Ultraconserved elements are associated with homeostatic control of splicing regulators by alternative splicing and nonsense-mediated decay

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Ultraconserved elements are associated with homeostatic control of splicing regulators by alternative splicing and nonsense-mediated decay


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Many alternative splicing events create RNAs with premature stop codons, suggesting that alternative splicing coupled with nonsense-mediated decay (AS-NMD) may regulate gene expression post-transcriptionally. We tested this idea in mice by blocking NMD and measuring changes in isoform representation using splicing-sensitive microarrays. We found a striking class of highly conserved stop codon-containing exons whose inclusion renders the transcript sensitive to NMD. A genomic search for additional examples identified >50 such exons in genes with a variety of functions. These exons are unusually frequent in genes that encode splicing activators and are unexpectedly enriched in the so-called “ultraconserved” elements in the mammalian lineage. Further analysis show that NMD of mRNAs for splicing activators such as SR proteins is triggered by splicing activation events, whereas NMD of the mRNAs for negatively acting hnRNP proteins is triggered by splicing repression, a polarity consistent with widespread homeostatic control of splicing regulator gene expression. We suggest that the extreme genomic conservation surrounding these regulatory splicing events within splicing factor genes demonstrates the evolutionary importance of maintaining tightly tuned homeostasis of RNA-binding protein levels in the vertebrate cell.

Keywords: SR proteins; splicing microarray; hnRNP proteins; splicing factor; autogenous regulation; epigenetics

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The nonsense-mediated decay (NMD) pathway is an mRNA surveillance mechanism that limits the translation of mRNAs with premature termination codons (PTCs) (Conti and Izaurralde 2005; Lejeune and Maquat 2005; Maquat 2005). In mammals, PTCs are partially determined by their position relative to the activity of the splicing machinery. The spliceosome leaves a protein mark called the exon–junction complex (EJC) at each junction of the spliced mRNA (Le Hir et al. 2000, 2001). During the first (pioneer) round of translation, the ribosome removes the EJC, and the mRNA is stabilized (Ishigaki et al. 2001; Chiu et al. 2004; Lejeune et al. 2004). If an EJC remains on the mRNA as the ribosome terminates, proteins within the EJC, notably the Upf3 or Upf3x and Upf2 proteins, recruit Upf1, which triggers NMD-mediated destruction of the mRNA (Lykke-Andersen et al. 2000, 2001; Kim et al. 2001; Wang et al. 2001; Schell et al. 2003; Kashima et al. 2006). This relationship between NMD and splicing has consequences for gene structure: The most 3’ exon–exon junction can be no >50–55 nucleotides (nt) downstream from the stop codon. This means that normal stop codons are nearly always found in the last exon, or within 55 nt of the end of the second-to-last exon of the gene (Nagy and Maquat 1998; Maquat 2005).

A second consequence of the relationship between NMD and splicing is that errors of splicing in which normal exons are skipped, or inappropriate intron segments are included will be destroyed by NMD if they generate a premature stop codon (Lejeune and Maquat 2005). Studies of human and mouse EST data predict that about one-fifth of the alternatively spliced human and mouse genes produce PTC-containing RNAs that should be subject to NMD (Lewis et al. 2003; Baek and Green 2005). How many of these are splicing errors is not clear.
Alternative splicing and NMD

(Sorek et al. 2004). Besides removing splicing errors, NMD may also play an important role in regulating the expression of many genes by coupling alternative splicing (AS) to decay [Lewis et al. 2003; Baek and Green 2005].

The regulatory role of coupling between AS and NMD is supported by several lines of experimental evidence. The expression profile in Upf1-depleted HeLa cells shows that ~5% of expressed transcripts are up-regulated [Mendell et al. 2004]. Splicing factors like polyadenylate-binding protein (PTB) and serine/arginine-rich 2 protein (SC35) are known to autoregulate their own expression by binding to their own mRNA to increase splicing of the NMD-sensitive isoform [Sorek et al. 2001; Wollerton et al. 2004]. In these cases the splicing regulation represses an exon, whose skipping creates a PTC by frame shifting [Wollerton et al. 2004], or activates the removal of cryptic introns in the 3′ untranslated region [UTR], in essence introducing an EJC downstream from the normal stop codon [Sorek et al. 2001].

Genes such as neural PTB (nPTB) [Rahman et al. 2002], TRA2-β [Stoilov et al. 2004], and SRp20 [Jumaa and Nielsen 1997], have been reported to create mRNA isoforms potentially encoding truncated proteins by alternative splicing, but their relationship to NMD has not been determined.

