

# Tools for understanding brain computations

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Brain and Cognitive Sciences,  
MIT**



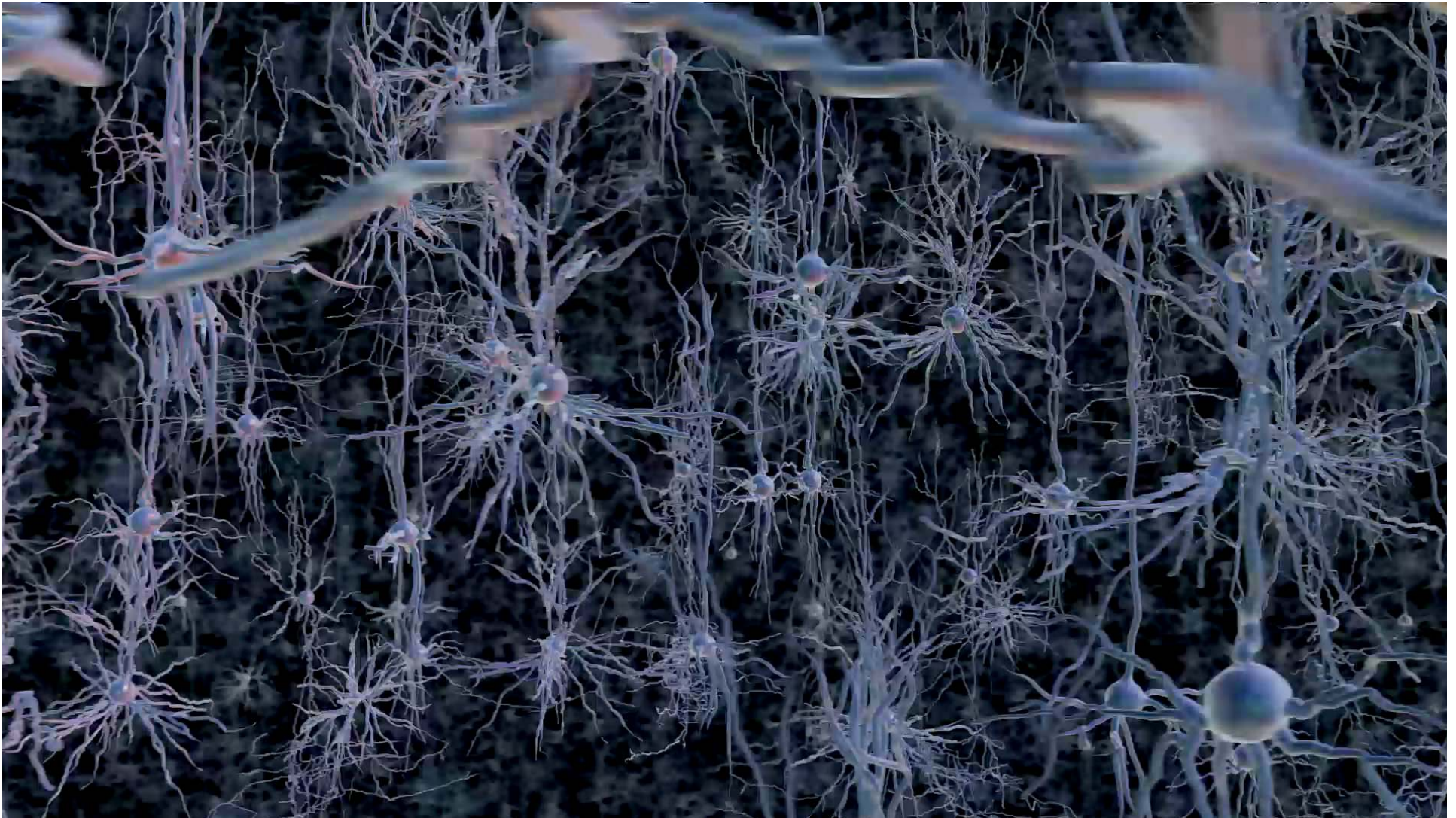
**Understanding complex biological systems, like the brain, is difficult because of wide-ranging spatial and temporal scales**

**Space:** Nanoscale building blocks, spanning centimeters



**Understanding complex biological systems, like the brain, is difficult because of wide-ranging spatial and temporal scales**

**Time:** Millisecond events, spanning seconds to years



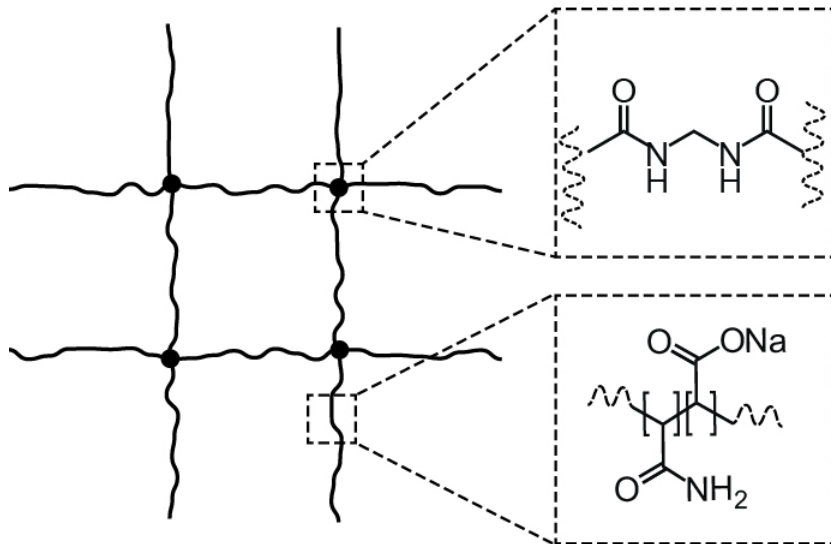
# **Map: molecules**

How can we map complex biological systems, like brain circuits, with molecular precision?

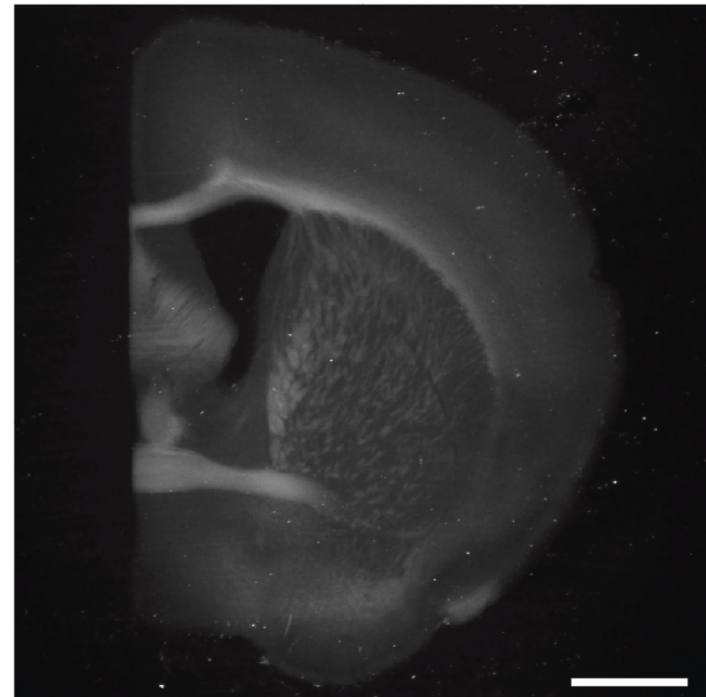
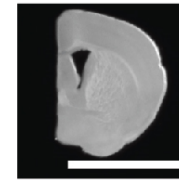


# Physical magnification: a new principle of microscopy

**Sodium polyacrylate:**  
polymer that swells in water  
(found in baby diapers)

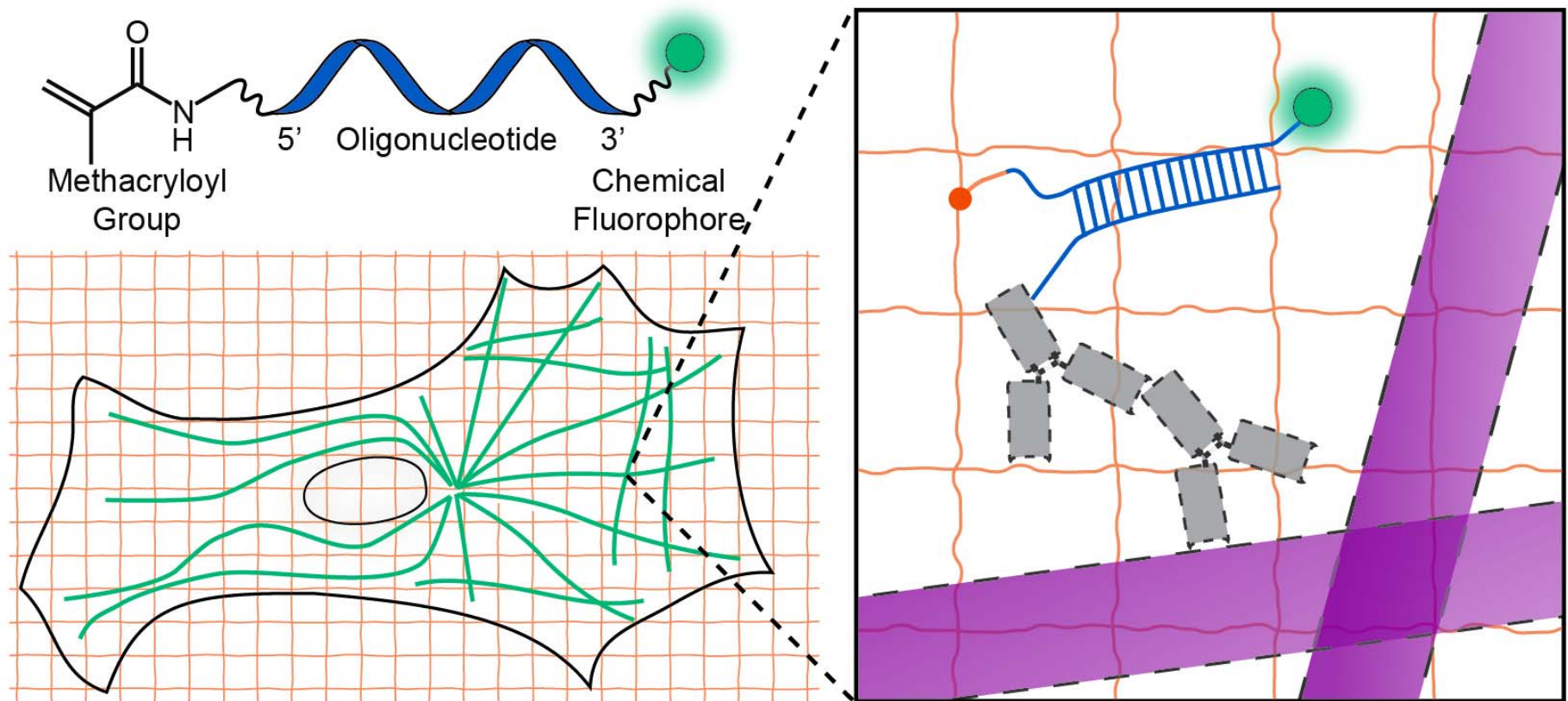


**Embed brain**  
in permeating polyacrylate  
network, then add water



**Chen\*, Tillberg\*, Boyden (2015) *Science* 347(6221):543-548. <http://expansionmicroscopy.org>  
Commercialized by Expansion Technologies, Inc. (ESB is co-founder)**

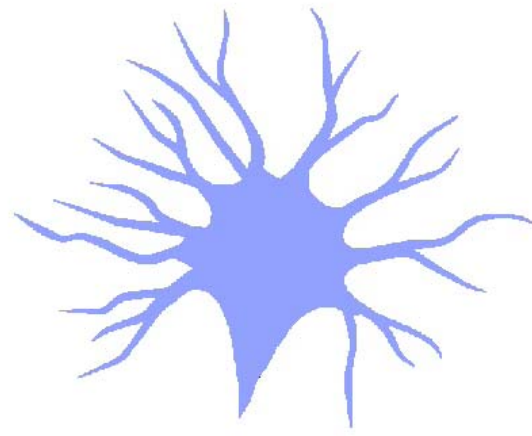
# Expansion Microscopy (ExM)



1. Preserve tissue by fixation and permeabilization.
2. Immunostain with a fluorescent label bearing a polymer-linkable group.
3. Diffuse in acrylate monomer, and polymerize, linking fluorescent label to polymer.
4. Use protease to digest everything but the polymer and polymer-anchored labels.
5. Add water to swell the polymer.

Chen\*, Tillberg\*, Boyden (2015) *Science* 347(6221):543-548.

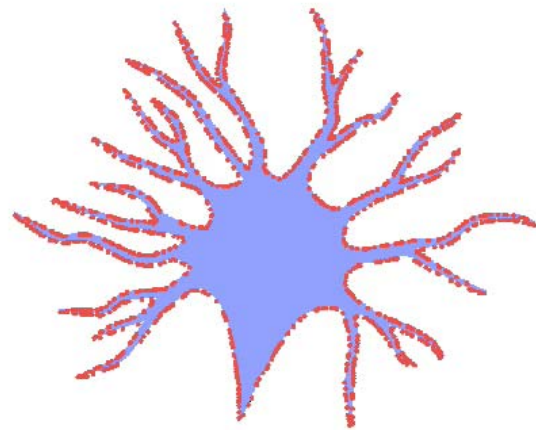
# ExM Process



10 microns

- Fix specimen
- Tri-functional label
- Form swellable gel
- Digest specimen
- Expand in water

# ExM Process

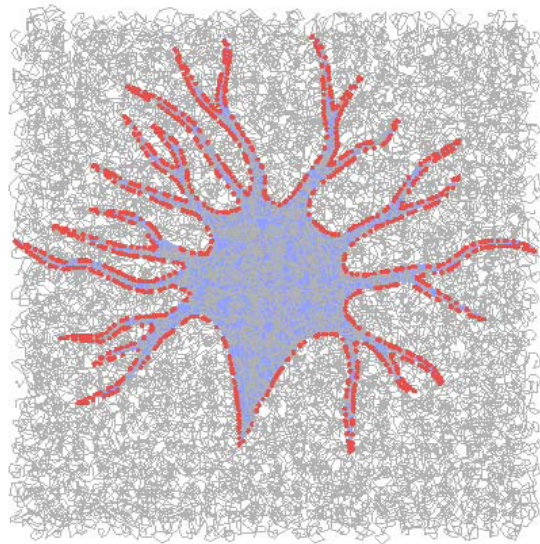


10 microns

- Fix specimen
- **Tri-functional label**
- Form swellable gel
- Digest specimen
- Expand in water



# ExM Process

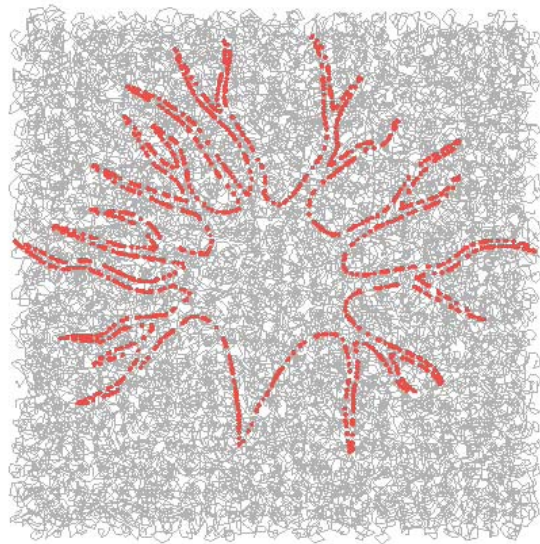


10 microns

- Fix specimen
- Tri-functional label
- **Form swellable gel**
- Digest specimen
- Expand in water

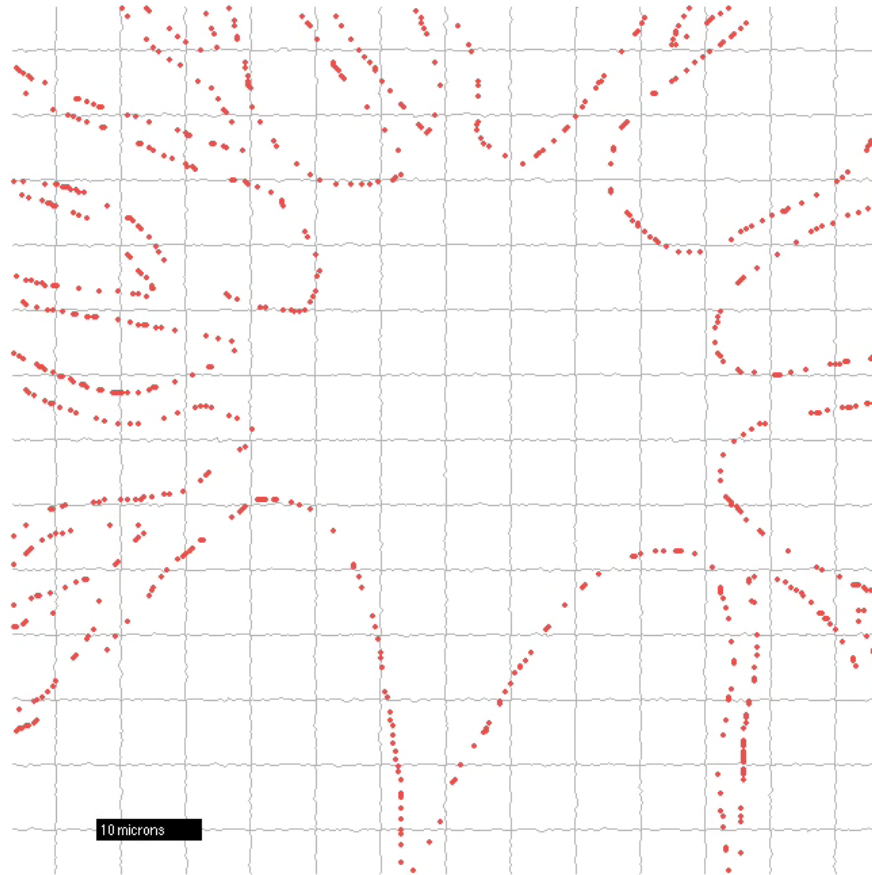
Chen\*, Tillberg\*, Boyden (2015) *Science* 347(6221):543-548.

