

# Ligand docking and virtual screening

CS/BioE/CME/Biophys/BMI 279

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# Outline

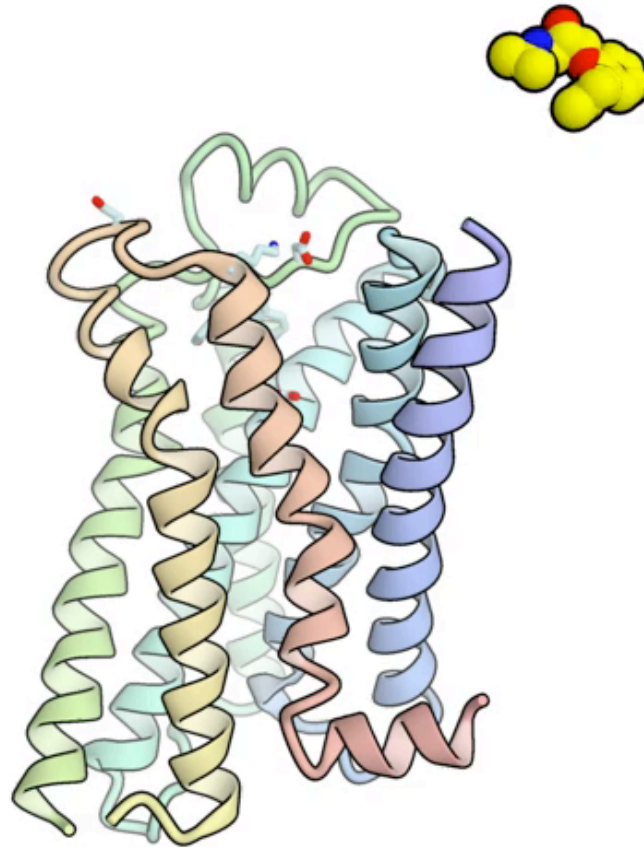
- Goals of ligand docking
- Defining binding *affinity* (strength)
- Computing binding affinity: Simplifying the problem
- Standard ligand docking methodology
- Virtual screening
- Alternative methods and current research directions

# Goals of ligand docking

# A drug binding to its target

(The majority of drug targets are proteins)

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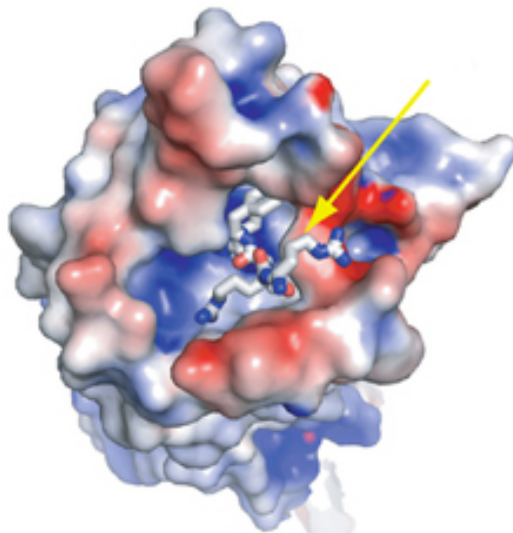


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Beta-blocker alprenolol binding to an adrenaline receptor

# Problem definition

- A *ligand* is any molecule that binds to a target macromolecule (e.g., a protein or RNA drug target)
  - We'll also use *ligand* to refer to any molecule (e.g., any candidate drug) that *might* bind to a given macromolecule
- *Ligand docking* addresses two problems:
  - Given a ligand known to bind a particular protein, what is its binding *pose* (that is, the location, orientation, and internal conformation of the bound ligand—basically, the position of each ligand atom when bound)
  - How *tightly* does a ligand bind a given protein (or other macromolecule)?



[http://www.nih.gov/researchmatters/october2012/images/structure\\_l.jpg](http://www.nih.gov/researchmatters/october2012/images/structure_l.jpg)

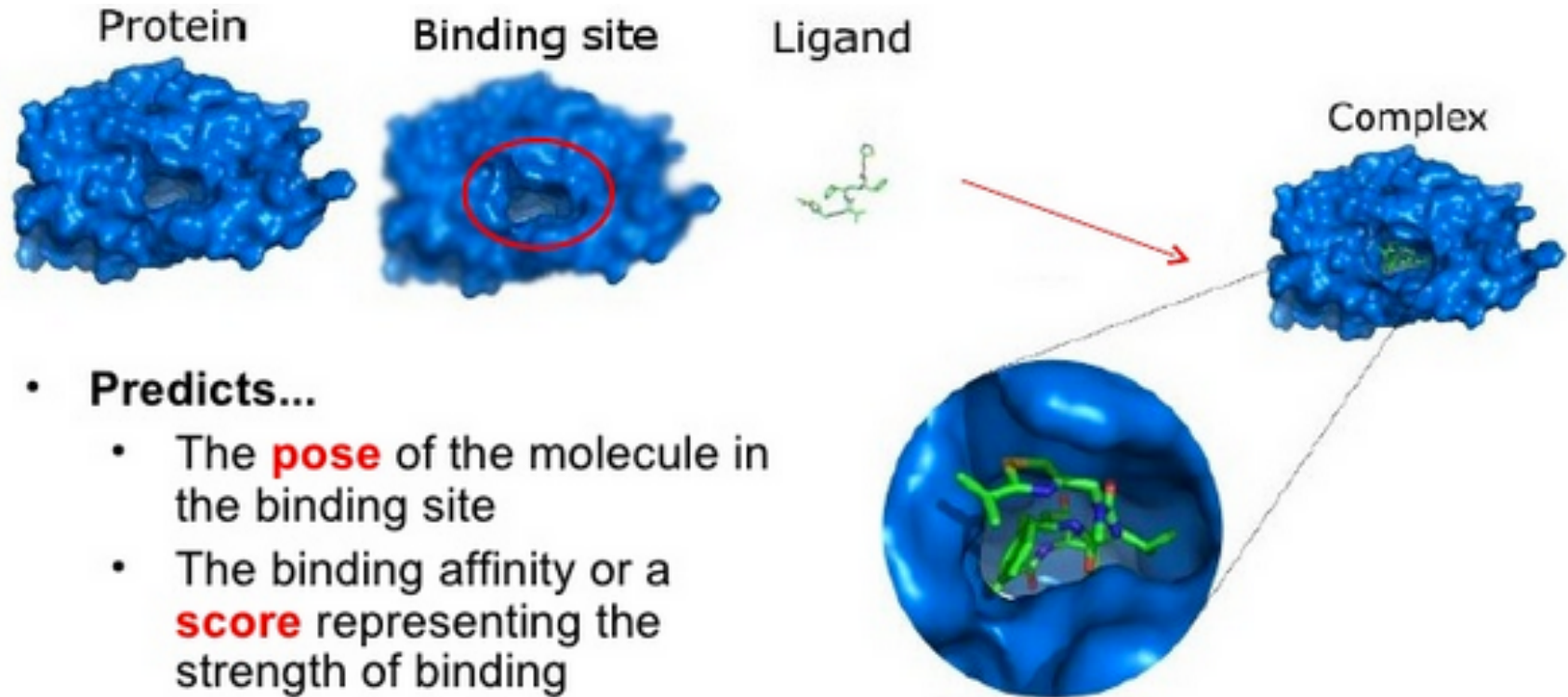
# Why is docking useful in drug discovery?