Recent work measuring PTC-containing RNAs by microarrays showed that a large proportion of them are expressed at low levels and are not sensitive to inhibition of NMD [Pan et al. 2006]. This result goes against the hypothesis that AS-NMD is a widespread mechanism of gene regulation in vertebrates [Lewis et al. 2003; Mendell et al. 2004; Baek and Green 2005], and raises more questions concerning which NMD-coupled splicing isoforms in the database represent regulated splicing events, and which represent splicing errors [Lewis et al. 2003; Sorek et al. 2004; Baek and Green 2005; Pan et al. 2006]. Since the levels of NMD-sensitive isoforms are kept low due to their efficient degradation, the database counts of ESTs for them is unlikely to reflect the frequency at which they are generated by splicing [Baek and Green 2005]. It has thus been a challenge to reveal functionally important NMD-coupled splicing events genome-wide and study their regulation mechanisms.

In this paper we use splicing-sensitive microarrays to identify a group of alternative RNA isoforms that accumulate after inhibition of NMD. We identify a biologically relevant class of NMD-sensitive isoforms that arises by inclusion of highly conserved exons that contain stop codons in all three reading frames. We used the properties of the array-identified exons to create a program to scan the genome for other examples and found many. These conserved stop codon exons are particularly common in genes encoding splicing activators known to activate exon inclusion, and frequently overlap ultraconserved elements in mammalian genomes [Bejerano et al. 2004]. Unlike previous examples of autogenous negative regulation of alternative splicing coupled to NMD (Sorek et al. 2001; Wollerton et al. 2004), the inclusion of these novel exons must be activated to block gene expression. We suggest that the exonic class of ultraconserved elements is critically required for homeostatic maintenance of splicing factor expression levels.

Results

Detection of AS-NMD using splicing-sensitive microarrays

Public databases of EST sequences hinted at the existence of coupling between alternative splicing and NMD. But discrimination of functionally important splicing from splicing errors in these ESTs has been a challenge [Sorek et al. 2004; Pan et al. 2006]. To experimentally detect biologically relevant examples of AS-NMD, we blocked NMD and used splicing-sensitive microarrays [Clark et al. 2002; Wang et al. 2003; Ule et al. 2005; Pan et al. 2006; Sugnet et al. 2006], seeking alternatively spliced RNAs whose levels increased compared with controls. To identify effects resulting from NMD inhibition, rather than secondary effect specific to a particular treatment, we used two different methods of NMD inhibition. In one method, we transfected mouse N2A cells with Upf1 small interfering RNA (siRNA) to down-regulate Upf1, a core component of the NMD machinery [Mendell et al. 2002]. Upf1 protein was reduced to at least 25% of its level in untreated cells or control siRNA-treated cells (Fig. 1A). In the second method, we treated N2A cells with the translation inhibitor emetine [Noensie and Dietz 2001], since translation is a prerequisite for NMD-mediated degradation. To test the effectiveness of these approaches, we measured accumulation of the alternative isoform resulting from skipping of

Figure 1. NMD pathway is blocked by Upf1 RNA interference (RNAi) and emetine treatment. (A) Western blot using antibodies against Upf1 and α-Tubulin. Lanes 1, 2, and 3 contain equal amounts of total cell protein 2 d after siRNA transfection. Lanes 4 and 5 contain 50% and 25% of lane 3. (B) RT–PCR of nPTB mRNA isoforms. nPTB primers are in constitutive exons 9 and 11 of nPTB mRNA. Exon 10 skipping will generate NMD-targeted nonsense isoforms. Nonsense isoforms are accumulated in Upf1 siRNA-transfected cells and emetine-treated cells. RT–PCR of β-actin mRNA was used as control.
exon 10 of neural polypyrimidine tract-binding protein (nPTB) mRNA (Rahman et al. 2002) by RT–PCR. This isoform is predicted to create a PTC, and the data shows that both treatments effectively blocked NMD (Fig. 1B). RNA from treated and untreated cells was converted to cDNA, labeled, and applied to Affymetrix alternative splicing microarrays as previously described (Wang et al. 2003; Ule et al. 2005; Sugnet et al. 2006). We analyzed the data using the method of Sugnet et al. (2006). Since the EST/mRNA sequences used for array design were not filtered by frequency [D. Kulp and M. Ares, unpubl.], many NMD-sensitive [and hence rarely present in EST libraries] RNAs can be measured by this array. Changes in levels of alternatively spliced isoforms were observed for all splicing modes [e.g., cassette exon, alternative 5' splice site, etc.] in each individual class of treatment [data not shown]. We focused on the 229 cassette exons for which isoform representation in the RNA pool appeared to change under either of the NMD-inhibiting treatments [Supplementary Table S1]. Using the University of California at Santa Cruz [UCSC] Genome Browser [Karolchik et al. 2003], we identified 28 cassette exons for which the predicted NMD isoform accumulated when NMD was blocked [Table 1]. We tested 25 of these by RT–PCR, and verified that 24 [96%] of them change as indicated by the array [Supplementary Table S1; see also Figs. 2, 3]. We conclude that both Upf1 siRNA knockdown and emetine treatment lead to many changes in isoform representation, of which a little more than 10% can directly be attributed to NMD. The nearly 90% of changes in isoform representation that do not conform to known rules of NMD could be due to perturbation of other translation and Upf1-dependent decay mechanisms (Kim et al. 2005) or could be secondary effects of blocking NMD.