# ExM Process



- Fix specimen
- Tri-functional label
- Form swellable gel
- **Digest specimen**
- Expand in water

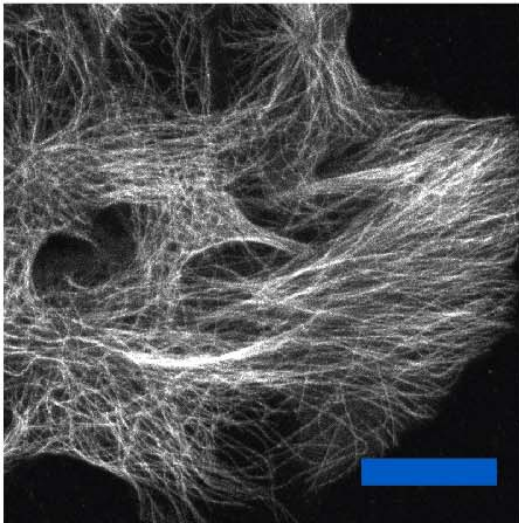
# ExM Process



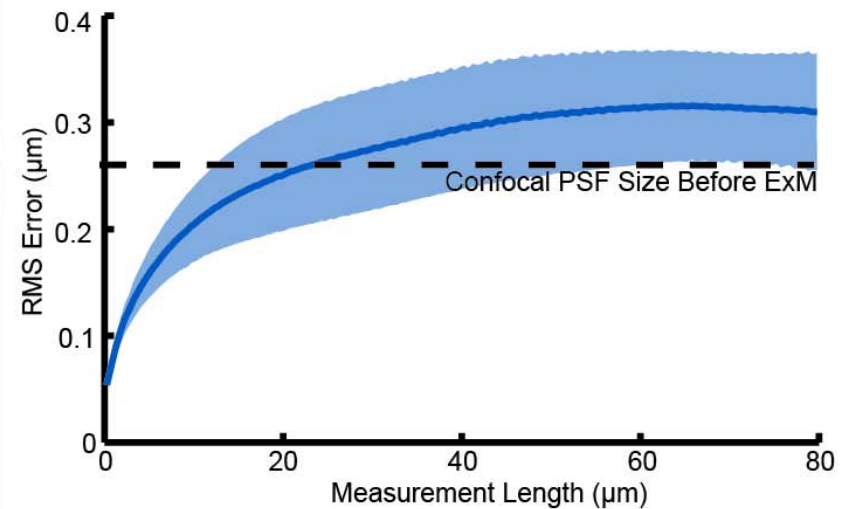
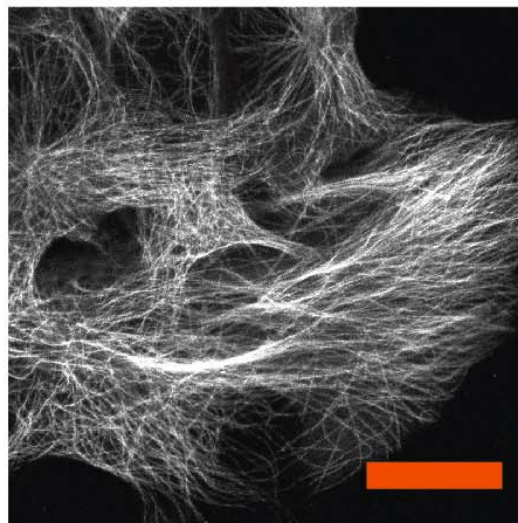
- Fix specimen
- Tri-functional label
- Form swellable gel
- Digest specimen
- **Expand in water**

# ExM preserves global isotropy: cultured cells, microtubules

Before expansion:  
confocal microscopy

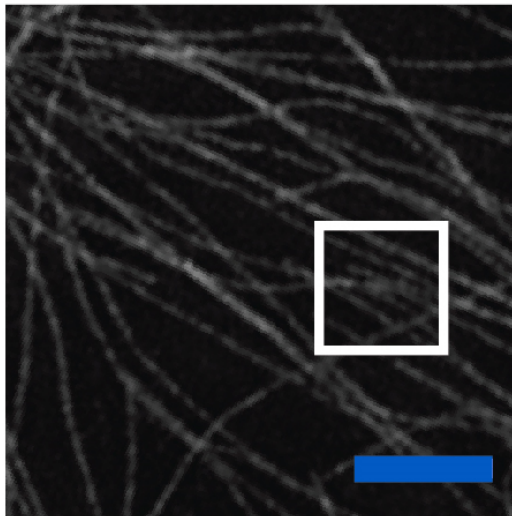


After expansion (~4.5x):  
confocal microscopy

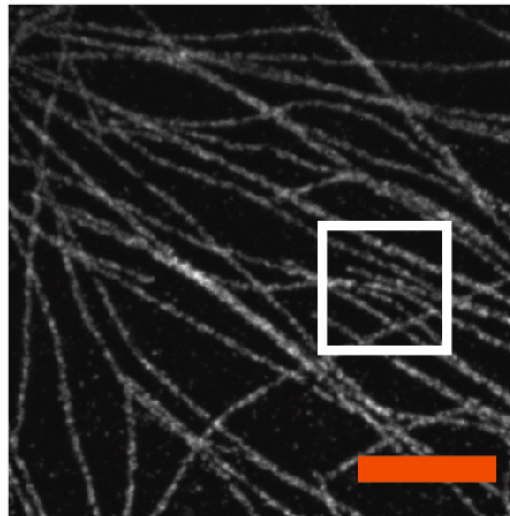


# ExM preserves nanoscale isotropy: cultured cells, microtubules

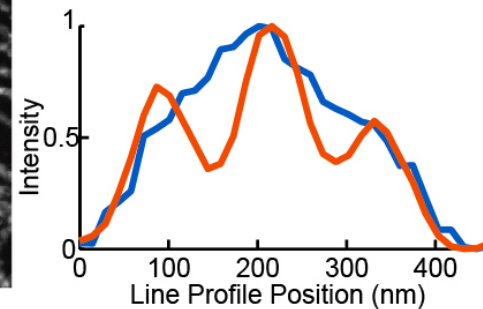
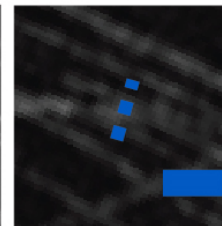
Before expansion:  
super-resolution  
structured illumination  
microscopy (SR-SIM)



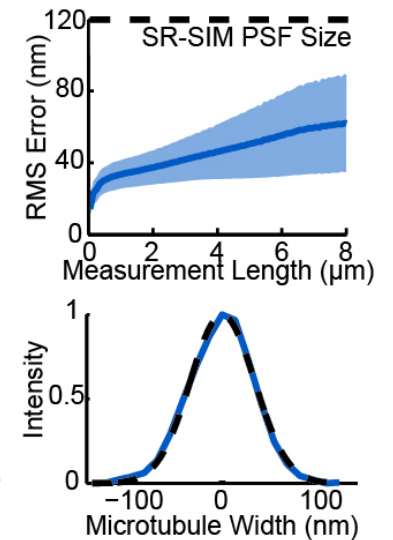
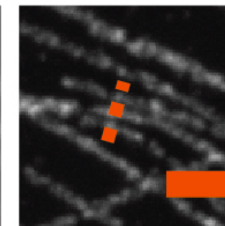
After expansion (~4.5x):  
confocal microscopy



Before:  
SR-SIM



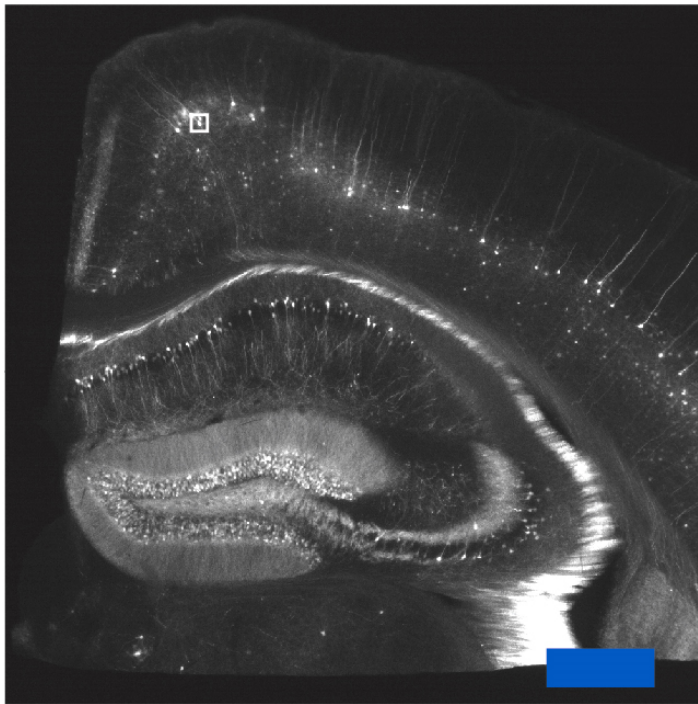
After:  
confocal



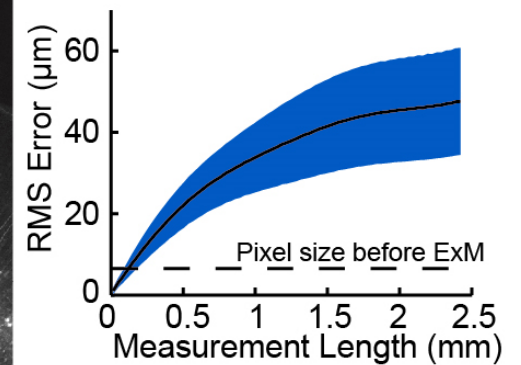
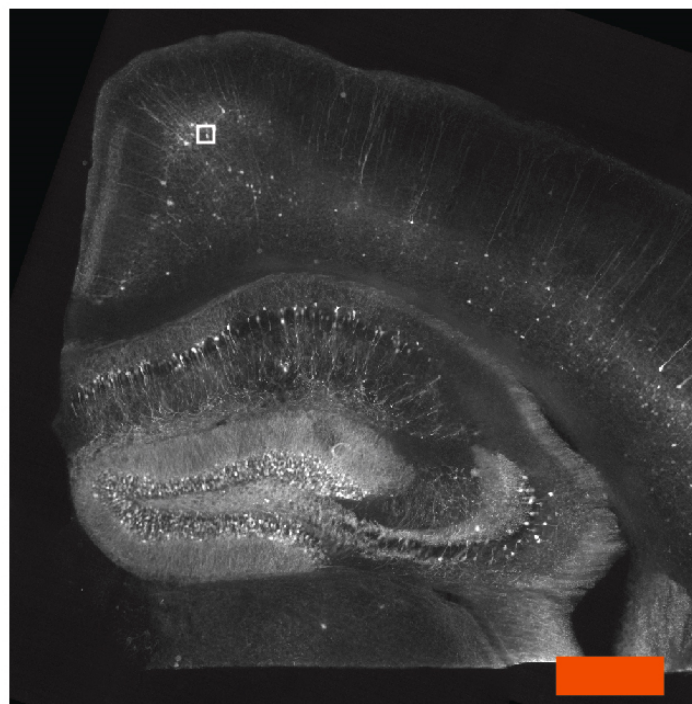