(area that docking is most widely used in)

- *Virtual screening*: Identifying drug candidates by considering large numbers of possible ligands
- *Ligand optimization*: Modifying a drug candidate to improve its properties e.g. solubility, binding selectivity, rate of metabolism in body
  - Docking can predict the candidate molecule's binding pose, which helps envision how modifying that molecule would change its binding strength and/or alter its effect on the target protein
  - Docking can predict binding strengths of related candidate molecules

Traditional method: carefully select compounds from library of existing compounds and perform experiments (time consuming + costly, limited to existing/known compounds)

# Ligand docking: a graphical summary



Defining binding *affinity* (strength)

# How do we specify how tightly a ligand binds to a protein?

- ***Binding affinity*** quantifies the binding strength of a ligand to a protein (or other target)
  - Conceptual definition: if we mix the protein and the ligand (with no other ligands around), what fraction of the time will the protein have a ligand bound?
    - This depends on ligand concentration, so we assume that the ligand is present at some standard concentration. Equivalently, given many protein copies, what fraction of them are bound at a specific instance?

Even tightly-binding ligands unbind at some point  
– there will always be an equilibrium between the bound and unbound states

# Binding affinity can be expressed in two ways

- A dissociation constant ( $K_D$ ), which is (roughly) the ligand concentration at which half the protein molecules will have a ligand bound **lower concentration = stronger binding (need fewer ligand molecules)**
  - For example, a “1 nanomolar (1 nM) binder” is a ligand that will occupy the binding site half the time at a concentration of 1 nM (i.e.,  $10^{-9}$  moles of ligand per liter of solute, typically water)
  - This is the most common way to express affinity
- The difference  $\Delta G$  in free energy of the bound state (all atomic arrangements where the protein has a ligand bound) and the unbound state (all other atomic arrangements)  **$\Delta G < 0$  for any binding ligands**
  - Typical units are kcal/mol or kJ/mol
  - Again, assume standard concentration of ligand
  - From  $\Delta G$ , one can compute the fraction of time the ligand will be bound

# Binding affinity: Clarifications

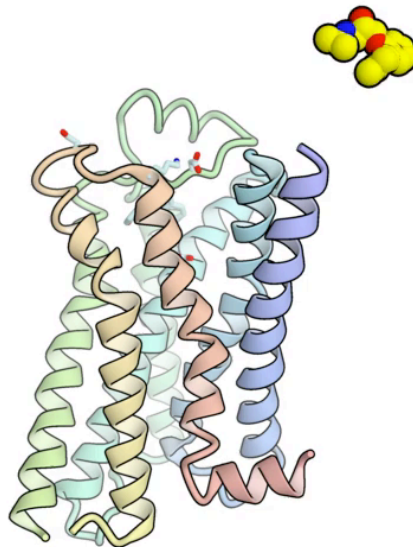
- Binding affinity is different from “how long the ligand remains bound” (the off-rate) or “how quickly the ligand binds” (the on-rate)
  - Binding affinity is a ratio of the on-rate and off-rate; you can’t calculate it from either one alone
  - These rates are also of interest in drug discovery, and predicting them is a different (and even more challenging) computational problem
- Binding affinity is different from “how strong are the inter-atomic forces between the ligand and the target when the ligand is bound”
  - Binding affinity also depends a great deal on what happens when the ligand isn’t bound (e.g., how favorable are the interactions of the ligand and the binding pocket with water)

# Computing binding affinity: Simplifying the problem

# A hypothetical direct approach to computing binding affinity

- Run a really long molecular dynamics (MD) simulation in which a ligand binds to *and* unbinds from a protein many times.
- Directly observe the fraction of time the ligand is bound.

0.00 us



# This direct approach rarely works

- It is so computationally intensive that one usually cannot do it for even a single ligand, let alone millions
  - The toughest part is the unbinding (dissociation)
    - Drug molecules usually take seconds to hours to unbind from their targets.
    - Microsecond-timescale molecular dynamics simulations usually take days.
  - We'd have to simulate *many* cycles of binding and unbinding.
- It is also limited by force field accuracy
  - Most molecular mechanics force fields are less accurate for small-molecule ligands than for proteins

# Question to discuss

- How would you compute a binding affinity?
  - Suppose you're given the structure of a target protein, and you want to compute the affinity of a particular ligand to that protein
  - To simplify the problem a bit, you may also assume that you're given the binding pose

# Standard ligand docking

(most common method to predict ligand binding affinity)

- Ligand docking is a fast, heuristic approach with two key components
  - A *scoring function* that very roughly approximates the binding affinity of a ligand to a protein given a binding pose
  - A *search method* that searches for the best-scoring binding pose for a given ligand