A class of conserved alternative exons brings stop codons into mRNA to trigger NMD

Previous studies of splicing and NMD have suggested a relationship between conserved and nonconserved alternative splicing events and NMD (Baek and Green 2005; Pan et al. 2006). To distinguish which of the 28 NMD isoforms detected by microarray might be generated by

<table>
<thead>
<tr>
<th>NMD-inducing splicing event</th>
<th>Gene name</th>
<th>Description</th>
<th>Conservation score</th>
<th>RT–PCR tests</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exon-skipping-induced NMD</td>
<td><strong>NFYB</strong></td>
<td>Nuclear transcription factor Y subunit β</td>
<td>0.99</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>DLGH4</strong></td>
<td>Post-synaptic density protein 95</td>
<td>0.97</td>
<td>True</td>
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<tr>
<td></td>
<td><strong>MRPL13</strong></td>
<td>60S ribosomal protein L13 mitochondrial</td>
<td>0.98</td>
<td>True</td>
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<tr>
<td></td>
<td><strong>WBSCR22</strong></td>
<td>Putative methyltransferase HUSSY-03</td>
<td>0.91</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>CCAR1</strong></td>
<td>Death inducer with SAP domain DIS</td>
<td>0.99</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>AK035230</strong></td>
<td>Protein phosphatase 1 regulatory subunit 12A</td>
<td>0.99</td>
<td>ND</td>
</tr>
<tr>
<td></td>
<td><strong>CDC16</strong></td>
<td>Cell division cycle protein 16 homolog</td>
<td>0.99</td>
<td>NI</td>
</tr>
<tr>
<td></td>
<td><strong>CCT8</strong></td>
<td>T-complex protein 1, θ subunit</td>
<td>0.97</td>
<td>ND</td>
</tr>
<tr>
<td></td>
<td><strong>BRD8</strong></td>
<td>Bromodomain-containing protein 8</td>
<td>0.99</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>RPNI</strong></td>
<td>Oligosaccharide protein glycosyltransferase</td>
<td>0.97</td>
<td>ND</td>
</tr>
<tr>
<td></td>
<td><strong>HSFI</strong></td>
<td>Heat-shock factor protein 1</td>
<td>0.98</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>RPA1</strong></td>
<td>Replication protein A1</td>
<td>0.78</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>NIPSNAP1</strong></td>
<td>NipSnap1 protein</td>
<td>0.99</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>BRD2</strong></td>
<td>Female sterile homeotic-related protein Frg-1</td>
<td>0.93</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>FARSIB</strong></td>
<td>Phenylalanyl-tRNA synthetase β chain</td>
<td>No align</td>
<td>True</td>
</tr>
<tr>
<td>Exon-inclusion-induced NMD</td>
<td><strong>NOL5</strong></td>
<td>Nucleolar protein 5, NOP58 homolog</td>
<td>0.98</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>SFRS4</strong></td>
<td>Splicing factor, arginine/serine-rich 4</td>
<td>0.98</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>NKIRAS1</strong></td>
<td>KAPPAB-RAS1</td>
<td>No align</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>RNPC2</strong></td>
<td>RNA-binding domain protein 2 [CAPERα]</td>
<td>0.99</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>1300007B12RIK</strong></td>
<td>Hypothetical protein</td>
<td>0.99</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>ZCCHC6</strong></td>
<td>Ser/Arg-related nuclear matrix protein</td>
<td>0.99</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>DAERTD196E</strong></td>
<td>Hypothetical cytochrome c family protein</td>
<td>0.94</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>AK031574</strong></td>
<td>Hypothetical Ub-associated protein</td>
<td>0.92</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>TLK1</strong></td>
<td>Serine/threonine-protein kinase</td>
<td>0.99</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>SFRS9</strong></td>
<td>Splicing factor, arginine/serine-rich 9</td>
<td>0.98</td>
<td>True</td>
</tr>
<tr>
<td></td>
<td><strong>G430041M01RIK</strong></td>
<td>Tra-2 protein homolog [TRA-2 α]</td>
<td>0.99</td>
<td>True</td>
</tr>
</tbody>
</table>