# ExM preserves global isotropy: brain, YFP-expressing neurons

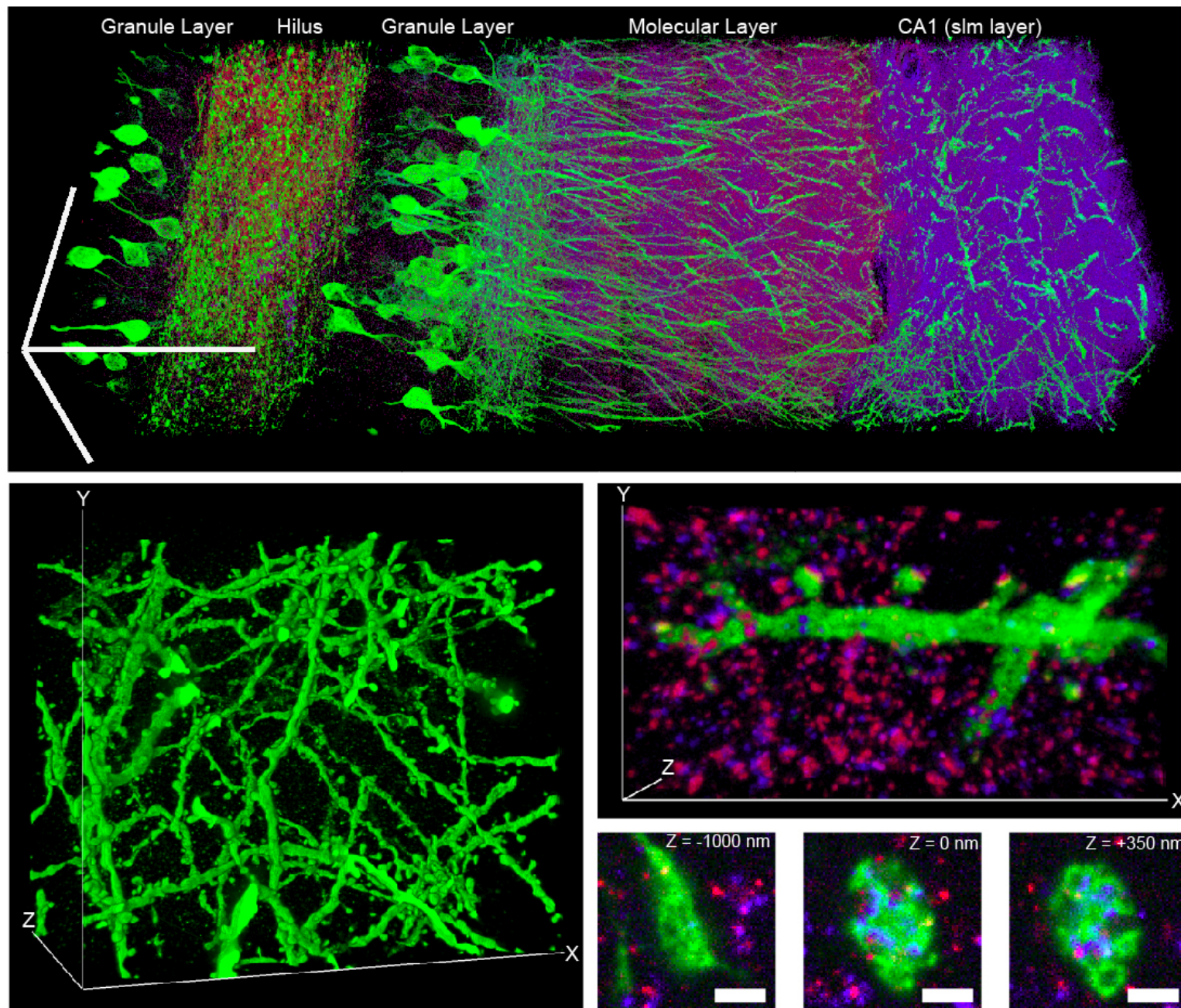
Before expansion:  
confocal microscopy



After expansion (~4.5x):  
confocal microscopy



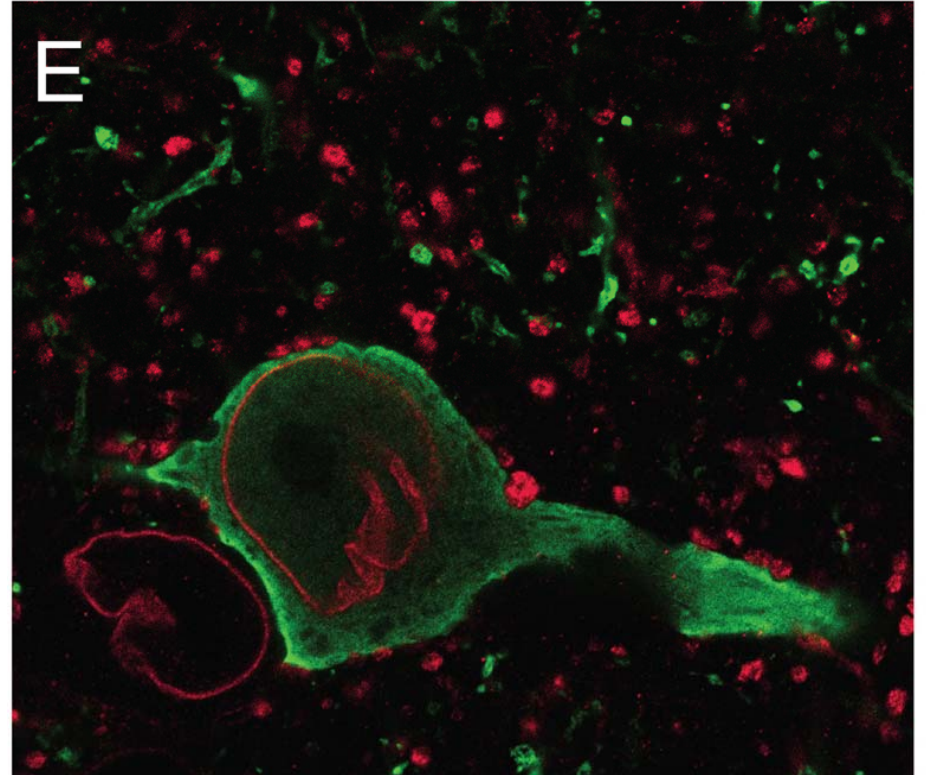
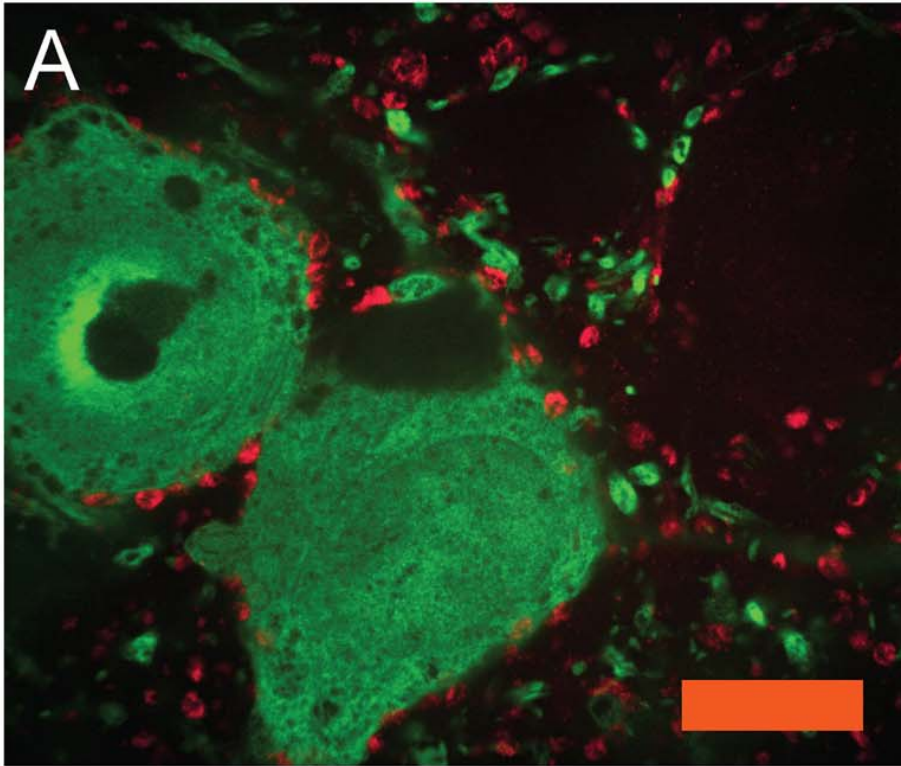
**ExM: voxel sizes of a super-resolution microscope, voxel rates of a diffraction-limited microscope**



**Chen\*, Tillberg\*, Boyden (2015) *Science* 347(6221):543-548.**



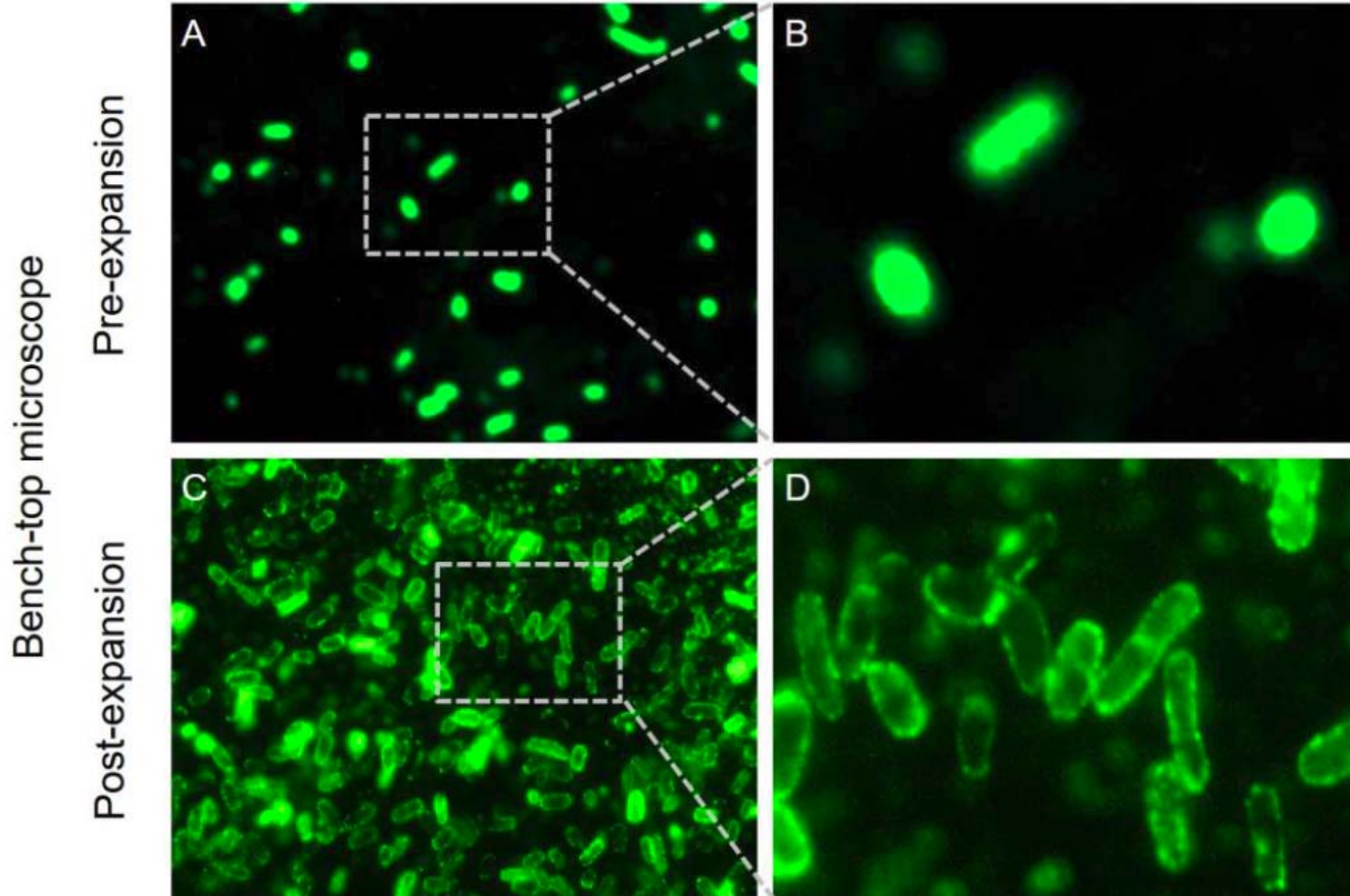
## Lots of interesting things to look at



ExM imaging of antibodies of interest in neuroscience and biology. Confocal images of expanded Thy1-YFP mouse brain cerebral cortex sections stained with anti-GFP (green) and antibodies against other proteins (red) as follows: (A) GAD65/67 (22), (B) ChAT (23), (C) CaMKII (24), (D) GABA (25), (E) Lamin A/C (26), (F) NMDAR2a/b (27). Scale bars: (A) 10  $\mu\text{m}$  in pre-expansion units (physical size post-expansion, 45  $\mu\text{m}$ ); (B) 10  $\mu\text{m}$  (47  $\mu\text{m}$ ); (C) 10  $\mu\text{m}$  (40  $\mu\text{m}$ ); (D) 10  $\mu\text{m}$  (44  $\mu\text{m}$ ); (E) 10  $\mu\text{m}$  (43  $\mu\text{m}$ ); (F) 10  $\mu\text{m}$  (43  $\mu\text{m}$ ).

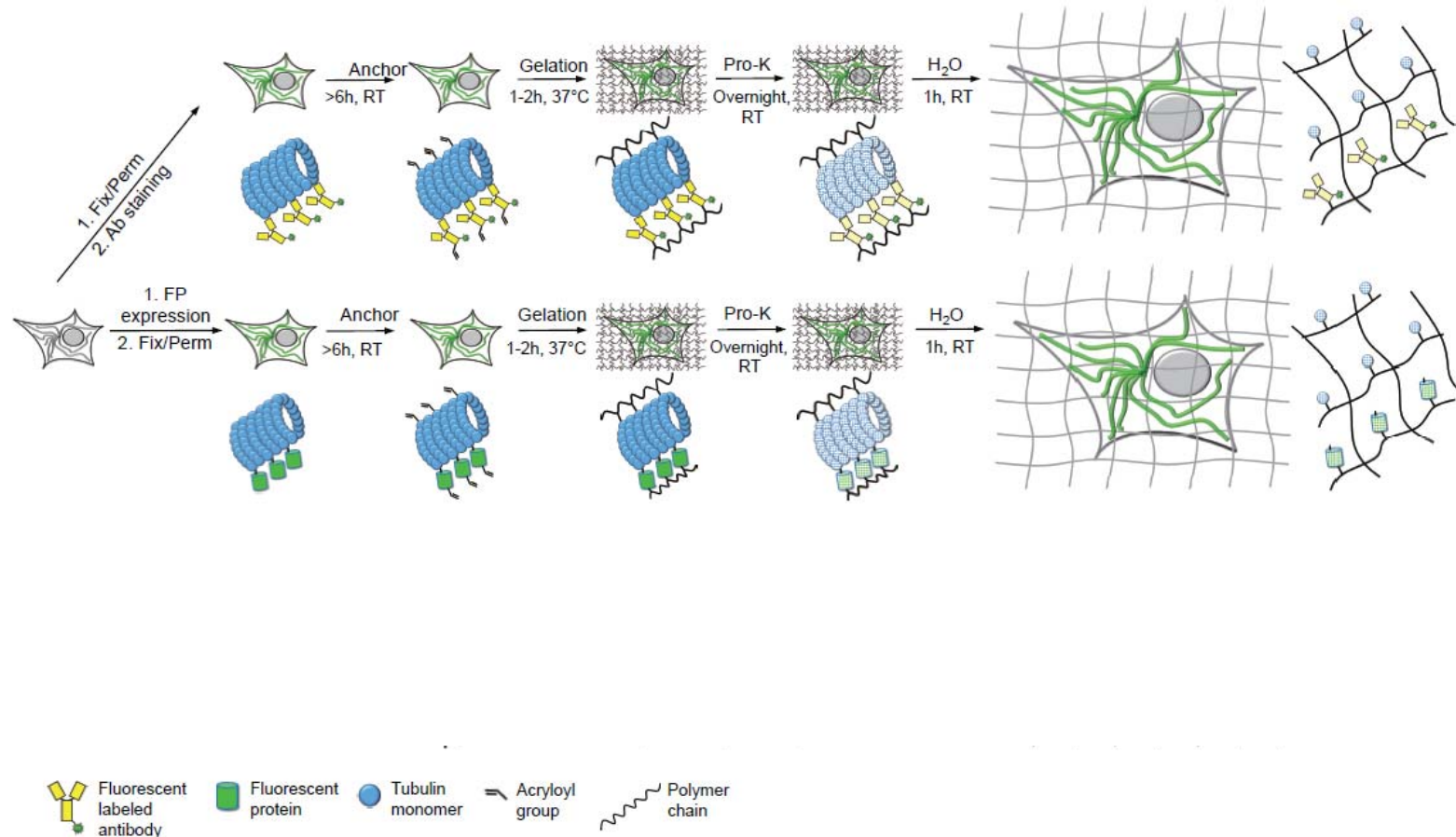
**Chen\*, Tillberg\*, Boyden (2015) *Science* 347(6221):543-548.**

# Bacteria



Zhang YS\*, Chang JB\*, et al. (2016) *Scientific Reports* 6:22691.