# Standard ligand docking

(most common method to predict ligand binding affinity)

- To predict the binding affinity of a ligand:
  - Docking software searches through poses of the ligand to find the pose with the best score
  - That pose is the predicted pose of the ligand, and its score is the predicted affinity
    - Here affinity is expressed as a binding energy:  
the lower the score, the more tightly the ligand binds

# Standard ligand docking

(most common method to predict ligand binding affinity)

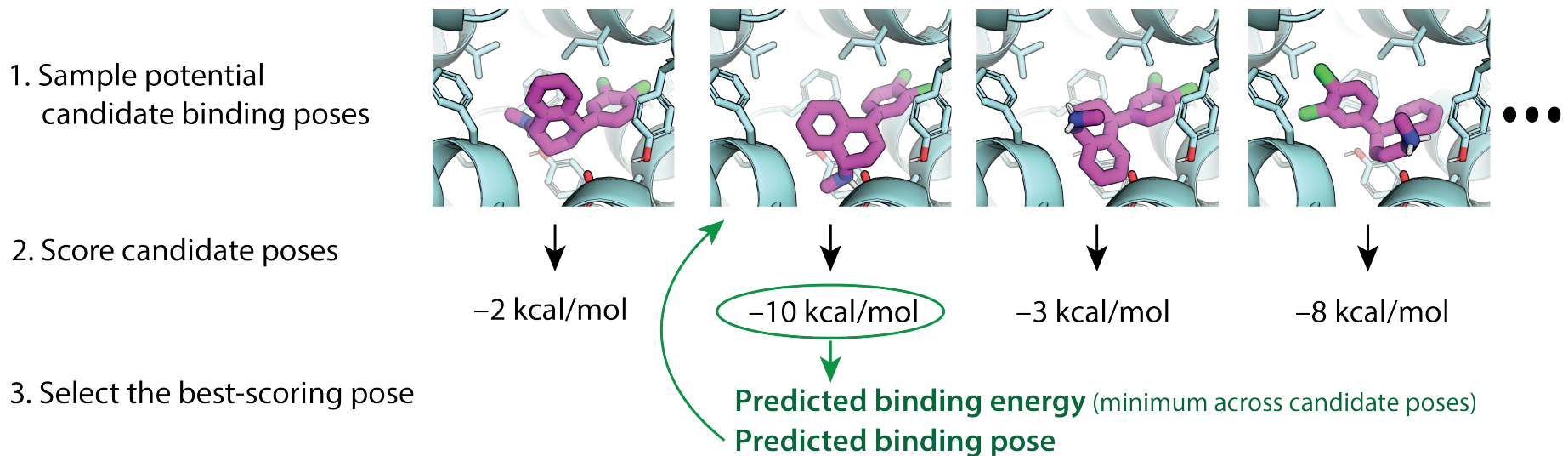


Figure by Ayush Pandit and Joe Paggi  
Paggi et al., Annual Review of  
Biochemistry, 93:389-410, 2024

Note that for docking to run reasonably quickly, one needs a good search strategy for the sampling step. Some search strategies iterate between generating candidate poses and scoring them.

# Ligand docking is approximate!

- For example, most ligand docking methods assume that the target protein is rigid and don't explicitly consider water molecules
- In reality, protein mobility, ligand mobility, and water molecules all play a major role in determining binding affinity
  - Docking is approximate but useful
  - The term *scoring function* is used instead of *energy function* to emphasize the highly approximate nature of the scoring function

# Docking software (a partial list)

Program ↕	Country of Origin ↕	Year Published ↕
AADS	India	2011
ADAM	Japan	1994
AutoDock	USA	1990
AutoDock Vina	USA	2010
BetaDock	South Korea	2011
DARWIN	USA	2000
DIVALI	USA	1995
DOCK	USA	1988
DockVision	Canada	1992
EADock	Switzerland	2007
eHiTS	UK	2006
EUDOC	USA	2001
FDS	UK	2003
FlexE	Germany	2001
FlexX	Germany	1996
FLIPDock	USA	2007
FLOG	USA	1994
FRED	UK	2003
FTDOCK	UK	1997
GEMDOCK	Taiwan	2004
Glide	USA	2004
GOLD	UK	1995
Hammerhead	USA	1996
ICM-Dock	USA	1997

Lead finder	Canada	2008
LigandFit	USA	2003
LigDockCSA	South Korea	2011
LIGIN	Germany	1996
LUDI	Germany	1992
MADAMM	Portugal	2009
MCDOCK	USA	1999
MDock	USA	2007
MolDock	Denmark	2006
MS-DOCK	France	2008
ParDOCK	India	2007
PhDOCK	USA	2003
PLANTS	Germany	2006
PRO_LEADS	UK	1998
PRODOCK	USA	1999
ProPose	Germany	2004
PSI-DOCK	China	2006
PSO@AUTODOCK	Germany	2007
PythDock	South Korea	2011
Q-Dock	USA	2008
QXP	USA	1997
rDock	UK	2013
SANDOCK	UK	1998
SFDOCK	China	1999
SODOCK	Taiwan	2007
SOFTDocking	USA	1991
Surflex	USA	2003
SYSDOC	USA	1994
VoteDock	Poland	2011
YUCCA	USA	2005

Most popular  
(based on citations  
2001–2011):

AutoDock  
GOLD  
DOCK  
FlexX  
Glide  
FTDOCK  
QXP

Sousa et al., Current  
Medicinal Chemistry  
2013

[http://en.wikipedia.org/wiki/  
Docking\\_\(molecular\)](http://en.wikipedia.org/wiki/Docking_(molecular))

# Standard ligand docking methodology

# Scoring functions

- Scoring functions used for docking typically capture chemists' intuition about what makes a ligand–target interaction energetically favorable. For example:
  - Hydrogen bonding
  - Hydrophobic interactions
- Parameters are fit based on known ligand–target structures and affinities
- These scoring functions are (very rough) attempts to approximate the binding *free energy*
  - By contrast, molecular mechanics force fields give *potential energy* associated with a particular arrangement of atoms