All stop codon exons have stop codons in all three reading frames except those indicated by *, which have stops in two frames including the coding frame of the protein. [ND] Not determined; [NI] not interpretable.
functionally relevant splicing rather than by splicing errors, we grouped the 28 splicing events into four classes according to whether exon skipping or inclusion triggered NMD, and whether the exon was conserved in vertebrate genomes or not. All of the exons whose skipping triggered NMD were >78% conserved as indicated by their conservation score (Table 1). One known AS-NMD mechanism is the regulated skipping of a conserved protein coding exon to trigger NMD, which has been reported in the PTB gene (Wollerton et al. 2004). However, the skipping of conserved protein coding exons has been frequently observed, and can be interpreted as a splicing error [Sorek et al. 2004; Baek and Green 2005]. Unfortunately, in the exon-skipping NMD class of events it is difficult to separate exon conservation that could be required for splicing regulation from that required to code for protein. This creates doubt about whether regulated exon-skipping NMD or splicing error is the underlying cause of this alternative splicing. Some of the exon-skipping NMD exons in Table 1 are embedded in conserved intron sequences, similar to those in the exon-skipping NMD-regulated PTB (Wollerton et al. 2004) and nPTB genes [Fig. 1; Rahman et al. 2004]. This suggests that these conserved exons could be new examples of genes regulated by the coupling of alternative splicing and NMD, but additional experimentation is required to test this.

The situation is different for the exon-inclusion NMD class (Table 1). The majority of the exon-inclusion NMD events we found belong to an unexpected class of highly conserved, nonprotein coding exons containing in-frame stop codons (Table 1). A detailed example is shown for NOL5, a nucleolar protein homologous to yeast Nop58p, component of C/D-Box methylation guide snoRNPs [Fig. 2]. A plot of the skip/include ratios calculated from the plots of data from each array for the NMD inhibited (Fig. 2A, filled characters) and control groups (Fig. 2A, open figures) shows that the inclusion of the NOL5 exon increases when NMD is blocked (Fig. 2A). The log2 difference in the slopes is 1.1922, indicating a 2.3-fold increase in NMD-blocked cells relative to non-NMD-blocked cells for this exon [Sugnet et al. 2006]. (B) RT–PCR validation of accumulation of NOL5 stop codon exon after blocking NMD. The stop codon exon-inclusion isoform (non-sense isoform) is specifically accumulated upon Upf1 siRNA transfection or emetine treatment as compared with control cell samples. Percentage of exon inclusion increases fourfold by Upf1 siRNA transfection or ninefold by emetine treatment, as measured by an Agilent Bioanalyzer. (C) NOL5 stop codon exon as seen in the UCSC Genome Browser [Karolchik et al. 2003]. Base position shows three reading frames. In each reading frame, black squares represent stop codons and the white squares represent methionine codons, whereas the gray squares represent nonmethionine amino acid codons. The NOL5 exon has stop codons in all three reading frames. The conservation track at the bottom shows high conservation in exonic and flanking intronic sequences of the NOL5 stop codon exon.
mouse, rat, and dog, as indicated by the conservation probability calculated and displayed in the UCSC Genome Browser (Fig. 2C; Supplementary Table S2; Karolchik et al. 2003; Siepel et al. 2005). These stop codon exons often maintain high conservation well into flanking intronic regions (Fig. 2C; Supplementary Table S4), as is seen for exons whose alternative splicing is conserved (Sorek and Ast 2003; Sugnet et al. 2004, 2006; Yeo et al. 2005). The stop codon exons found in RNPC2 and TRA2A overlap the so-called “ultraconserved” elements in the vertebrate genome, previously defined to be the 481 regions of ≥200 base pair (bp) without variation in the human, mouse, and rat genomes (Bejerano et al. 2004). Unlike the exon-skipping NMD exons, conservation of the stop codon exons cannot be due to protein coding, and therefore strongly implies a regulatory function for these exons.

Within this group of exons we found two that are present only in the mouse and do not appear in other vertebrates (FARS1B and NKIRAS1). These could represent important species-specific splicing (Modrek and Lee 2003), but could also be recently exorized short interspersed repetitive elements (SINEs) (see Lev-Maor et al. 2003; Bejerano et al. 2006; also see below). There are many examples of such exons in the genome and both exons show homology mouse SINE elements (Supplementary Table S2). These two SINE-derived exons were presumably identified along with the conserved class because they are among the most highly expressed and actively decayed SINE-derived exons in the mouse N2A cells we tested.