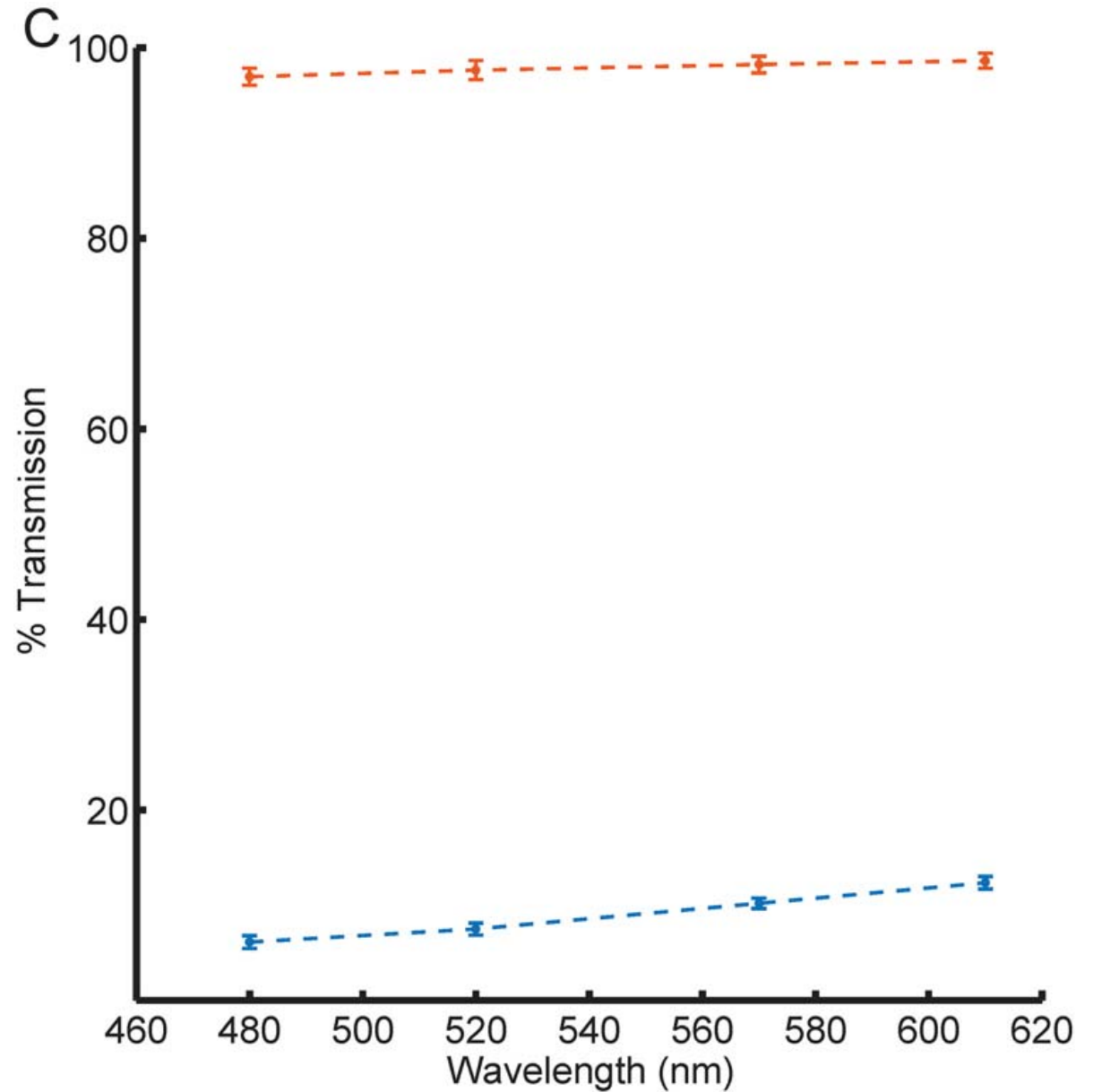
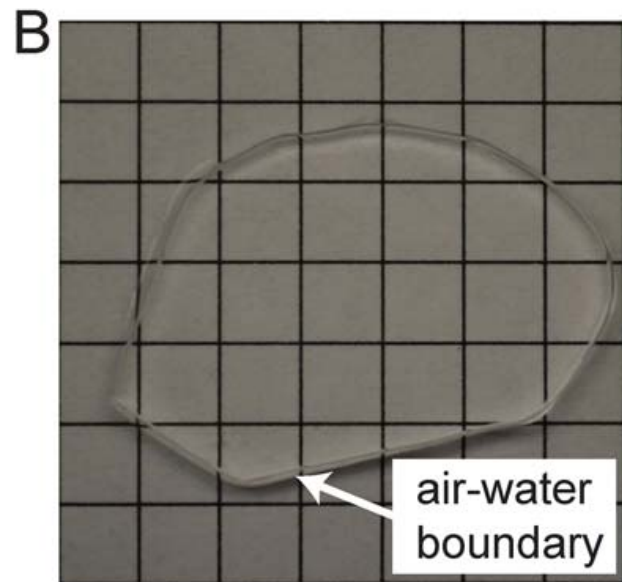
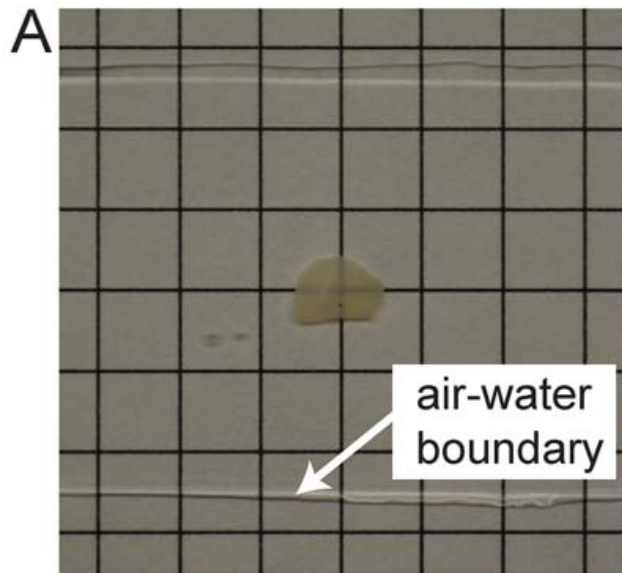
# proExM: ExM with commercially available chemicals, like a normal immunohistochemistry workflow



Paul W. Tillberg, Fei Chen, Kiryl D. Piatkevich, Yongxin Zhao, Chih-Chieh (Jay) Yu, Brian P. English, Linyi Gao, Anthony Martorell, Ho-Jun Suk, Fumiaki Yoshida, Ellen DeGennaro, Douglas H. Roossien, Robert Desimone, Dawen Cai



# Expanded samples are almost completely transparent



Chen\*, Tillberg\*, Boyden (2015) *Science* 347(6221):543-548.

# **Record: dynamics**

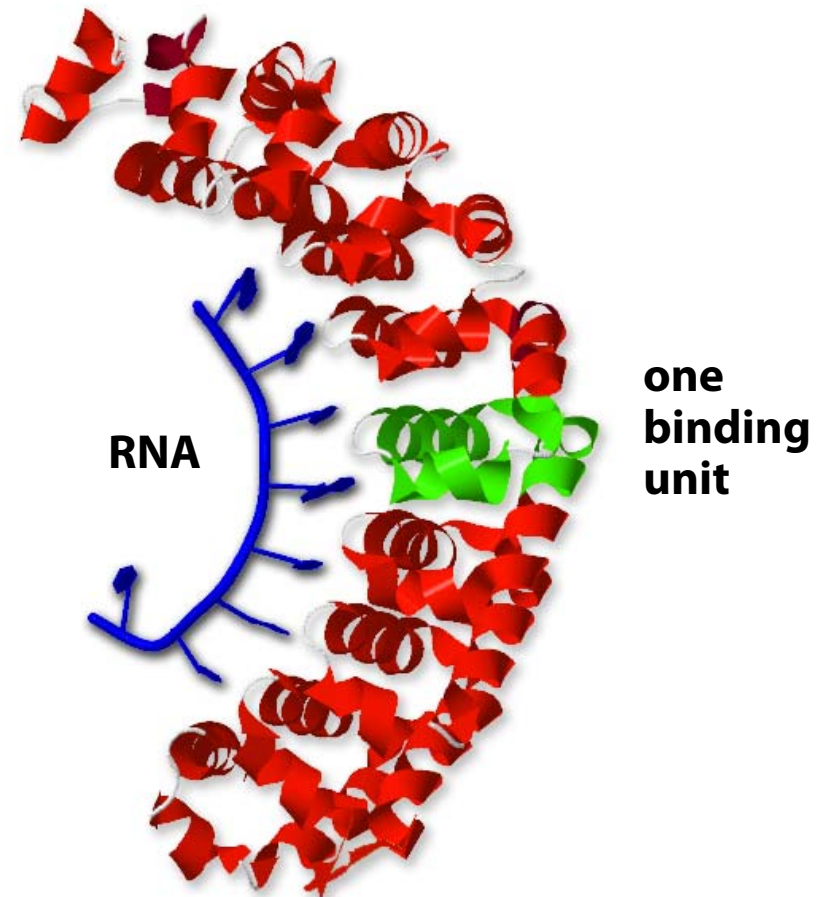
How can we record biological dynamics with precise temporal, and broad spatial, scale?

# A scaffold for designing RNA-binding proteins

**Pumilio homolog 1 (PumHD):**  
sequence-specific RNA-binding protein that regulates translation and mRNA stability by binding the 3'-UTR of mRNA targets.

It's made of a series of **36 amino acid** repeats. Each **repeat** binds a single RNA base.

Each 36 amino acid binding unit is a helical structure with **three amino acids** involved in contact with RNA.

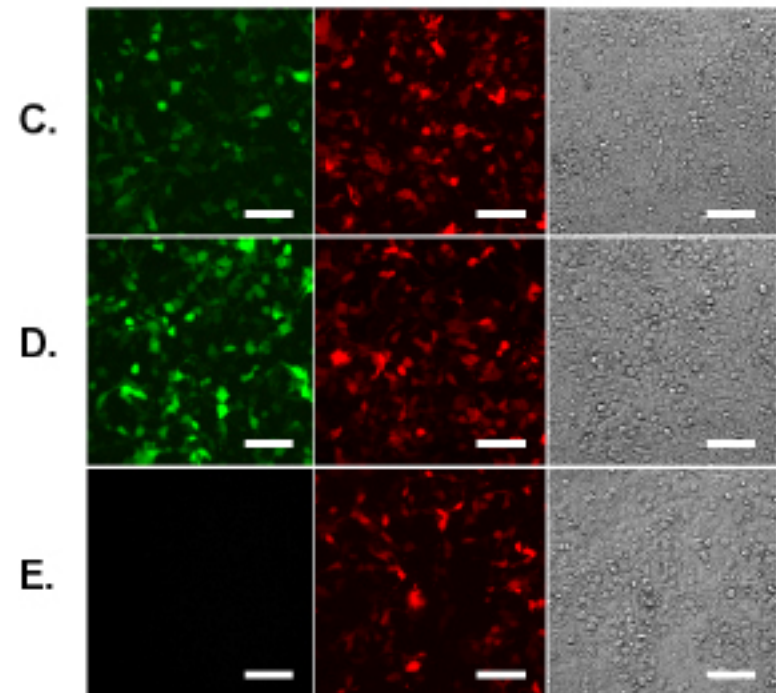
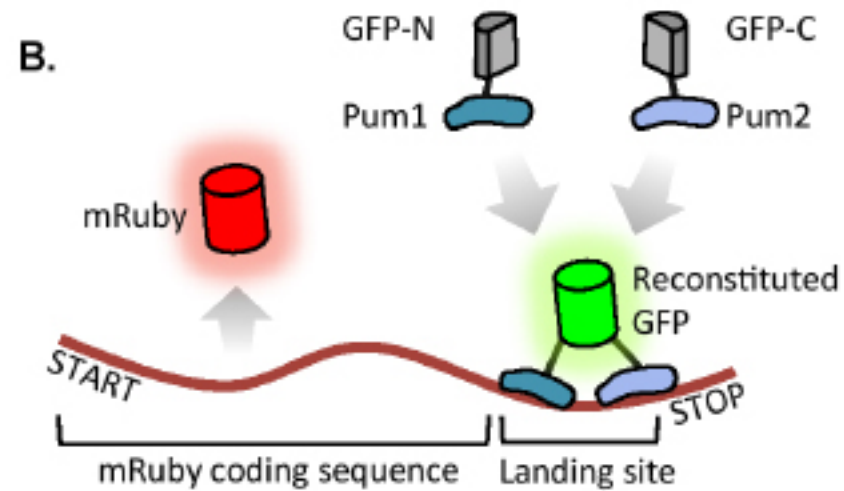
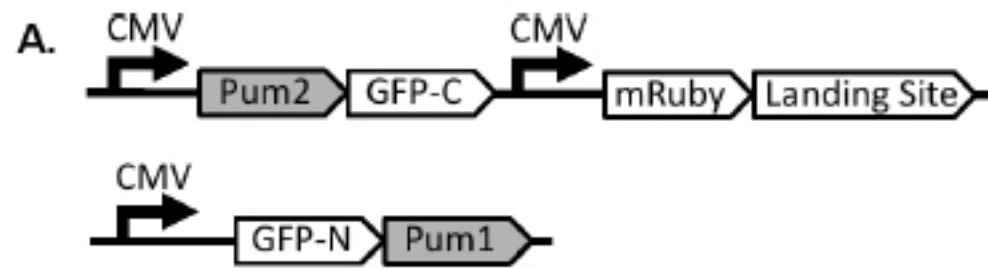


human Pumilio1 in  
complex with RNA;  
PDB 1M8X

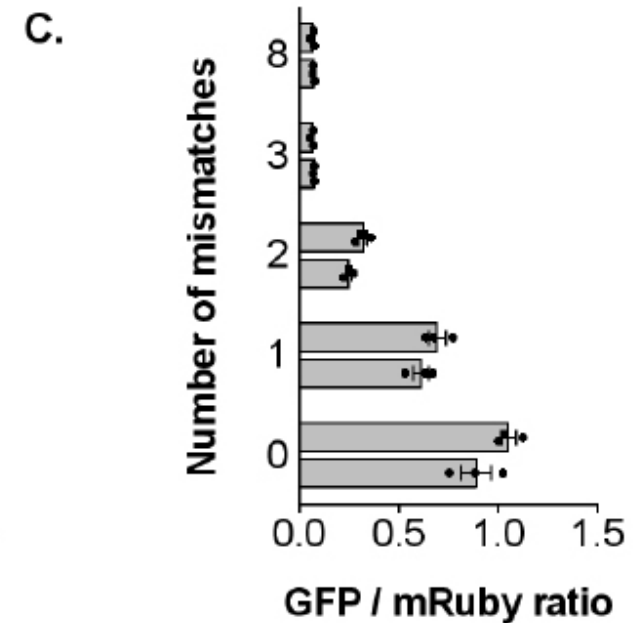
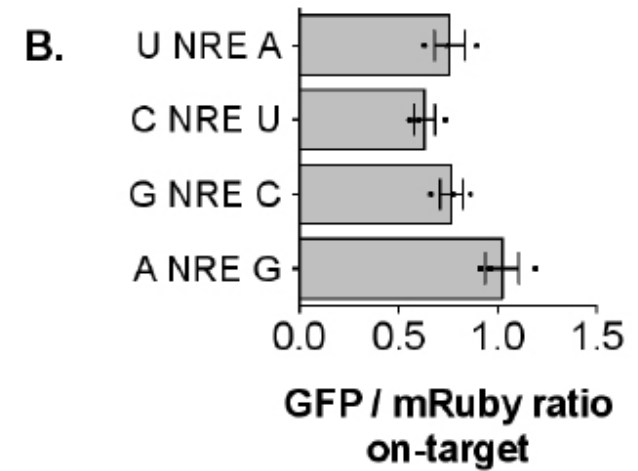
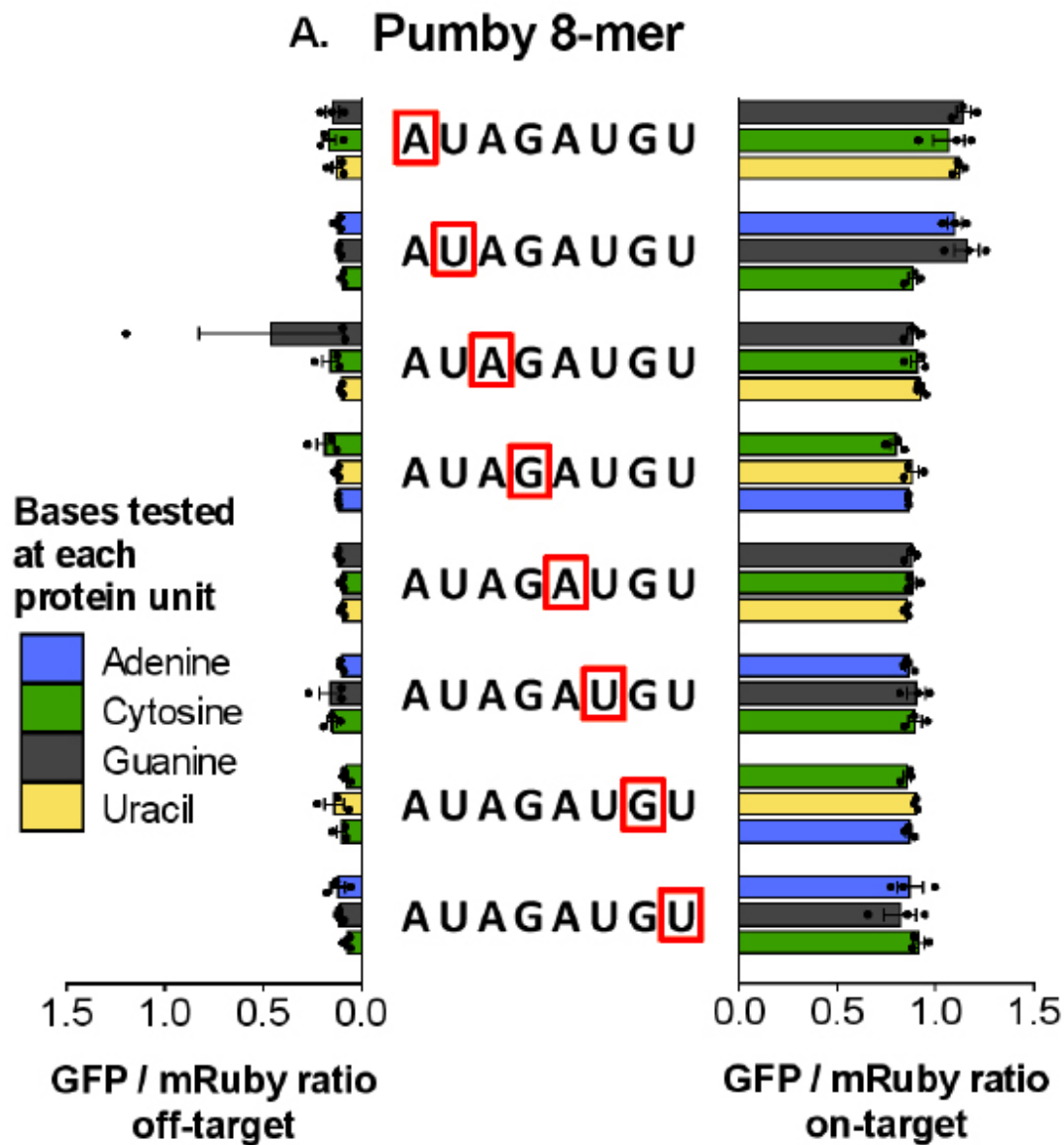
Adamala\*, Martin-Alarcon\*, Boyden (2016) in press.





