Treatment of 4T1 Metastatic Breast Cancer with Combined Hypofractionated Irradiation and Autologous T-Cell Infusion

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INTRODUCTION

The ability of radiation to induce remission of tumors is dependent on the injury or death of tumor cells themselves and/or the stromal and vascular cells within the tumors (1–3). A combination of DNA damage, activation of apoptosis and production of reactive oxygen species contribute to tumor remissions (1–3). In addition, radiation can be used to enhance systemic T-cell antitumor immunity that can improve therapeutic efficacy (4–23). Recent studies have shown that the ability of a single dose of radiation (20 Gy) to slow the growth of primary melanoma tumors is dependent on immune cells, since the slowing observed in wild-type mice failed to occur in immunodeficient nude mice, and slowing was abrogated by depleting the CD8+ T cells of the tumor-bearing mice with monoclonal antibodies (4, 5). Multiple smaller doses of radiation instead of the single dose were ineffective in slowing tumor growth, and chemotherapy administered after the single dose interfered with tumor regression and the associated immune response (4). Additional studies showed that radiation exposure increased tumor immunogenicity, stimulated antigen-presenting cells and promoted migration and entry of T cells into tumors (6–23).

Tumor irradiation has been combined with immunotherapy such as transduction of tumor cells with DNA-encoding immunogenic peptides, stimulatory ligands or chemokines (4, 5). The combined approach, which includes injections of dendritic cells, Flt-3 ligand or anti-CTLA4 monoclonal antibodies after radiotherapy, has been shown to induce systemic immunity in mice such that tumor growth at distant sites is slowed (12–17). Durable complete remissions with weakly immunogenic tumors were not achieved unless the tumors were small (<1 cm) and nonmetastatic (12–17).

Advances in the use of confocal radiation beams that are targeted to a tumor in 3 dimensions minimize irradiation to adjacent normal tissues [stereotactic body radiation therapy (SBRT)] and allow for administration of single doses as high as 30 Gy or up to 3 daily doses of 20 Gy each for a total of 60 Gy (24, 25). The efficacy of SBRT to induce solid tumor remission has been shown to be superior to that of fractionated irradiation with multiple small doses administered over several weeks (24, 25).

In the current study, we compared the efficacy of high-dose hypofractionated irradiation (3 × 20 Gy) alone to the...
combination of irradiation and autologous T-cell infusion for the treatment of metastatic 4T1 breast tumors in mice. Previous studies have shown that infusion of autologous T cells expanded ex vivo from tumor-infiltrating cells (TILs) or transfected with DNA constructs that encode T-cell antigen receptors that recognize tumor antigens can induce complete remission in patients with melanoma and lymphoid leukemias (26–28). The T-cell infusions were most effective after conditioning with lymphodepletive agents (26–28). In addition, the antitumor activity of vaccination with irradiated mouse colon tumor cells and adjuvant is markedly enhanced by autologous T-cell infusion after lymphodepletive total-body irradiation (29). The results of the current study show that the combination of local tumor irradiation and autologous T-cell infusion after lymphodepletion is more effective than irradiation alone.

MATERIALS AND METHODS

Animals
BALB/c (H-2b) wild-type female mice were purchased from Jackson Laboratories (Bar Harbor, ME). The Stanford University Committee on Animal Welfare (Administration Panel of Laboratory Animal Care) approved all mouse protocols used in this study.

Cell Lines
The 4T1 cell line was obtained from ATCC®. The 4T1-LUC/GFP cell line was lentivirally transduced (30–32).

Irradiation
Irradiation was performed with a Philips X-ray unit (200 kV, 10 mA; Philips Electronic Instruments Inc., Rahway, NJ) at a rate of 84 cGy/min with a 0.5 mm copper filter. For local tumor irradiation (LTI), unanesthetized mice were placed in lead jigs through which established tumors in the hindquarter were protruded for irradiation to an area of approximately 2 cm diameter (33).

Cell Preparation, Splenectomy and Collection of T Cells for Autologous Transplantation
Single cell suspensions from the spleen were enriched for T cells using anti-Thy1.2 columns according to previously described procedures (29). Collected T cells were cryopreserved with 10% dimethyl sulfoxide (DMSO) and frozen in liquid nitrogen.