A class of conserved stop codon exons in mammalian genomes

Our array experiment is limited to splicing events that are represented on our array and are expressed in mouse neuroblastoma cells at a level that meets our stringent criteria for detection and analysis [Sugnet et al. 2006]. To find additional conserved stop codon exons in mammalian genomes, we searched mouse and other mammalian EST and cDNA databases for internal (i.e., not 5’ or 3’ UTR) exons with ≥80% sequence conservation in mouse, human, rat, and dog genomes (Bejerano et al. 2004). Unlike the exon-skipping NMD exons, conservation of the stop codon exons cannot be due to protein coding, and therefore strongly implies a regulatory function for these exons.

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these alternative 5′ or 3′ splice sites would function identically to a cassette stop codon exon to trigger NMD.

We tested 30 of the 55 new stop codon exons found in the computational search by RT-PCR of N2A cell RNA, and 15 were detectably expressed in both the emetine-treated and untreated control N2A cells [Fig. 3]. Of these, 14 (93%) clearly show accumulation of nonsense isoforms upon blocking NMD with emetine, and one showed no change [Fig. 3]. We conclude that the majority of conserved stop codon exons are actively included in pre-mRNA by alternative splicing, and destabilize the RNA isoforms that contain them through NMD [Fig. 3]. From the conservation of these exons and the consequence of their inclusion for expression of the genes that carry them, we infer that stop codon exons are an important negative regulatory element for post-transcriptional gene regulation. We define a stop codon exon as a conserved, alternatively spliced, noncoding exon internal to a transcript that triggers NMD.

Stop codon exons are enriched in RNA-splicing factor genes and ultraconserved elements

To ask whether conserved stop codon exons are found more often in certain functional gene classes than others, we analyzed the functional annotations associated with the 65 stop codon-containing genes in the mouse genome using FuncAssociate (Berriz et al. 2003). Of these, 11 genes function in RNA splicing [GO Biological Process GO: 0,008,380] [Supplementary Table S3]. The likelihood that the association of stop codon exons with splicing factors is due to chance is small [p value = 3.60e-10] [Supplementary Table S3].

Because of the unexpectedly high conservation of stop codon exons and their association with RNA-splicing proteins, we examined the distribution of ultraconserved element among stop codon exons. There are ~136,000 exons in the UCSC Genome Browser Known Genes track (Karolchik et al. 2003), and 111 exonic ultraconserved elements [Bejerano et al. 2004]. Of the 66 conserved stop codon exons (in 65 different genes) we identified in the mouse, the human orthologs of nine overlap or are entirely contained within ultraconserved elements. This is an association statistically unlikely to be due to chance [p value = 4.1e-18, Fisher’s exact test]. Of the nine ultraconserved elements that overlap stop codon exons, eight are found in seven different RNA-splicing-associated genes (RNPC2, TRA2A, SFRS3, SFRS6, SFRS7, SFRS10, and TIAL1). Previously, Bejerano et al. (2004) demonstrated that the “exonic” class of ultraconserved elements is associated with RNA-splicing and -binding protein genes. Combining this finding with our data showing that stop codon exons are strongly enriched in splicing factor genes and frequently contained within ultraconserved elements, we conclude that the exonic class of ultraconserved elements contains members essential for the proper regulation of splicing factor genes by AS-NMD (Table 2).

Exonic ultraconserved elements within splicing factor genes generally link alternative splicing to NMD with a polarity consistent with autogenous regulation

Given the link between stop codon exons, ultraconserved elements, and exon-inclusion NMD, we searched all the human exonic ultraconserved elements in RNA-binding protein genes [Bejerano et al. 2004] using the UCSC Genome Browser [Karolchik et al. 2003] for more instances of ultraconserved elements connecting alternative splicing to NMD. Of the 29 exonic ultraconserved elements in RNA-binding protein genes in human [Bejerano et al. 2004], 15 have human and/or mouse EST evidence suggesting the presence of AS-NMD in those regions. Of the 15, eight are stop codon exons, and the other seven appear to link alternative splicing to NMD regulation via other kinds of splicing events such as exon skipping or activation of a 3′ UTR intron (Table 2). One example is the ultraconserved element of the splicing

Table 2. Human ultraconserved elements in splicing factor genes host-splicing-NMD regulation