# Example: Glide scoring function

- Glide (widely used commercial docking software) uses the following “GlideScore” function:

$$\begin{aligned} \Delta G_{\text{bind}} = & C_{\text{lipo-lipo}} \sum f(r_{\text{lr}}) + \\ & C_{\text{hbond-neut-neut}} \sum g(\Delta r) h(\Delta \alpha) + \\ & C_{\text{hbond-neut-charged}} \sum g(\Delta r) h(\Delta \alpha) + \\ & C_{\text{hbond-charged-charged}} \sum g(\Delta r) h(\Delta \alpha) + \\ & C_{\text{max-metal-ion}} \sum f(r_{\text{lm}}) + C_{\text{rotb}} H_{\text{rotb}} + \\ & C_{\text{polar-phob}} V_{\text{polar-phob}} + C_{\text{coul}} E_{\text{coul}} + \\ & C_{\text{vdW}} E_{\text{vdW}} + \text{solvation terms} \end{aligned}$$

Friesner et al., Journal of  
Medicinal Chemistry  
47:1739-49 (2004)

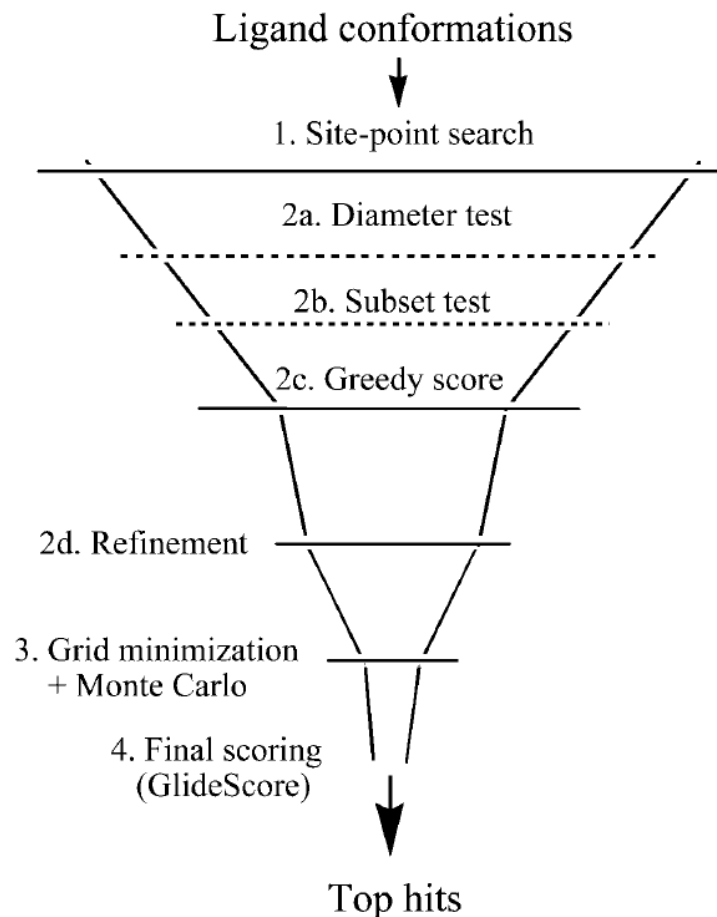
- The first term rewards contacts between hydrophobic atoms of the ligand and protein, and is a function of the distance between them
- The next three terms reward specific kinds of hydrogen bonds, and are functions of both distance and angle for each hydrogen bond
- Glide uses many additional terms as well

# Search methods

- Docking software searches for the best-scoring pose for each ligand
- The search space is huge, because one needs to consider all combinations of ligand position, ligand orientation, and ligand conformation (shape)
- To search this space efficiently, docking software typically employs either or both:
  - Hierarchical methods in which one uses approximate measures to identify promising *groups* of poses, then evaluates subgroups in more detail
  - Monte Carlo methods

# Example: Glide search

- Glide uses a hierarchical search method
- It first identifies a set of “reasonable” conformations for each ligand, by varying internal torsion angles
- For each ligand, it scans possible positions and orientations, using a rough measure of fit to binding pocket
- The most promising approximate poses undergo further “refinement”
- Candidate poses are ranked by the scoring function



Friesner et al., *J Med Chem* 47:1739, 2004

# Virtual screening

# Virtual screening: the basics

- Goal: identify ligands that bind to a target—particularly ligands that are very different from any known binder
- Typical process
  - Select a virtual library of chemical compounds
  - Use docking to estimate the affinity of each
  - Buy or make the compounds with the best predicted affinities and do experiments to test how well they bind
  - Optional: Optimization of experimentally validated binders by testing related chemical compounds