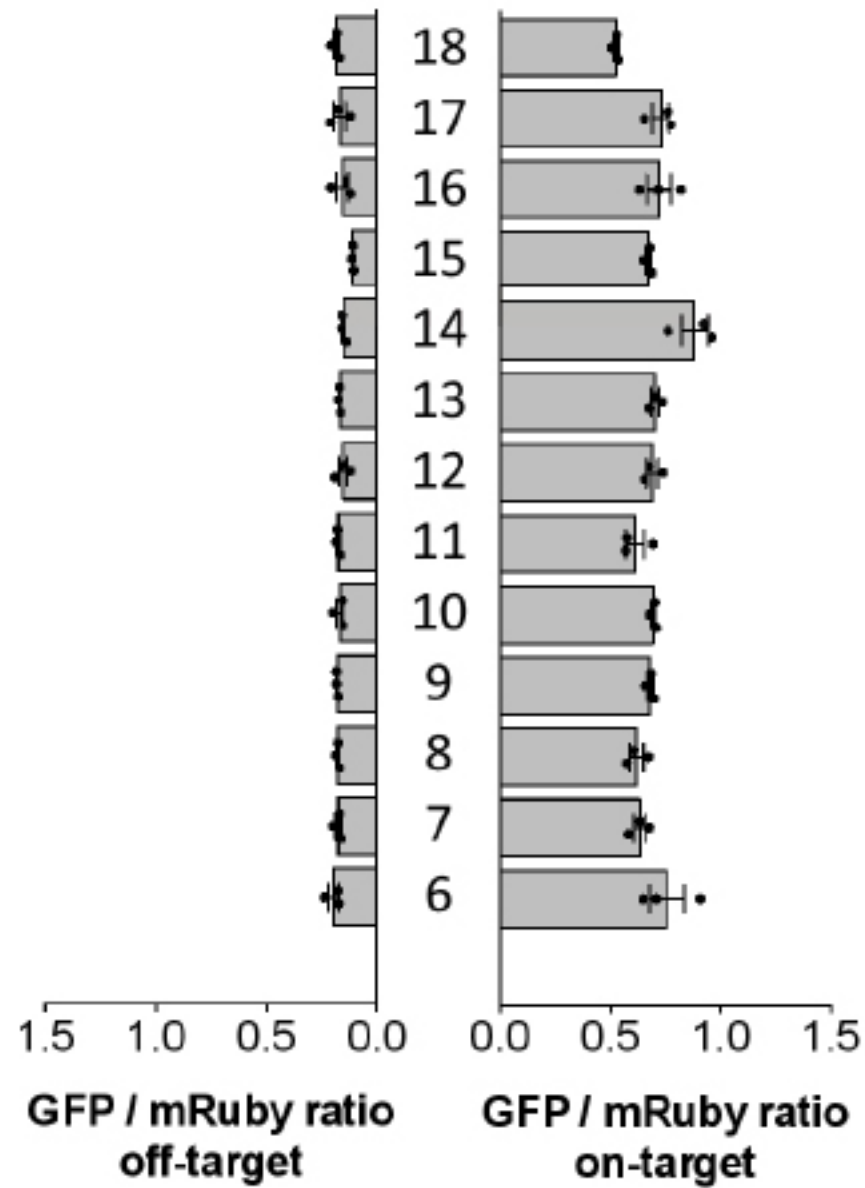


Adamala\*, Martin-Alarcon\*, Boyden (2016) in press.

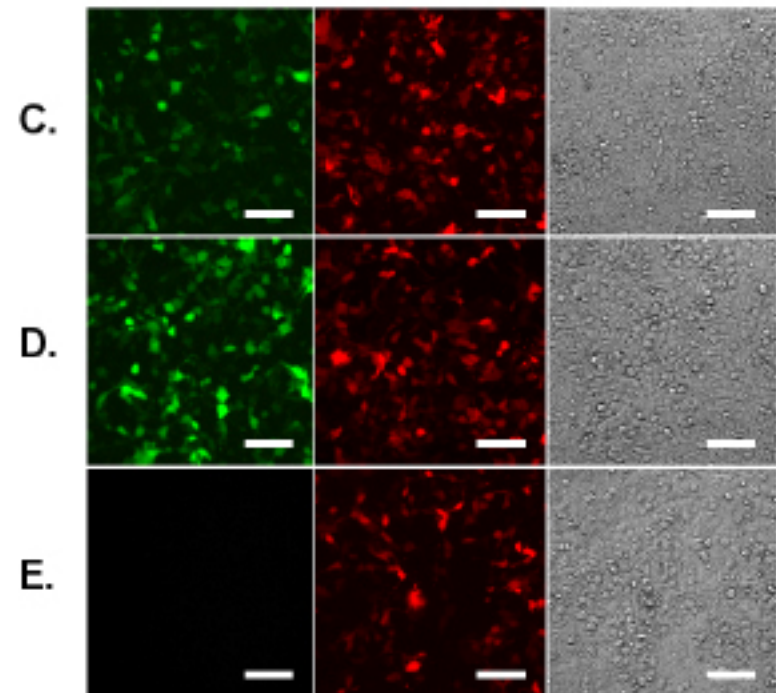
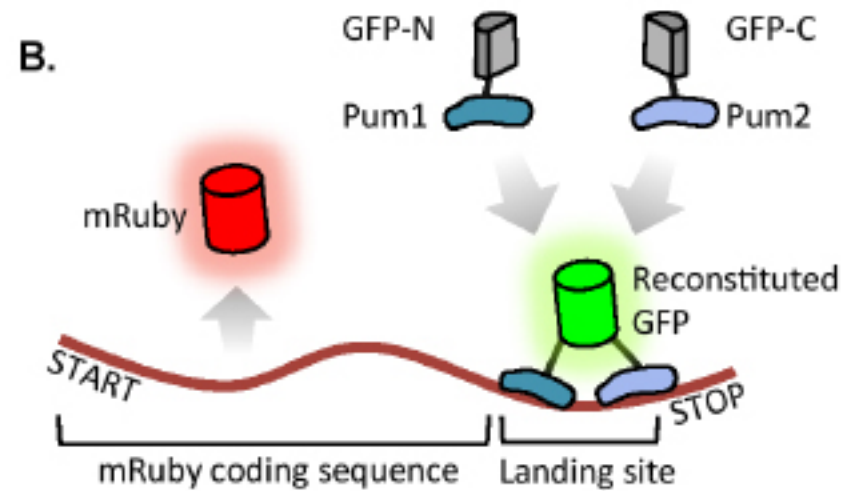
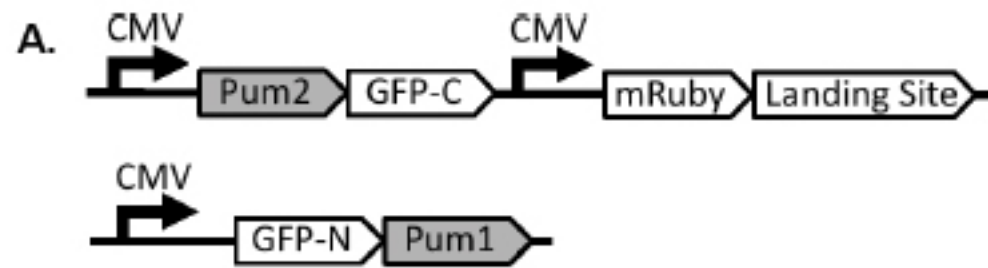


Adamala\*, Martin-Alarcon\*, Boyden (2016) in press.

#### D. Pumby n-mer

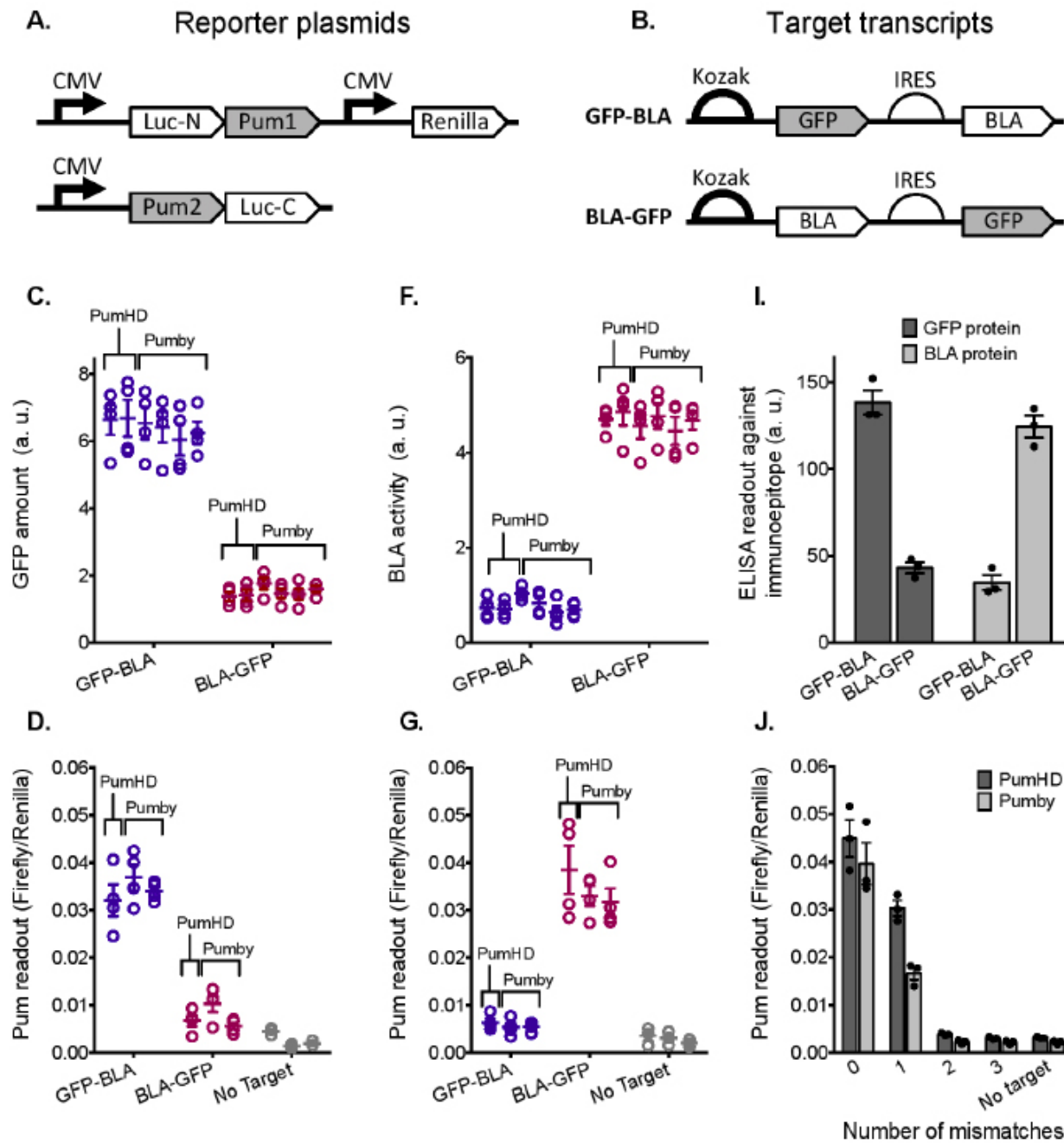


Adamala\*, Martin-Alarcon\*, Boyden (2016) in press.

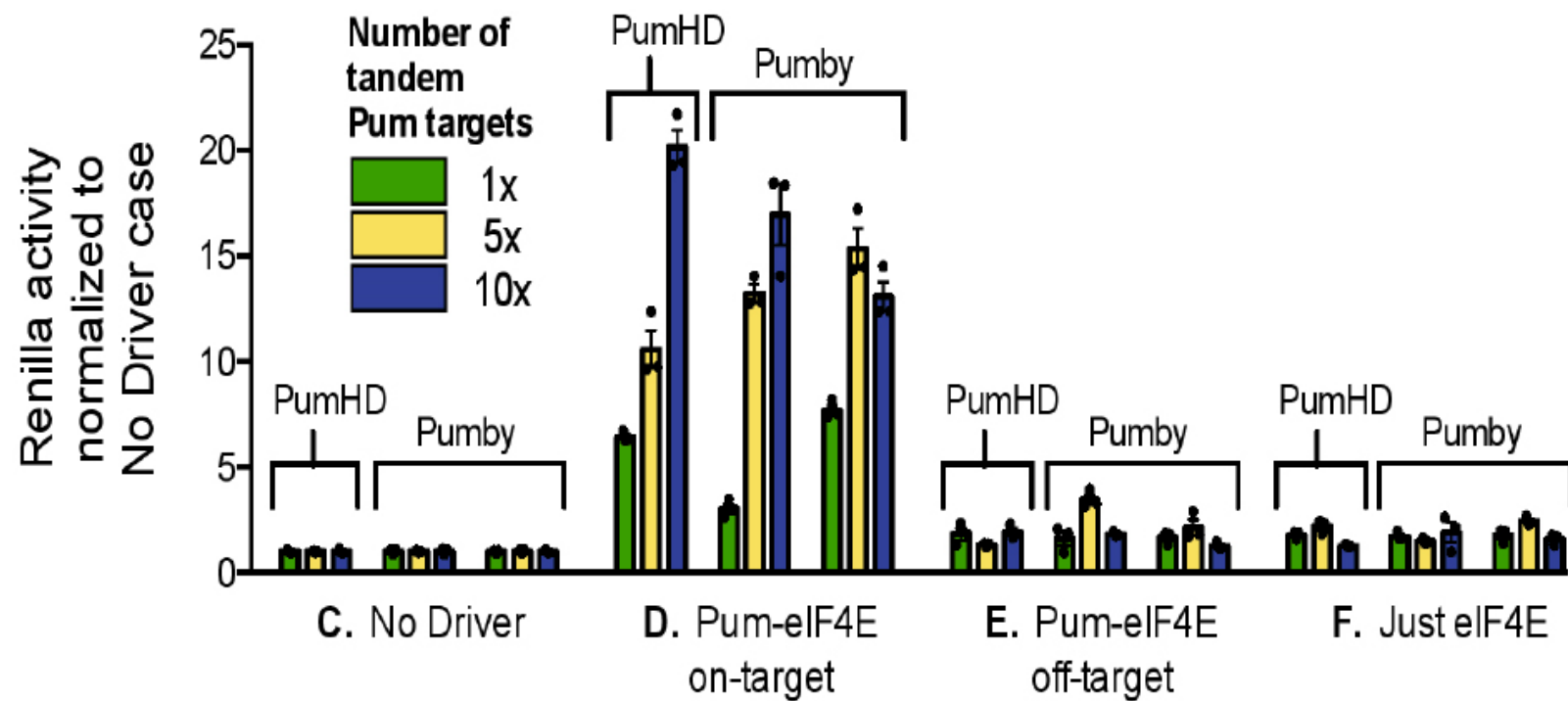
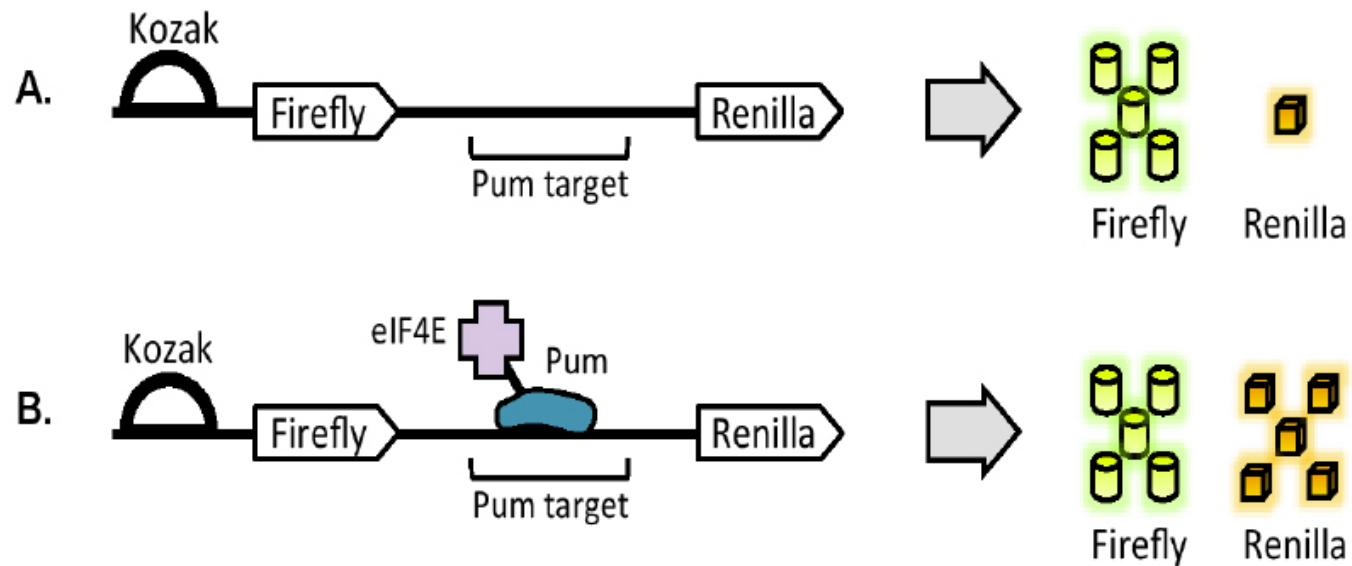


Adamala\*, Martin-Alarcon\*, Boyden (2016) in press.