<table>
<thead>
<tr>
<th>Splicing regulator</th>
<th>Ultra element</th>
<th>Gene</th>
<th>Protein</th>
<th>Splicing-NMD event</th>
</tr>
</thead>
<tbody>
<tr>
<td>SR proteins and splicing activators</td>
<td>uc.28</td>
<td>SFRS11</td>
<td>Splicing factor p54</td>
<td>Stop codon exon</td>
</tr>
<tr>
<td></td>
<td>uc.50</td>
<td>SFRS7</td>
<td>Splicing factor 9G8</td>
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<td></td>
<td>uc.138</td>
<td>SFRS10</td>
<td>Tra2-β [Stoilov et al. 2004]</td>
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<td>uc.189</td>
<td>SFRS3</td>
<td>SRp20 [Jumaa and Nielsen 1997]</td>
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</tr>
<tr>
<td></td>
<td>uc.208, 209</td>
<td>TRA2A</td>
<td>Tra2-α</td>
<td>Stop codon exon</td>
</tr>
<tr>
<td></td>
<td>uc.418</td>
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<td>SF2/ASF, SRp30a</td>
<td>3′ UTR intron activation</td>
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<td>uc.456</td>
<td>SFRS6</td>
<td>SRp55</td>
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<td>RNPC2</td>
<td>CAPERα</td>
<td>Stop codon exon</td>
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<td>uc.313</td>
<td>TIAL1</td>
<td>TIAR [Le Guiner et al. 2001]</td>
<td>Stop codon exon</td>
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<td>PTBP2</td>
<td>Neural hnRNP I, nPTB (Rahman et al. 2004)</td>
<td>Exon-skipping frameshift</td>
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<td>uc.144</td>
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<tr>
<td></td>
<td>uc.263</td>
<td>HNRPK</td>
<td>hnRNP K</td>
<td>Exon-skipping frameshift</td>
</tr>
<tr>
<td></td>
<td>uc.443</td>
<td>HNRPM</td>
<td>hnRNP M</td>
<td>Exon-skipping frameshift</td>
</tr>
<tr>
<td></td>
<td>uc.151</td>
<td>ZFR</td>
<td>Zinc-finger RNA-binding protein</td>
<td>Exon-skipping frameshift</td>
</tr>
</tbody>
</table>

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repressor gene PTBP2 [nPTB] (Bejerano et al. 2004; Rah-ram et al. 2004), which contains a 54-nt exon whose skipping would be predicted to cause a frame shift and NMD (Fig. 1B; Table 2). In another example, the ultra-conserved element uc.418 in SFRS1, encoding the splicing activator ASF/SF2, overlaps an alternative intron in the 3′ UTR whose removal causes NMD by placing an EJC downstream from the normal stop codon (Table 2; Wang et al. 1996; Sureau et al. 2001).

A striking aspect of the findings in Table 2 is that for splicing factors known mostly to be activators, such as SR proteins, the splicing event that triggers NMD within its pre-mRNA is a splicing activation event, whereas for factors known mostly to be repressors the splicing event that triggers NMD is a splicing-repression event. This polarity is identical to that expected for homeostatic auto- or cross-regulatory maintenance of proper splicing factor levels in the cell. Thus, in general, NMD is triggered for splicing activator mRNAs by activating a splicing event [exon inclusion or intron activation], and for splicing repressor mRNAs by repressing a splicing event [exon skipping], in order to bring levels of splicing factor back toward a set point [Fig. 4; see Discussion]. We suggest this polarity is the key to maintaining an appropriate balance between positively acting and negatively acting general splicing factors to ensure proper levels of basal exon inclusion.

Retrotransposons, stop codon exons, and ultraconserved elements associated with AS-NMD

Retrotransposons that land in introns can become “exonized” or spliced into mRNA of the gene into which they have inserted, often introducing stop codons into mRNA (Lev-Maor et al. 2003; Mendell et al. 2004; Bejerano et al. 2006). Recent transposition events will appear nonconserved; however, ancient retroposons that became exonized and subject to purifying selection early in the vertebrate lineage would be conserved in extant vertebrates and could achieve ultraconserved status despite their humble beginnings (Bejerano et al. 2004, 2006). The clearest example is the LF-SINE still active in the Coelacanth genome, which gave rise to two distinct ultraconserved elements, one of which overlaps an ultraconserved coding exon of the PCBP2 gene (Bejerano et al. 2006). To investigate whether retrotransposon exonization generally plays a role in creating stop codon exons for exon-inclusion NMD regulation during evolution, we used the UCSC Genome Browser repeating elements track (Jurka 2000; Karolchik et al. 2003) and found that four of our 66 conserved stop codon exons clearly overlap members of a SINE family that predates our last common ancestor with the dog [MIR/MIRb family] (Supplementary Table S2, Smit 1999). Three others appear derived from long interspersed repetitive elements (LINEs) and long terminal repeat elements (LTR). None of the retrotransposon-derived stop codon exons overlap with the ultraconserved class. Two detected by arrays [NKIRAS1 and FARSLB] are derived from more recent SINE element insertions found only in the mouse (Table 1; Supplementary Table S2). We conclude that retrotransposons contribute to, but do not entirely explain the origin of stop codon exons in the vertebrate genome (Supplementary Table S2).