# Virtual screening: the basics

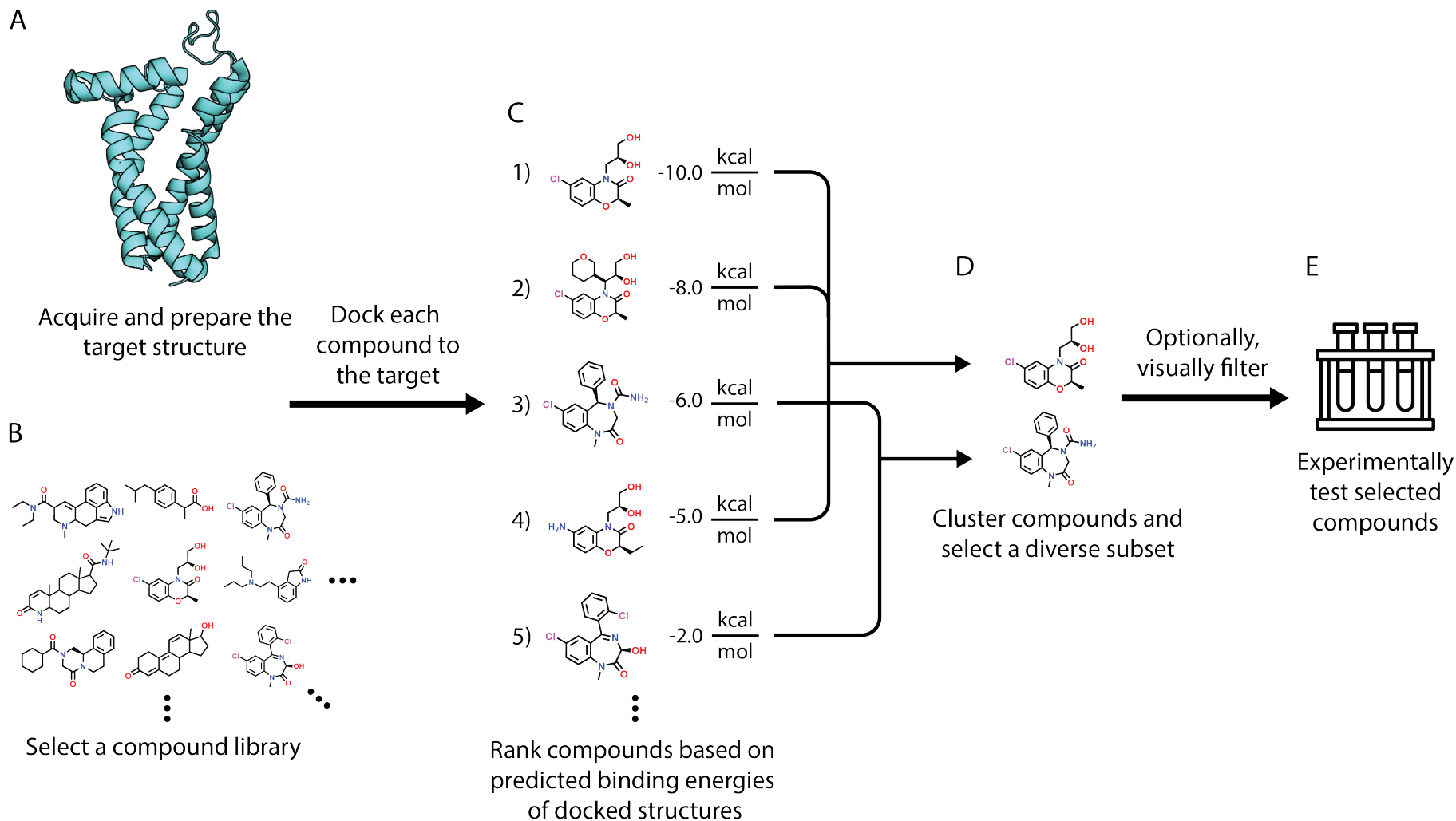


Figure by Ayush Pandit and Joe Paggi  
Paggi et al., Annual Review of  
Biochemistry, 93:389-410, 2024

# New: “Ultra-large” virtual libraries

- In virtual screening, one typically uses libraries of compounds that can be easily ordered from vendors, so that one can easily test the top-ranked ones
- A few years ago, a few million compounds were available from vendors
- Now it's billions or trillions
  - Thanks to the advent of the make-on-demand approach (pioneered by Enamine in Ukraine)
  - Idea: gigantic library of compounds that have not yet been made but that vendor can make quickly and cheaply with high probability
- This has increased the utility of virtual screening
  - A few million compounds can be tested experimentally by “high-throughput screening” robots, but this doesn't scale to billions and requires that all the compounds be synthesized in advance



# How well does docking work?

depends on what you want to do!

- The best standard docking protocols:
  - Predict a reasonably accurate pose (for ligands that do in fact bind the target protein) about half the time
    - Most of the time, some highly-ranked poses is reasonably accurate, but it may not be ranked first
  - Are not very useful when it comes to estimating affinities
  - Provide very useful (though far from perfect) results for virtual screening
    - Many recent, experimentally verified success cases
    - Virtual screening is different from affinity estimation because, in virtual screening, missing some actual binders in the library is fine as long as one can identify other binders with reasonable confidence.

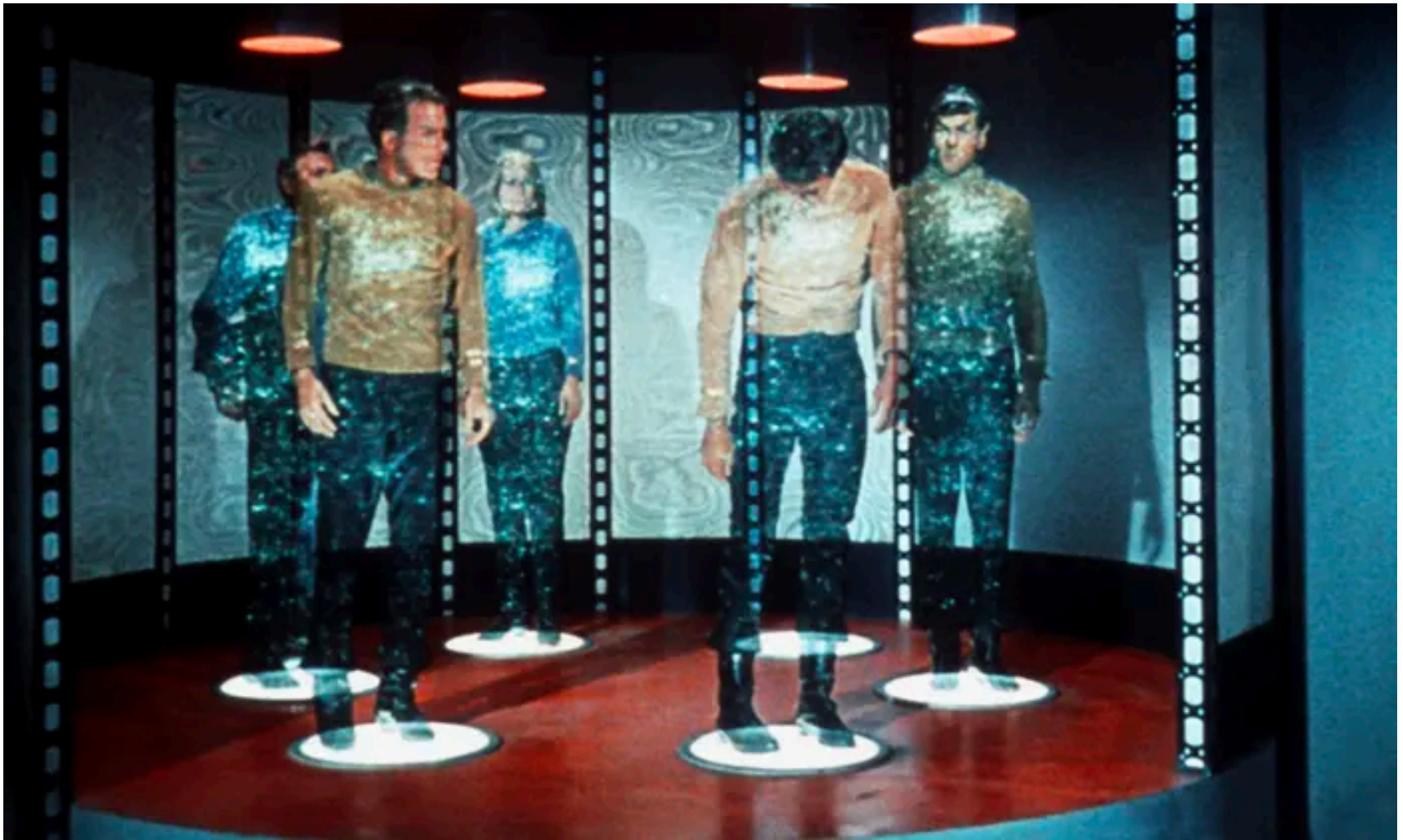
More information in Paggi et al., Annual Review of Biochemistry, 93:389-410, 2024