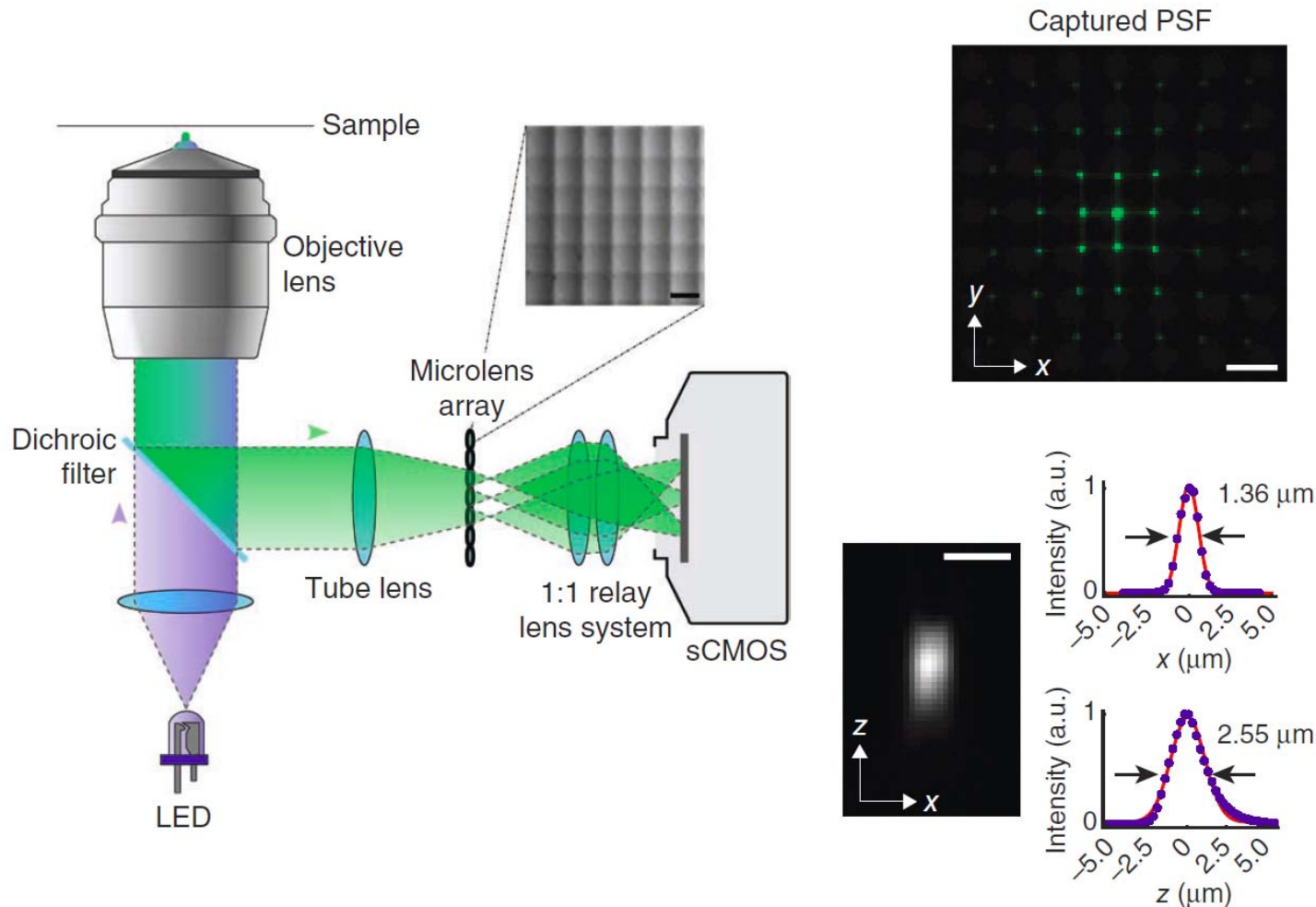


Adamala\*, Martin-Alarcon\*, Boyden (2016) in press.



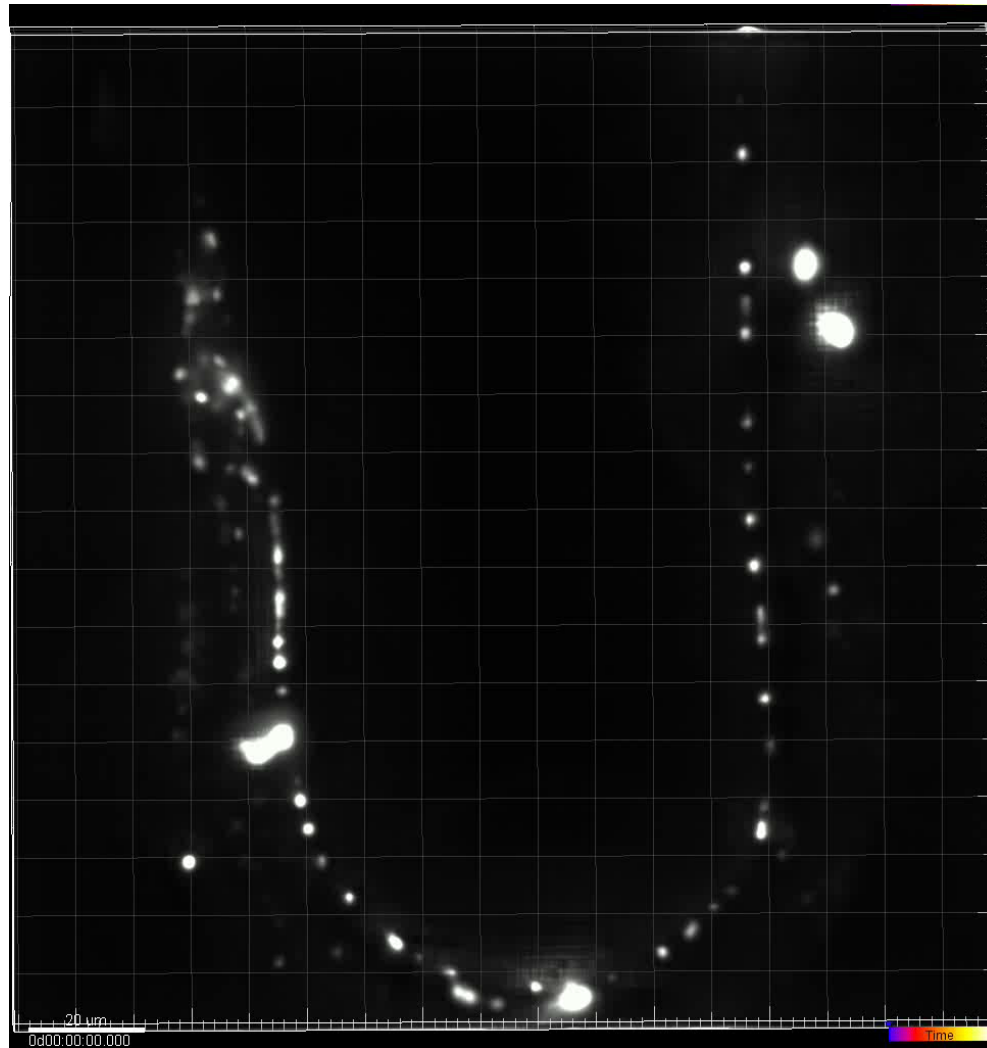
Adamala\*, Martin-Alarcon\*, Boyden (2016) in press.

# Simultaneous, whole-animal, 3-D microscopy: light-field imaging



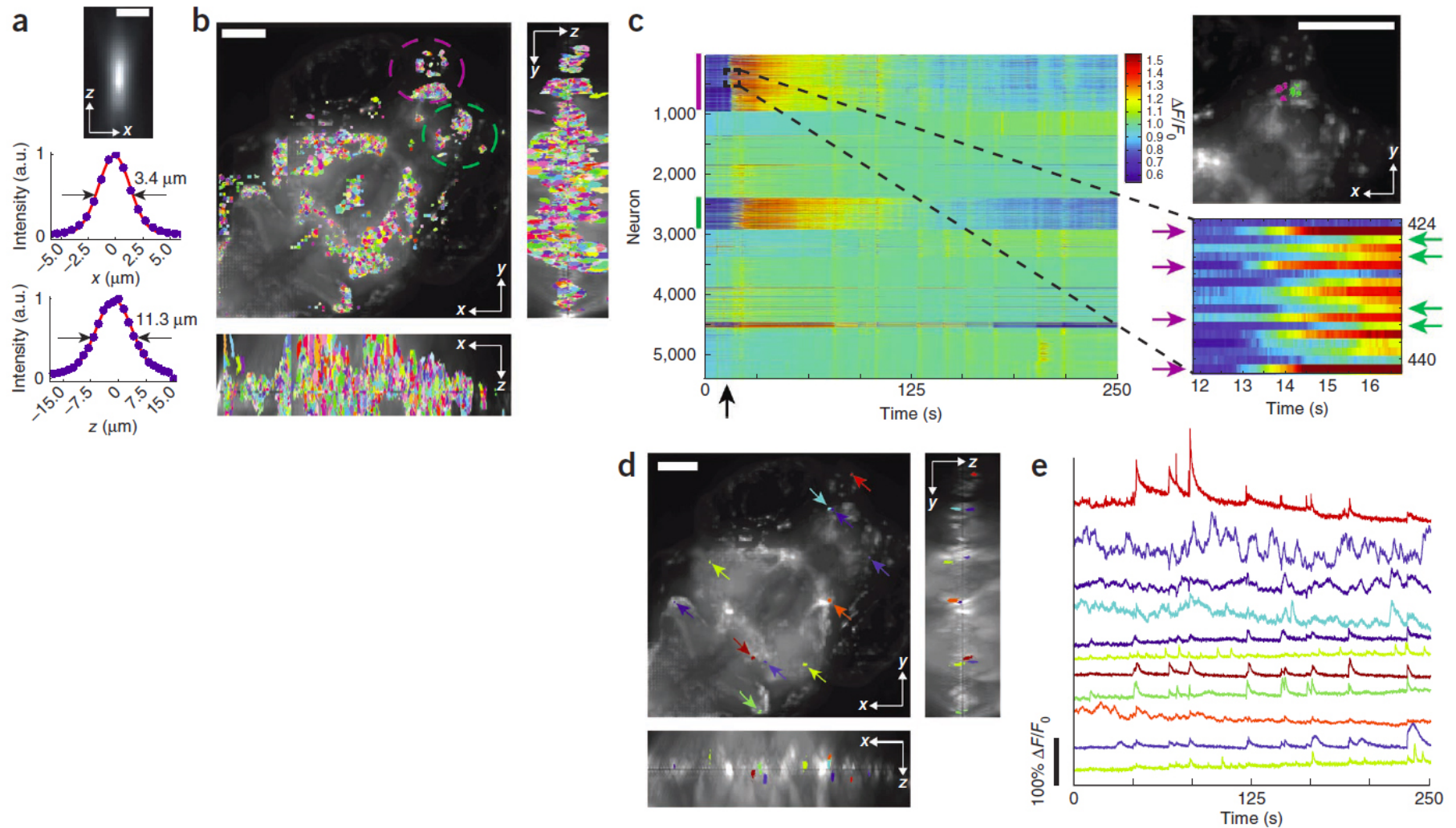
Prevedel\*, Yoon\*, et al. (2014) *Nature Methods* 11:727-730.  
Plans, parts lists, software at <http://lightfieldscope.org>

# Imaging neural activity throughout organism with known connectome



Prevedel\*, Yoon\*, et al. (2014) *Nature Methods* 11:727-730.

# Imaging zebrafish neural activity in 3-D (at 20 Hz, below)

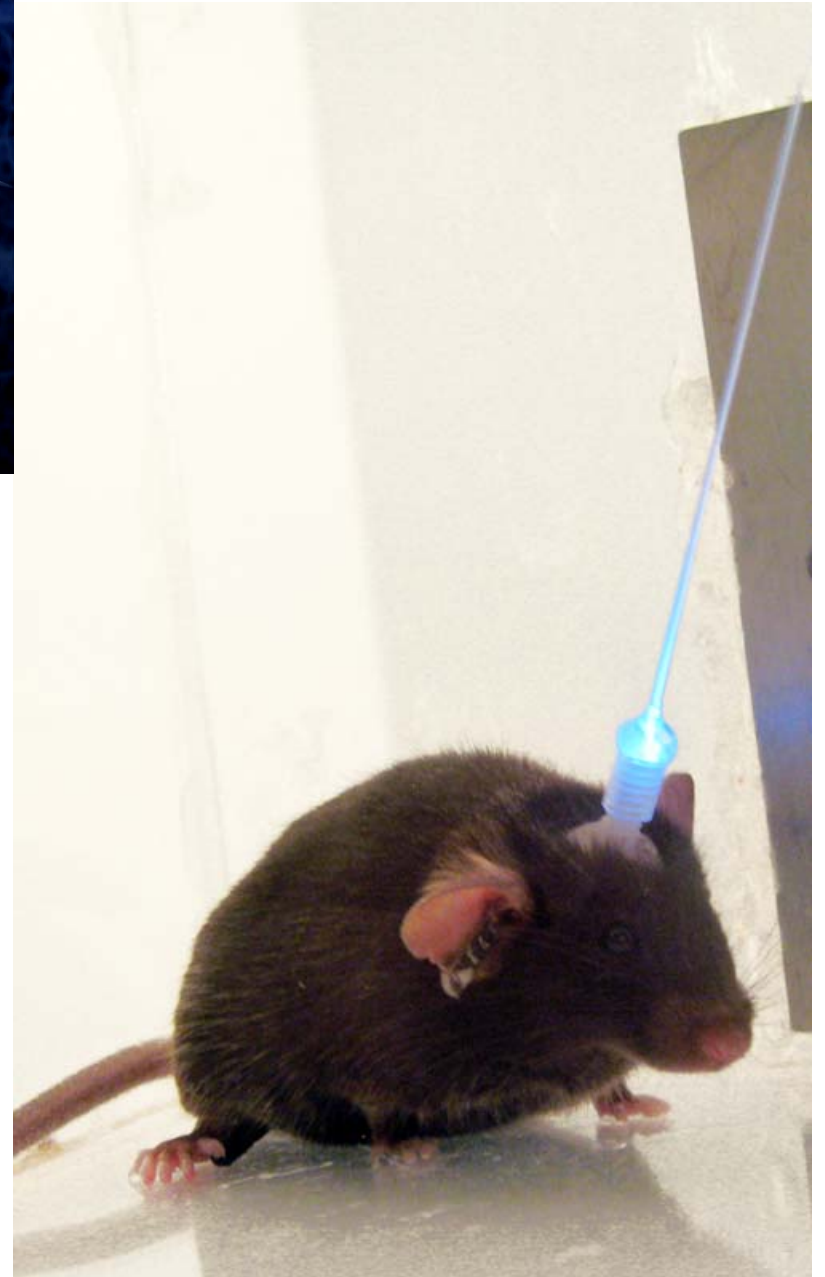
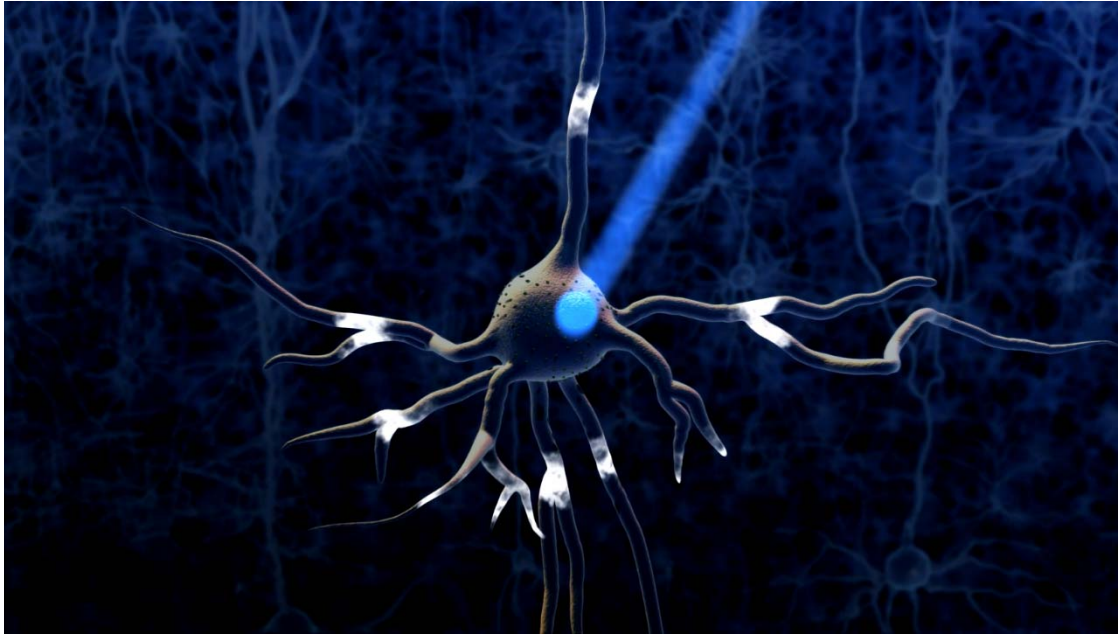