Discussion

In this study we have used splicing-sensitive microarrays to identify a class of alternative splicing events that actively produce NMD-sensitive isoforms. A striking class of highly conserved stop codon exons comprises a large fraction of the events we found (Table 1; Fig. 3). Using the features of these experimentally defined exons, we searched genomic data, finding and then validating more examples (Fig. 3; Supplementary Table S2). The abundance of alternative splicing events that give rise to NMD isoforms (Lewis et al. 2003; Back and Green 2005) prompted the suggestion that coupled alternative splic-
Ultraconserved Regions that control AS-NMD in splicing factor genes are ultraconserved

Ultraconserved elements are genomic regions that are longer than 200 bp and identical in sequence in the human, mouse, and rat genomes [Bejerano et al. 2004]. In our data, eight stop codon exon-containing genes (RNPC2, TRA2A, SFRS3, SFRS6, SFRS7, SFRS10, TIAL1, and 2900045N06Rik) overlap nine ultraconserved elements, and are statistically overrepresented in the 65 stop codon exon-containing genes relative to all genes. We also showed that more than half (15 of 29) of exonic ultraconserved elements found in RNA-splicing and RNA-binding protein genes appear to be associated with AS-NMD regulation (Table 2). Besides stop codon exon-inclusion NMD, many (eight of 15) of these ultraconserved exons are subject to other modes of AS-NMD regulation, such as exon-skipping NMD (Table 2, Fig. 4). In addition, even those stop codon exons whose conservation is extremely high but falls just short of ultraconserved may have exceedingly important or constrained functions. Together, these observations argue that the unusual evolutionary conservation of the special regions where alternative splicing leads to NMD in certain genes such as those encoding splicing factors could be due to a strong functional demand for combined AS-NMD auto-regulation and cross-regulation.

A model for homeostasis of splicing factor gene expression to maintain proper basal levels of exon recognition

In general, SR proteins are considered splicing activators that promote exon recognition through exonic splicing enhancers [ESEs], whereas hnRNP proteins act antagonistically to SR proteins, providing a regulatory counter-weight through exonic or intronic splicing silencers [ESSs or ISSs] [Black 2003; Matlin et al. 2005]. The ESE and ESS/ISS sequences that bind splicing factors are remarkably short and degenerate, and present in bewildering combinations, even in ordinary exons [Fairbrother et al. 2002; Black 2003; Wang et al. 2004; Matlin et al. 2005]. Although this generality has its exceptions, the expression of two sets of opposing splicing regulators with potential to influence the recognition of every one of ∼136,000 exons in the vertebrate cell must reflect a grand compromise. Such a compromise is also the ground state on which alternative splicing regulation must operate, since a regulated exon must be neither too strongly repressed nor too strongly activated in this ground state; otherwise, it may not be subject to the regulatory influence of specialized splicing factors. It therefore follows that the levels of the two sets of opposing splicing regulators must themselves be stably, as well as jointly, regulated.

We found that the polarity (activation or repression) of the splicing event that triggers NMD in the opposing groups of splicing factor genes has a strong positive correlation with the polarity of each group’s action in splicing (Table 2). This creates a general negative feedback on the mRNA levels of the splicing factors in accordance with their activity and contribution to the basal state [Fig. 4]. If positive regulators are in general too high, they act to lower the level of splicing activator mRNA by exon-inclusion NMD, or intron-activation NMD [Table
As the level of negatively acting splicing factors becomes too high, they act to lower the levels of splicing repressor mRNAs by exon-skipping NMD (Table 2; Fig. 4; Wollerton et al. 2004). We envision this stable balance in the activities of splicing activator and repressor proteins to be a self-perpetuating, stable state of gene expression, in essence, an RNA-based epigenetic state. Although there are many other mechanisms by which individual splicing factor activities are regulated (e.g., see Huang and Steitz 2005), we propose that the general demand for regulating a stable balance between antagonistic splicing factor activities at the mRNA level explains the ultraconserved nature of the genomic sequences that host AS-NMD.

The idea that autogenous control by RNA-binding proteins can maintain a stably inherited state of gene expression has precedence in studies of the Drosophila splicing regulator Sxl. Once activated early in development, Sxl maintains an epigenetic female state by activating its own splicing in a positive feedback loop (Bell et al. 1991). In the case of opposing splicing regulators in vertebrates, the feedback we illustrate in Figure 4 is primarily negative, but it seems likely there will be cross-regulatory influences that add layers to this simple model, and that these will be focused on the alternative splicing events that trigger NMD. For example, the repression of the stop codon exon of a splicing activator mRNA could occur if splicing repressor proteins become too high, and this would boost the levels of activators to counteract the increased level of repressors. Conversely, increasing levels of splicing activators could increase recognition of the key regulatory exon in splicing repressor mRNAs that would lead to increases in splicing repressor mRNA. There are many examples of alternative splicing regulation of splicing factor mRNAs by other splicing factors. For example, SRp30c (SFRS9) controls alternative splicing of hnRNP A1 (Simard and Chabot 2002), ASF/SF2 (SFRS1) represses the SRp20 (SFRS3) stop codon exon and antagonizes SRp20 autoregulation (Jumaa and Nielsen 1997). It will be important to understand the regulatory influences of other splicing factors on the regions where AS-NMD is taking place in order to understand the influences that maintain and regulate the activities of the many splicing regulators.