For rigorous (though outdated) comparisons of docking packages, see Leach et al., J Med Chem 49:5851 (2006); Warren et al., J Med Chem 49:5912 (2006)

# Alternatives methods and current research directions

Optional material

# MD-based approaches



<https://www.theguardian.com/technology/2015/jan/23/german-scientists-teleporter-transporter-3d-printing-star-trek>

# MD-based approaches

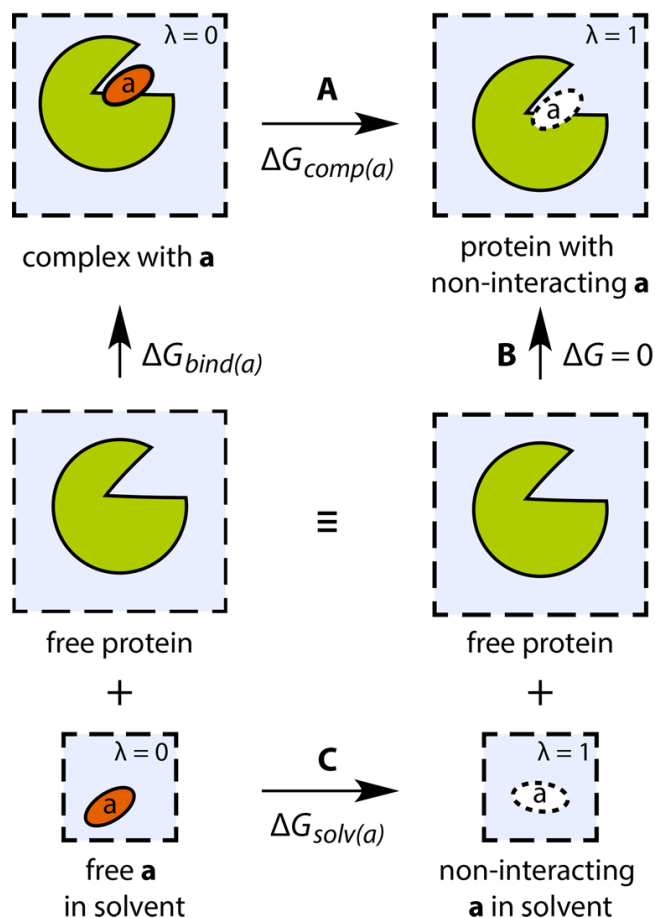
- It turns out that one can compute binding affinities by molecular dynamics simulation without waiting for ligands to spontaneously dissociate (unbind) and bind
- Instead, in “alchemical” methods such as free energy perturbation (FEP), one performs a series of simulations in which the ligand gradually “dematerializes” from its bound position and “materializes” in an unbound position. *This works because binding affinity does not depend on the binding pathway.*
  - These methods currently represent the most accurate way to predict binding affinities, particularly for comparing binding energies of chemically similar ligands, which is how they’re typically used
    - One can determine a difference in binding affinity between two similar ligands by “mutating” one ligand into the other in simulation.
  - These methods assume a known binding pose for each ligand
  - These methods are still very expensive computationally and thus very difficult to use on large numbers of ligands