Prevedel\*, Yoon\*, et al. (2014) *Nature Methods* 11:727-730.



# **Control: neural activity**

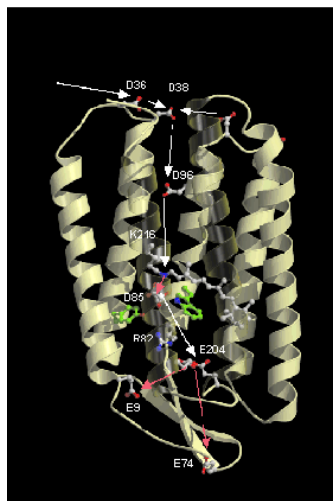
How can we control high-speed neural electrical activity with single-cell, millisecond-timescale precision?



## Bacteriorhodopsins: Light-driven proton pumps



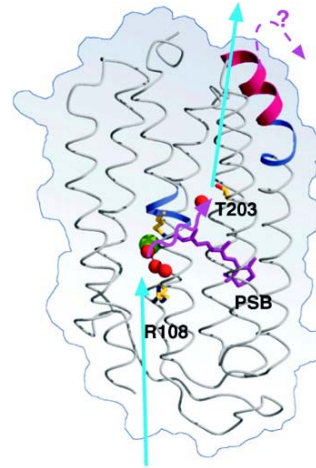
<http://www.genome.duke.edu/genomelife/2011/03/systems-under-stress/>



[http://www.biochem.mpg.de/523002/Protein\\_BR](http://www.biochem.mpg.de/523002/Protein_BR)

D. Oesterhelt and W. Stoeckenius (1971)  
Rhodopsin-like Protein from the Purple  
Membrane of *Halobacterium halobium*.  
*Nature New Biology* 233:149-152.

## Halorhodopsins: Light-driven chloride pumps



<http://www.sciencemag.org/content/288/5470/1390.full>

Matsuno-Yagi A, Mukohata Y (1977)  
Two possible roles of  
bacteriorhodopsin; a comparative  
study of strains of *Halobacterium*  
*halobium* differing in  
pigmentation. *Biochem Biophys*  
*Res Commun* 78:237-43.

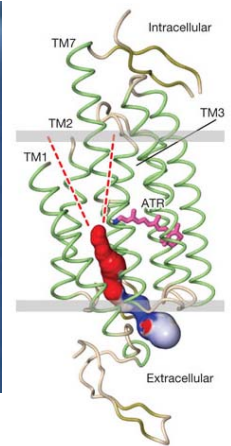
Matsuno-Yagi A, Mukohata Y (1980)  
ATP synthesis linked to  
light-dependent proton uptake in a  
mutant strain of *Halobacterium*  
lacking bacteriorhodopsin. *Arch*  
*Biochem Biophys*, 199:297-303.

Schobert B, Lanyi JK (1982)  
Halorhodopsin is a light-driven  
chloride pump. *J Biol Chem*,  
257:10306-13.

## Channelrhodopsins: Light-driven ion channels



<http://starcentral.mbl.edu/microscope/portal.php?pagetitle=assetfactsheet&imageid=3245>

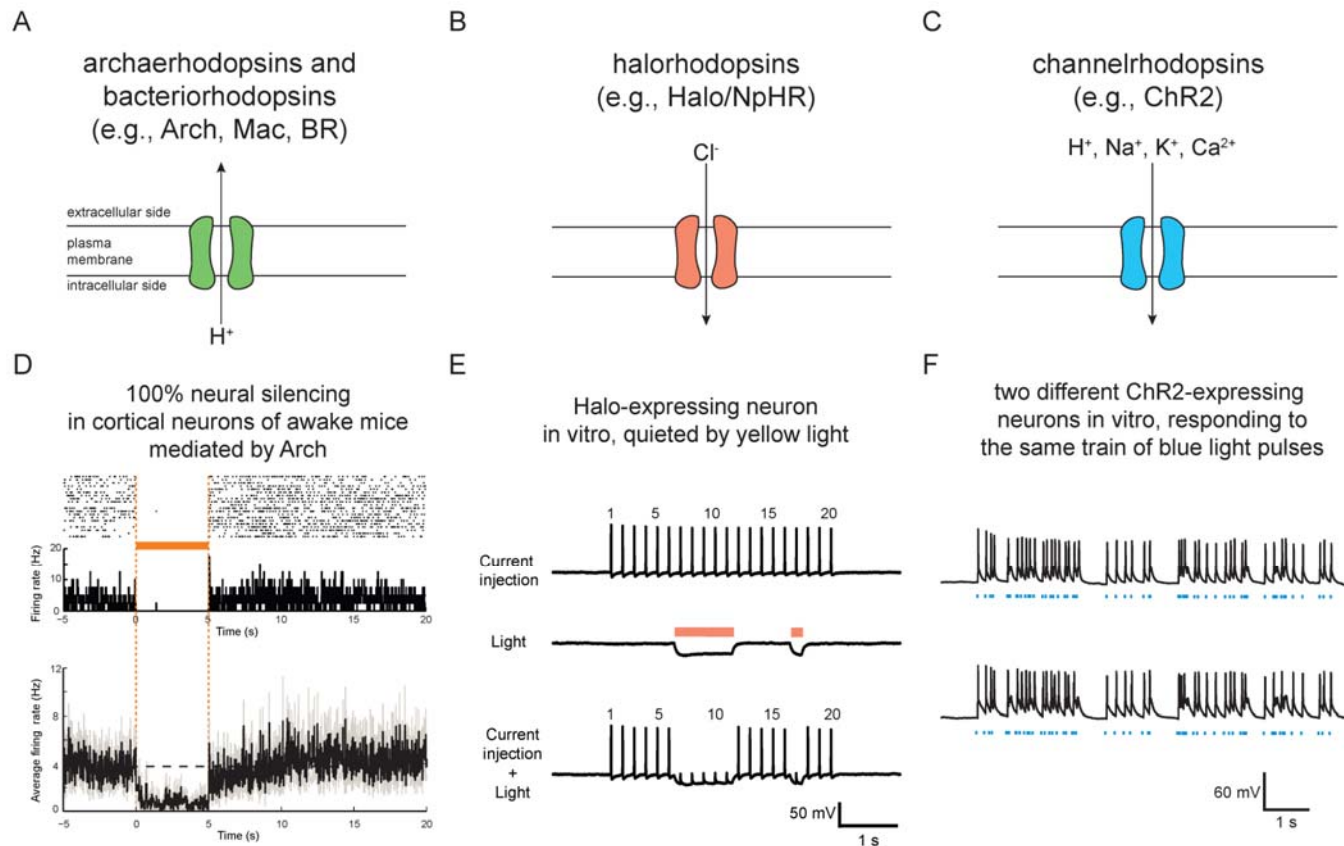


<http://www.nature.com/nature/journal/v482/n7385/full/nature10870.html>

Nagel G, Ollig D, Fuhrmann M, Kateriya S,  
Musti AM, Bamberg E, Hegemann P (2002)  
Channelrhodopsin-1: a light-gated proton  
channel in green algae. *Science*, 296:2395-8.

Nagel G, Szellas T, Huhn W, Kateriya S,  
Adeishvili N, Berthold P, Ollig D, Hegemann  
P, Bamberg E (2003) Channelrhodopsin-2, a  
directly light-gated cation-selective  
membrane channel. *Proc Natl Acad Sci U S A*,  
100:13940-5.

# Optogenetics: microbial opsins, seven-transmembrane proteins, binding endogenous all-trans-retinal

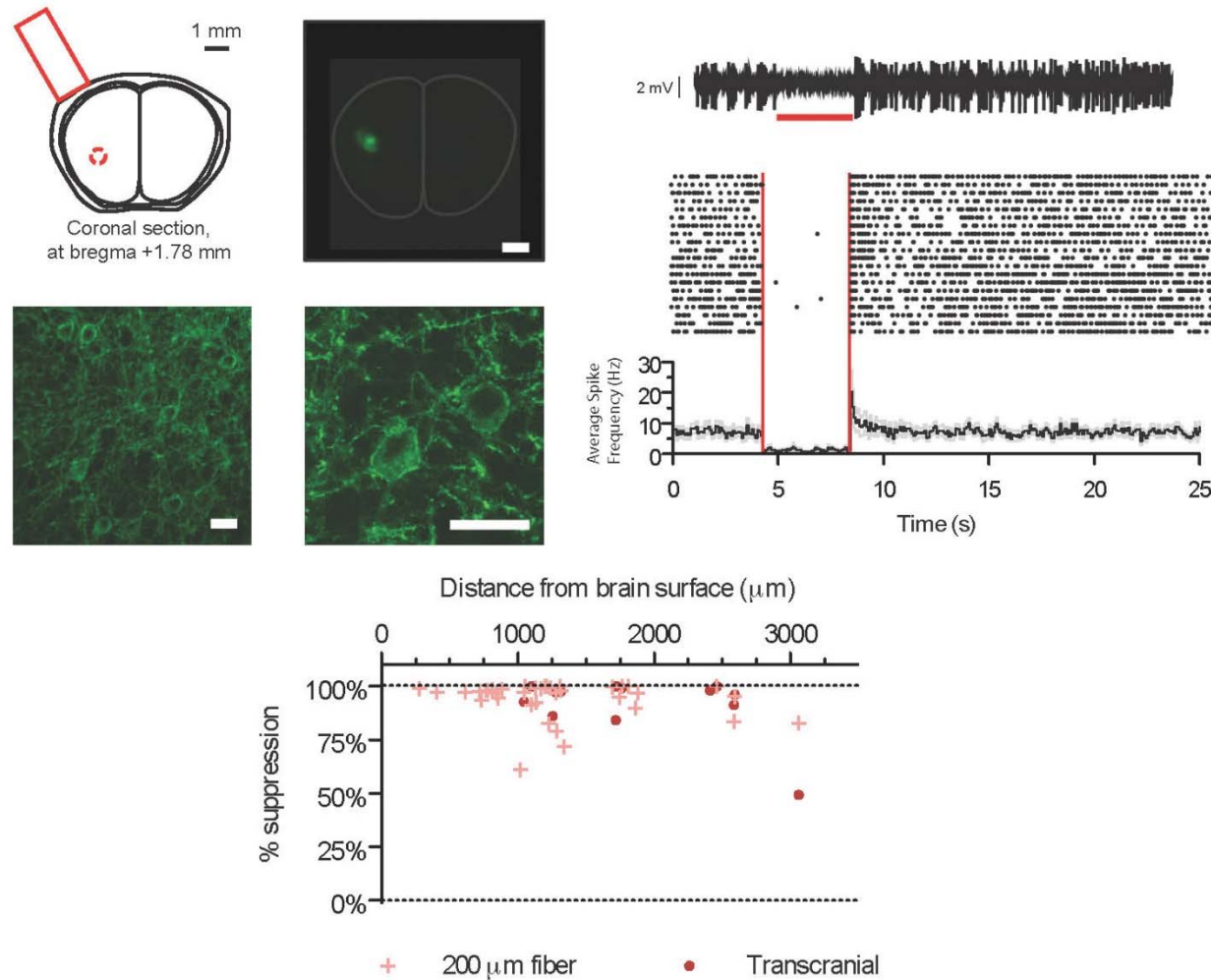


Boyden (2011) *Faculty of 1000 Biology Reports* 3:11.

Sequences, plasmids, links to DNA/virus providers, at <http://syntheticneurobiology.org/protocols>

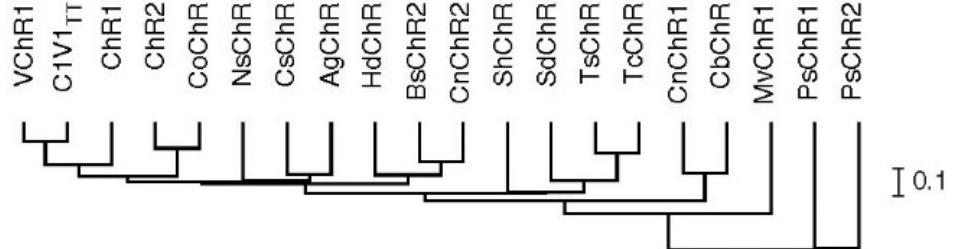
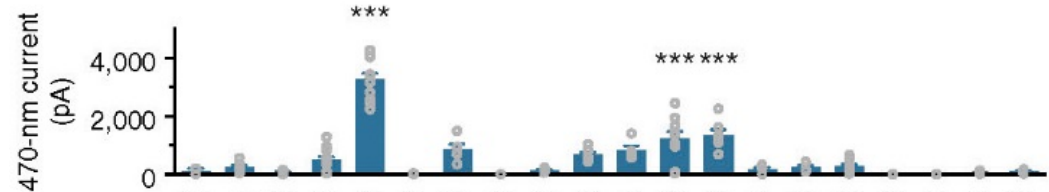
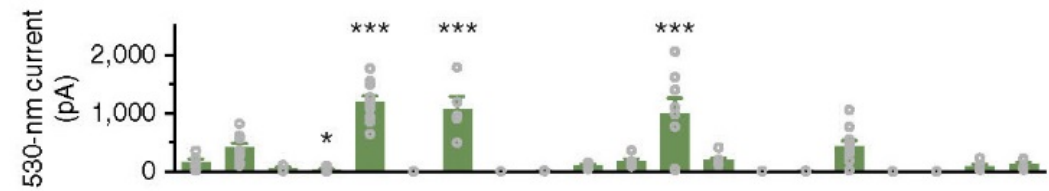
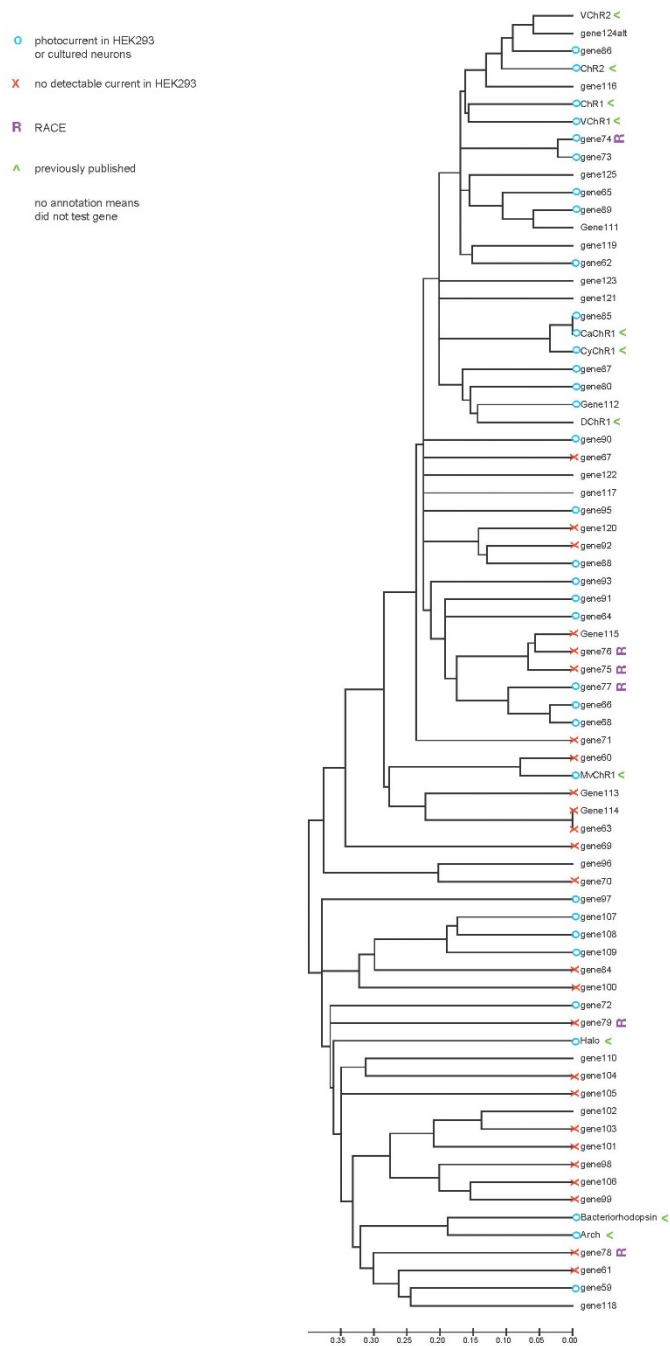