Materials and methods

Cell culture, transfection, and drug treatment

Dr. Douglas L. Black (University of California at Los Angeles, Los Angeles, CA) generously provided mouse N2A cells. The cells were maintained in Dulbecco’s modified Eagle’s medium (DMEM) with 10% fetal bovine serum (FBS). For transfection, siRNA or minigenes were transfected into the cells using Lipofectamine 2000 reagent (Invitrogen) according to the manufacturer’s instructions. Sequences of Upf1 and negative control siRNA were 5’-GAUCCGAUCCGUUCAUCCGdTdT-3’ and 5’-UAUUGUCGACAUCCGUGCCTdTdT-3’, respectively. For emetine treatment, cells were exposed to 100 µg/mL emetine dihydrochloride hydrate (Fluka) 10 h before harvesting.

Western blotting and RT–PCR

Proteins from extracts of treated cells were separated in SDS-polyacrylamide gels, transferred to nitrocellulose, and probed with polyclonal goat anti-REN1 (Upf1) antibody (Bethyl) or monoclonal anti-a-Tubulin as loading control. Bound antibodies were detected with ECL-plus Western Blotting Detection System (Amersham) as instructed by the manufacturer.

Total RNA was extracted using Trizol reagent (Invitrogen). cDNA was generated from 1 µg of total RNA using SuperScript II Reverse Transcriptase (Invitrogen) using oligo-dT primer, according to the manufacturer’s instructions. For PCR, ~50 ng of cDNA was used as a template with primer pairs designed to measure splicing of the target exons (available on request). Reactions used Taq polymerase (Promega) and were run for 25–35 cycles at annealing temperatures appropriate for the primer pairs used. PCR products were checked using agarose gels stained with ethidium bromide, and were quantitated on an Agilent Bioanalyzer running the DNA 1000 chip.

Mouse splicing microarray

The Affymetrix mouse splicing microarray is described in Sugnet et al. (2006). Because EST/mRNA sequences used for array design were not filtered by frequency, many rare NMD-sensitive isoforms can be measured by this array. Three individual total RNA samples from Upf1 siRNA-transfected cells and control siRNA-transfected cells, and two individual total RNA samples each from emetine-treated cells and control cells were reverse transcribed, fragmented, end-labeled with biotin, and hybridized to arrays as described (Sugnet et al. 2006). Arrays were then stained and scanned, normalized, and analyzed as described (Sugnet et al. 2006). We analyzed the resultant microarray data using the method of Sugnet et al. (2006), reanalyzing the data three times, each time grouping the samples in one of three ways: Emetine versus Control, Upf1 siRNA versus Control siRNA, and Upf1 siRNA plus Emetine versus All controls. To simplify the identification of splicing isoforms that might be NMD substrates, we focused our analysis on the cassette exon mode of alternative splicing. Microarray data is deposited in the Gene Expression Omnibus (GEO) under accession number GSE6611 at http://www.ncbi.nlm.nih.gov/geo.

Calculation of conservation score

The multiple alignment and conservation score of each alternatively spliced exon comes from data in the UCSC Genome Browser conservation track (Karolchik et al. 2003). This track is made using the phastCons program (Siepel et al. 2005), which is based on a phylo-HMM, a type of probabilistic model that describes both the process of DNA substitution at each site in a genome and the way this process changes from one site to the next. The score is a probability that the alignment in question was derived by evolutionary conservation.

Bioinformatic identification of stop codon exon

Genome-wide identification of alternatively spliced exons relied on mouse ExonWalk data (Sugnet 2005). The ExonWalk method (Karolchik et al. 2003; Sugnet 2005) merges cDNA evidence together to predict full-length isoforms, including alternative transcripts. To predict transcripts that are biologically functional rather than the result of technical or biological noise, ExonWalk requires that every intron and exon either (1) be present in cDNA libraries of another organism (and also present in mouse), (2) have three separate cDNA GenBank (Benson et al. 2002) instructions. Sequences of Upf1 and negative control siRNA were 5’-GAUCCGAUCCGUUCAUCCGdTdT-3’ and 5’-UAUUGUCGACAUCCGUGCCTdTdT-3’, respectively. For emetine treatment, cells were exposed to 100 µg/mL emetine dihydrochloride hydrate (Fluka) 10 h before harvesting.
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