# MD-based approaches

Calculating binding energy of  
a single ligand

## A. Absolute free energy perturbation

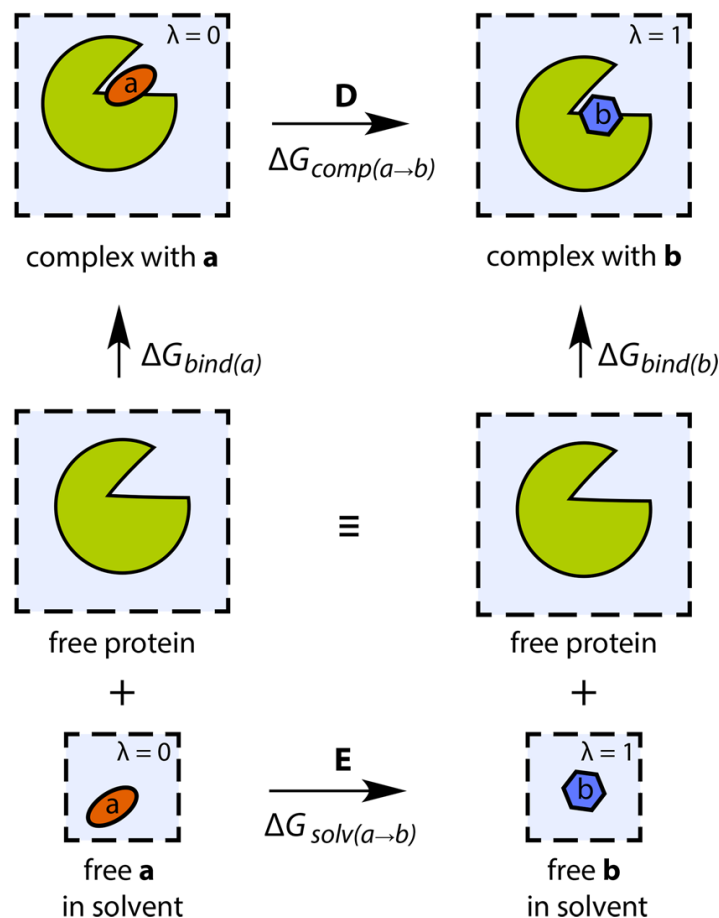
$$\Delta G_{bind(a)} = C + B - A$$



Calculating difference in binding  
energies of two similar ligands

## B. Relative free energy perturbation

$$\Delta\Delta G_{bind(a \rightarrow b)} = D - E$$



# “Ligand-based” approaches

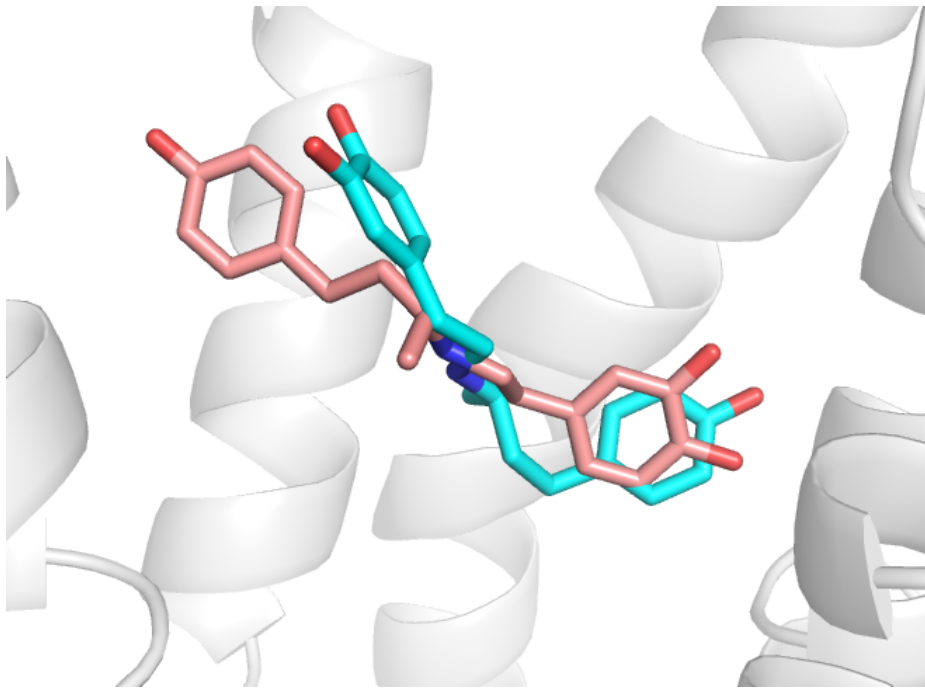
- If one has experimentally measured affinity values for many ligands at a particular target, one can ignore the target structure entirely and simply make affinity predictions based on similarity of query ligand to previously characterized ligands
- These approaches, which date back many decades, are a type of machine learning
- They generally work well only when one has experimentally characterized ligands similar to the query ligand

## Current research area:

# Machine learning approaches for virtual screening

- Both academic research groups and companies are working on deep learning approaches to develop more accurate scoring functions
- The idea is to fit general functional forms (as described by large neural networks), rather than assuming specific functional forms based on approximations to physics
- A variant of this approach is to do reasonably accurate, time-consuming calculations for a subset of the compounds in the library, and then use the results to predict binding affinities of other compounds with faster ligand-based methods

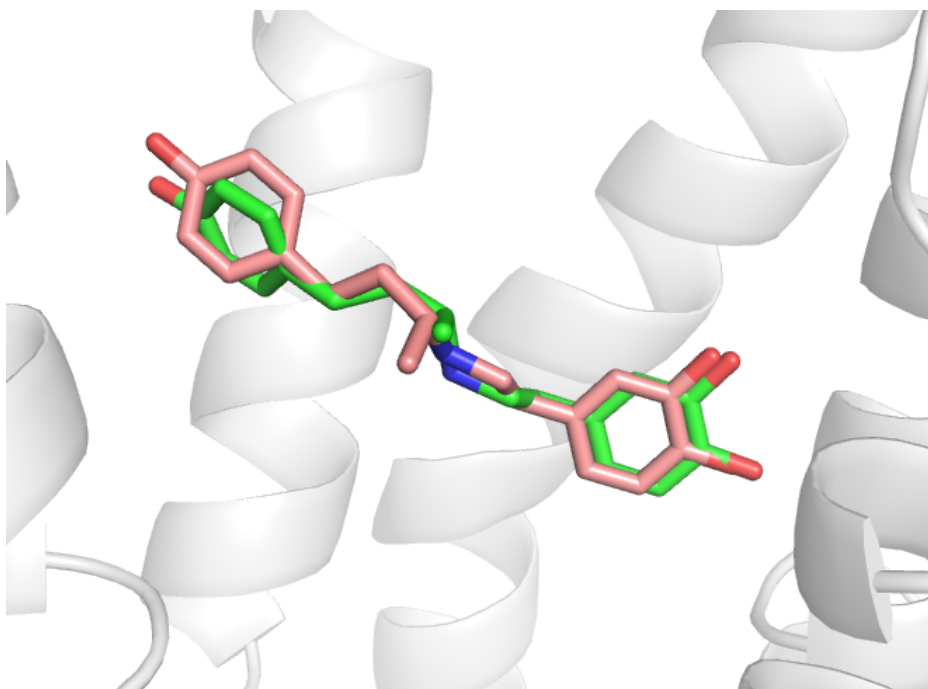
Another machine learning approach:  
experimental information on unrelated ligands can  
substantially improve docking predictions