# Noninvasive optogenetic neural silencing: Jaws

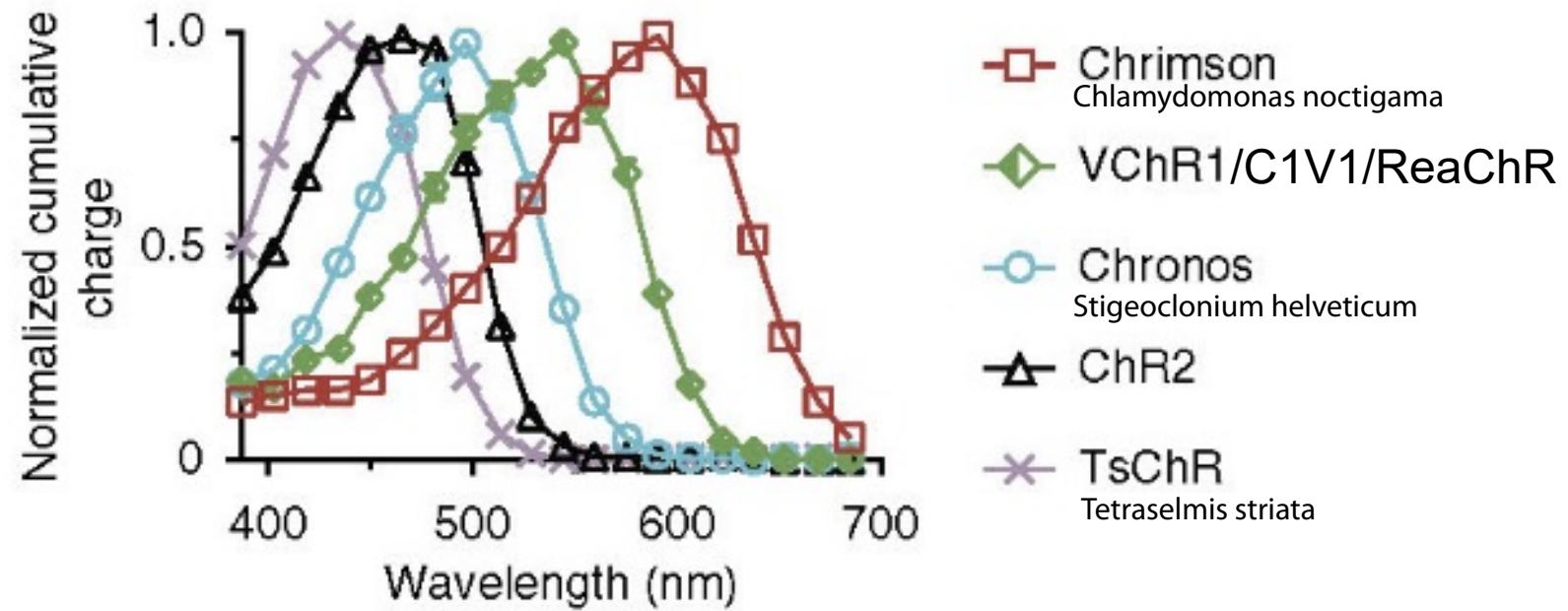


Chuong et al. (2014) *Nature Neuroscience* 17:1123-1129.



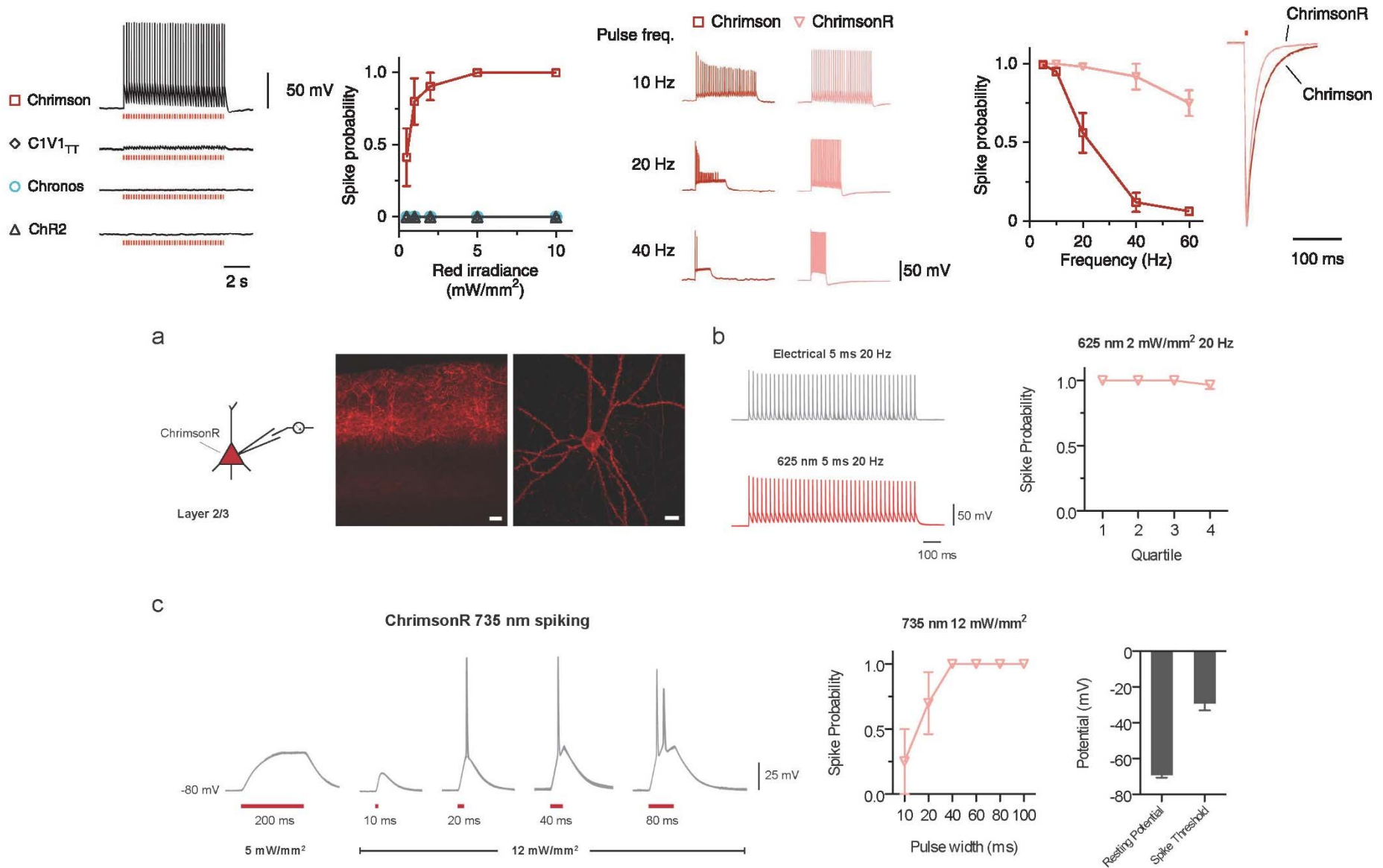


Klapoetke et al. (2014) *Nature Methods* 11:338–346.



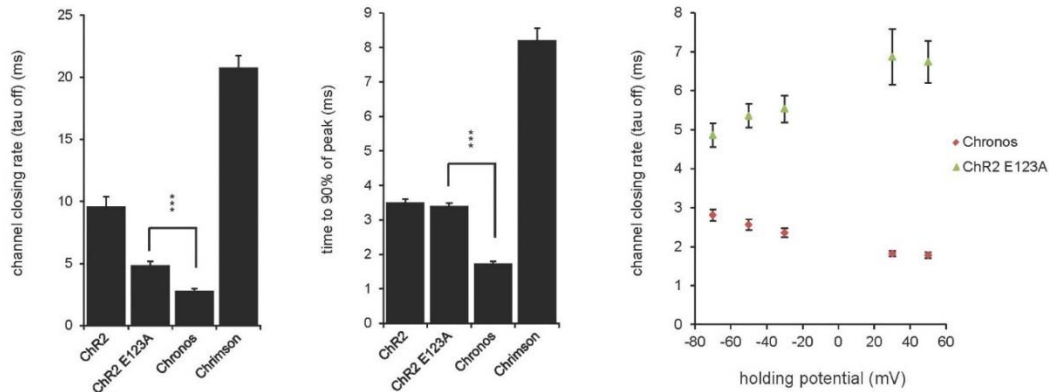
Klapoetke et al. (2014) *Nature Methods* 11:338–346.

# Chrimson: quasi-infrared neural stimulation

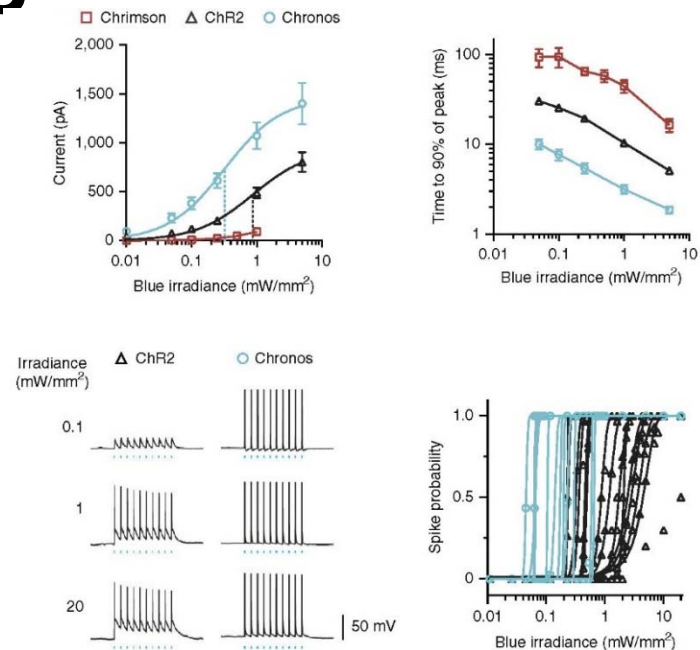


Klapoetke et al. (2014) *Nature Methods* 11:338–346.

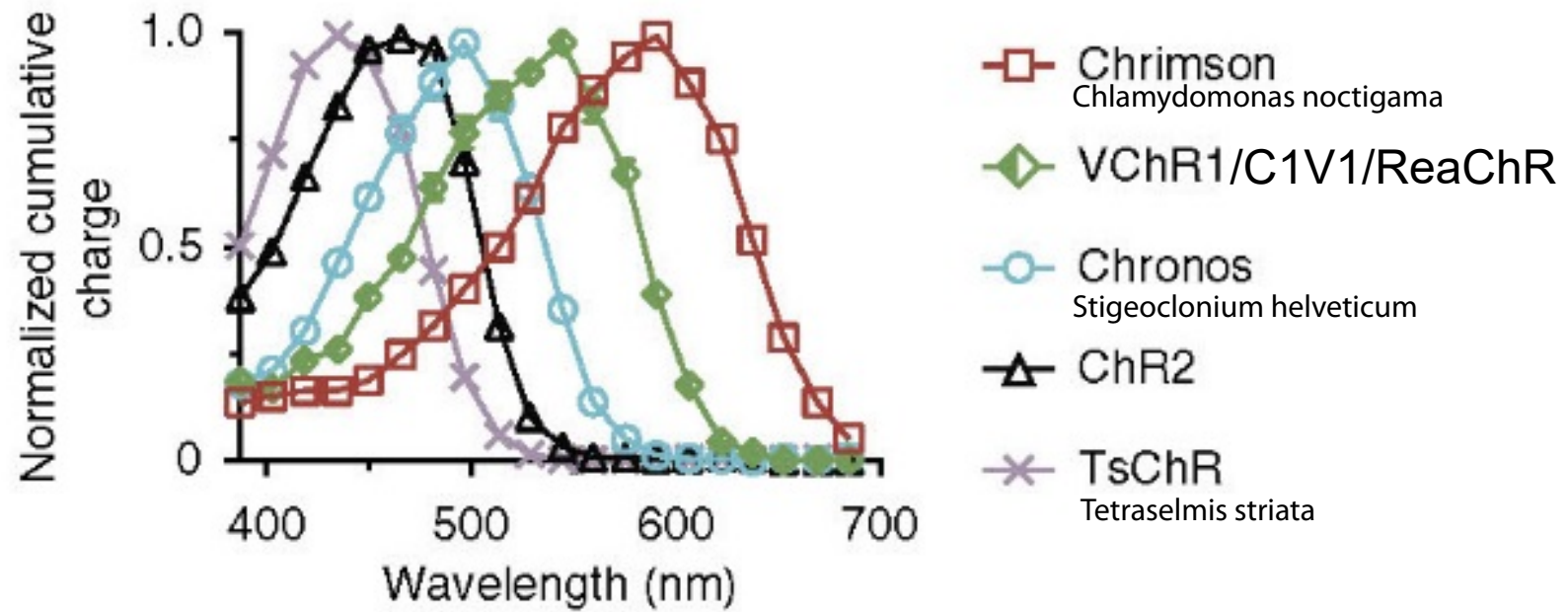
# Chronos: a very fast channelrhodopsin...



...that is also very light sensitive!



Klapoetke et al. (2014) *Nature Methods* 11:338–346.



Klapoetke et al. (2014) *Nature Methods* 11:338–346.



#### **Graduate Students, Postdocs, Staff**

Adam Marblestone  
Aimei Yang  
Amauche Emenari  
Andrew Payne  
Annabelle Singer (Georgia Tech)  
Asmamaw Wassie  
Brian Allen  
Christian Wentz  
Changyang Linghu  
Cynthia Smith  
Daniel Goodwin  
Daniel Martin-Alarcon  
David Rolnick  
Deblina Sarkar  
Demian Park  
Desiree Dudley  
Erica Jung  
Fei Chen  
Giovanni Talei Franzesi  
Grace Huynh  
Guangyu Xu (U.Mass.)  
Harbaljit Sohal  
Ho-Jun Suk  
Ingrid van Welie  
Ishan Gupta  
Jae-Byum Chang  
Jake Bernstein  
Jay Yu  
Jorg Scholvin  
Justin Kinney  
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# **Synthetic Neurobiology Group**

## **<http://syntheticneurobiology.org/>**

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#### **Conflict of Interest Statement**

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