

Rejuvenating Tithonus

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Tennyson tells us of Tithonus, who was endowed with immortality and was never permitted to perish, despite his wishes to that end. A biological parallel might be pluripotent stem cells, which can be indefinitely propagated *in vitro* in an uncommitted, self-renewing state, and can form all lineages in the mammalian body [1]. However, pluripotency is an inherently transient condition *in vivo*. Thus, pluripotent cells *in vitro*, such as embryonic stem cells (ESCs), continually show proclivities towards differentiation, similarly to Tithonus seeking the termination of his immortality [1]. To counteract this, several molecular actors, including the transcription factor *Prdm14*, within self-renewing pluripotent cells restrain forward developmental progression [2,3]. Four papers, including one in this issue of *EMBO reports*, depict mechanisms by which *Prdm14* serves as the elixir of youth for pluripotent stem cells [4–7], acting at the levels of developmental signalling and chromatin.

At the level of signalling, *Prdm14* desensitizes pluripotent cells to differentiation-inducing extrinsic signals. *In vivo*, the pluripotent cells of the blastocyst-stage epiblast—from which ESCs are derived—express *Fgf4*, which sustains adjacent extra-embryonic endoderm cells. However, endogenous fibroblast growth factor (FGF) signalling within ESC cultures *in vitro* promotes disassembly of the undifferentiated state and directs commitment to extra-embryonic or primitive streak lineages [8]. *Prdm14* safeguards ESCs from autocrine FGF signalling by binding to and down-regulating expression of *Fgfr* genes [5,7],

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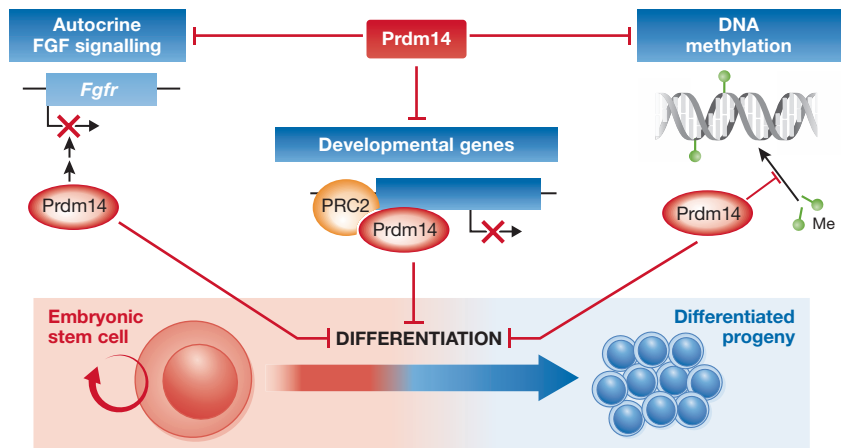


Fig 1 | *Prdm14* represses autocrine FGF signalling, occupies and suppresses multiple differentiation-associated genes and globally suppresses DNA methylation. This collectively restricts differentiation and entraps pluripotent cells in an uncommitted state. FGF, fibroblast growth factor.

apparently through the repression of distal enhancer elements that control *Fgfr* expression. Genetic removal of *Prdm14* renders ESCs more susceptible to the inductive effects of FGF signalling [5,7]. Yet, *Prdm14* acts more broadly than just counteracting the effects of FGF signalling, because epistatic experiments show that even if FGF/MAPK (mitogen-activated protein kinase) signalling is pharmacologically inhibited, *Prdm14* is still important for undifferentiated ESC self-renewal [5,7].

To identify the multiplicity of targets through which *Prdm14* might act, two studies used chromatin immunoprecipitation followed by sequencing to systematically chart genes across the genome bound by *Prdm14* [2,3]. By combining this *cis*-regulatory data with gene expression changes after perturbing *Prdm14* expression, two more studies found that *Prdm14* directly occupies a variety of regulatory genes that control germ layer formation and anterior–posterior patterning—for example, *Nodal*, *Snai1* and *Hhex*—and down-regulates their expression [5,7]. How

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does *Prdm14* prevent activation of these developmental programmes?

Prdm14 physically interacts with polycomb repressive complex 2 (PRC2; [4,7]), a chromatin-modifying complex that suppresses gene transcription and deposits the H3K27me3 histone modification. This suggests that *Prdm14* targets PRC2 to developmental genes to inhibit their expression. Indeed, transcriptional and chromatin state analyses have revealed that on genetic ablation of *Prdm14*, PRC2 and H3K27me3 fail to accumulate at archetypic *Prdm14* target genes [4,5]. Thus, in the absence of *Prdm14*, differentiation-associated transcriptional programmes are activated to some extent in undifferentiated ESC.

Reports also suggest that *Prdm14* forestalls differentiation by globally attenuating DNA methylation [5–7]. DNA methylation,

which is often associated with transcriptional repression, is required for differentiation but is dispensable for the self-renewal of undifferentiated cells [9]. Accordingly, various pluripotent cell types are generally characterized by genome-wide DNA hypomethylation [6]. *Prdm14* impedes DNA methylation in ESCs, apparently at a global level, as *Prdm14* deletion leads to a widespread increase of DNA methylation. *Prdm14* broadly represses DNA methylation by binding to and downregulating the expression of DNA methyltransferase genes, such as *Dnmt3a* and *Dnmt3b* [5–7].

Prdm14 thus pre-empts the precocious differentiation of pluripotent cells through a variety of molecular means. However, simply assigning *Prdm14* the role of a general antagonist of differentiation is an oversimplification. The particular lineage choices that *Prdm14* represses remain unclear, as pluripotency factors can often specifically inhibit particular prospective developmental fates whilst promoting others [1]. More broadly, the phenotypic importance of *Prdm14* remains controversial. In typical

***Prdm14* impedes DNA methylation in ESCs [...] at a global level [...] by binding to and downregulating the expression of DNA methyltransferase genes**

serum-containing culture conditions for ESC propagation, ESCs deteriorate if *Prdm14* is genetically ablated. Yet, within '2i' conditions, in which differentiation-inducing signalling is minimized, *Prdm14*^{−/−} ESCs still thrive despite modest upregulation of differentiation genes and a plethora of other defects [5,7]. Thus, if differentiation signals are withdrawn, ESCs become less reliant on *Prdm14*. Similarly, *Prdm14*^{−/−} mice do not show peri-implantation lethality—the typical consequence of disrupting pluripotent epiblast formation *in vivo*—rather, they have later deficiencies in germline development [10]. Perhaps *Prdm14* suppresses a battery of developmental genes by recruiting PRC2, but when extrinsic activating signals are minimized, these differentiation-associated genes cannot be brought into play, and *Prdm14* becomes less important. Equally, other pluripotency factors—for example, *Esrrb*—are dispensable for self-renewal if serum is withdrawn [11], suggesting that some pluripotency factors largely serve to dampen differentiation genes otherwise induced by serum, and thus serve in a more ancillary capacity. In any case, development *in vivo* is accompanied by *Prdm14* downregulation [10] and persistent *Prdm14* expression perturbs proper differentiation [5]. How developmental morphogens suppress *Prdm14* to support differentiation remains a pertinent question, the answer to which is how, at last, Tithonus might be laid to rest.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

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