Experimental structure  
Computational prediction (docking)

Beta agonist dobutamine bound to  
 $\beta$ 1-adrenergic receptor

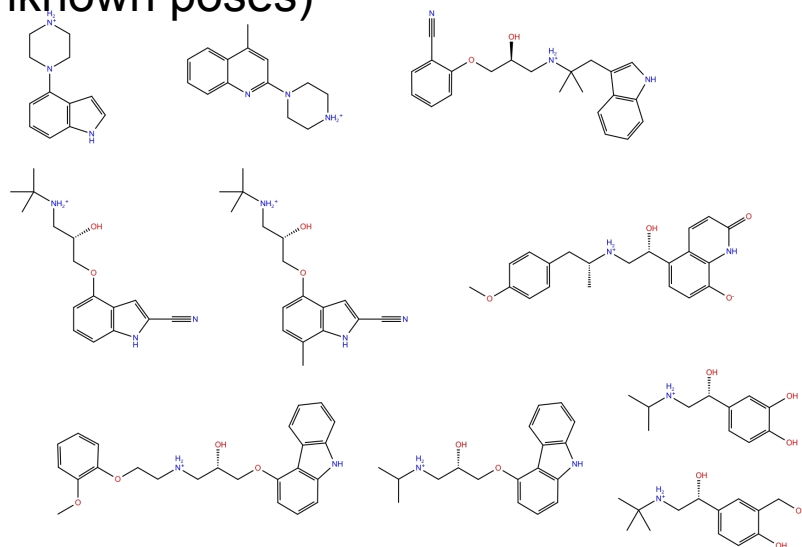
# Another machine learning approach: experimental information on unrelated ligands can substantially improve docking predictions



Beta agonist dobutamine bound to  
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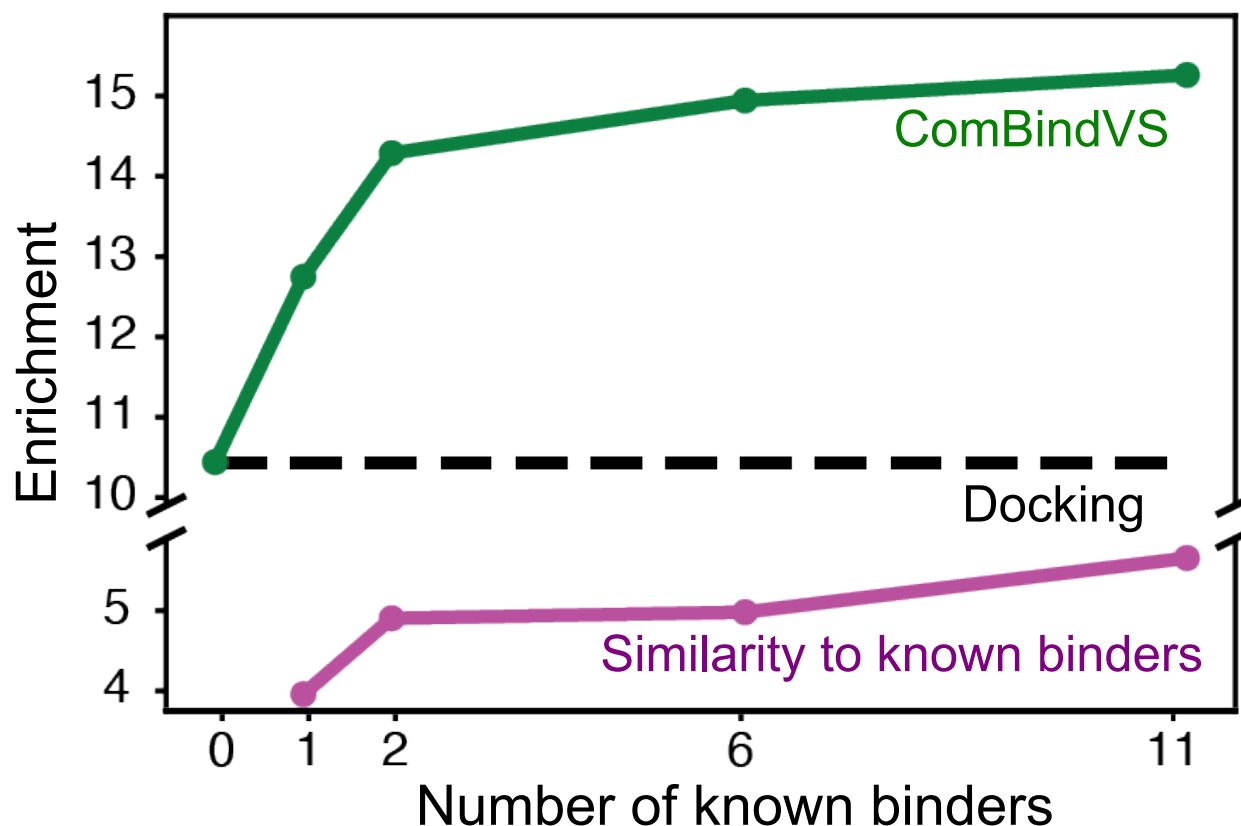
Experimental structure  
Computational prediction (ComBind)

Prediction informed by the fact that the  
compounds below bind this target (in  
unknown poses)



# A similar approach (ComBindVS) improves virtual screening

Average performance across 102 targets in DUD-E benchmark set



Note: All ligands screened are very different from known binders.

# Current research area:

## Generative models

- Instead of learning a scoring function, one can learn to directly generate ligand binding poses or even ligands themselves
- Ligand binding poses
  - Both RoseTTAFold All-Atom and AlphaFold 3 can generate structures of protein-ligand complexes, providing a reasonably accurate docking pose without assuming a fixed conformation of the protein
- Ligands
  - Given a 3D structure of a protein binding pocket, generate ligands that bind tightly to that pocket. This is a very active research area.