In Vivo Bioluminescence Imaging (BLI)
In vivo BLI was performed according to the method of Edinger et al. (32).

RESULTS

Local Tumor Irradiation Alone or in Combination with Cyclophosphamide Fails to Eradicate Distant 4T1 Metastases
We studied the effect of high-dose irradiation on 4T1 breast cancer that metastasizes to the lungs after subcutaneous injection in BALB/c mice (34, 35). Figure 1A shows the growth of the 4T1 tumor transduced with the luciferase gene using BLI after subcutaneous injection into the right hindquarter. All mice died of tumor progression within 55 days whether or not tumor cells were transduced with the luciferase gene (Fig. 1A) and growth was similar to that reported for orthotopic injections (35). Both BLI signals above the diaphragm and histopathological analysis at day 21 showed that tumors had spread to the lungs by that time (Fig. 1B and C). The enlarged area of BLI signal below the diaphragm is associated with the growth of the primary tumor with subcutaneous local spread. When a single radiation dose of 30 Gy was administered to day 14 tumors, slowing of tumor growth was observed without development of complete remission (data not shown). However, 3 daily radiation doses of 20 Gy each resulted in complete remission in 2 out of 5 mice (Fig. 1A and D), and the mice survived for more than 100 days. After tumor remission, the irradiated skin area showed hair loss, scarring and contraction without ulceration.

In further studies, tumors were allowed to grow for 21 days before treatment. A single radiation dose of 30 Gy to primary tumors showed only modest slowing of local tumor growth, and had little impact on metastatic spread or survival (data not shown). When the subcutaneous tumors were irradiated with 3 daily doses of 20 Gy each, the primary tumors showed marked regression, but still spread to distant sites as shown by BLI. Seven of eight mice died by day 84, with 1 of 8 in remission (Fig. 1D). Next, we tried to augment radiotherapy with cyclophosphamide (CY) treatment. When a single intraperitoneal dose of 500 mg/kg of cyclophosphamide was administered after the third dose of local tumor irradiation, tumor growth slowed, but 6 of 8 mice died by day 84 and 1 relapsed at day 98 (Fig. 1E). This dose of cyclophosphamide depleted T cells in the spleen almost eightfold in nontumor-bearing mice. The mean ± SE absolute number of T cells in spleens of untreated animals was 29 ± 5 × 10⁶ and was reduced to 9 ± 2 × 10⁶ and 3.9 ± 0.7 × 10⁶ at days 3 and 7, respectively after cyclophosphamide injection.

Local Tumor Irradiation in Combination with Cyclophosphamide and Autologous T-Cell Infusion Can Eradicate Distant 4T1 Metastases

Based on the reported effect of lymphodepletive regimens to enhance T-cell immunity when given prior to T-cell infusion (27, 28), a group of tumor-bearing mice were treated with local tumor irradiation followed by collection of T cells within 24 h (Fig. 2). To collect total T cells, splenectomy was performed, and T cells were enriched using an anti-Thy1.2 mAb column and cryopreserved. Just after T-cell collection, the mice were given a single dose of 500 mg/kg cyclophosphamide. Forty-eight hours later, when the number of endogenous cells was declining, 2.5 × 10⁶ T cells were thawed and infused intravenously. In this group, 4 of 7 mice showed no evidence of primary and
metastatic tumor up to day 180. Survival of the group that had their local tumor irradiated, and were given cyclophosphamide and T cells was significantly increased according to the log rank test \( (P, 0.01) \) compared to the group that had their local tumor irradiated and were given cyclophosphamide without T cells (Fig. 2). Thus, the combination of local tumor irradiation, cyclophosphamide and T-cell therapy was more effective than either local tumor irradiation alone or local tumor irradiation in combination with cyclophosphamide.

**T-Cell Subsets in Autologous Infusions**

The T-cell subsets in the spleens of tumor bearing mice that were used for the autologous infusions were analyzed by immunofluorescent staining. Figure 3 compares the two-color flow cytometric analyses of spleen cells from normal mice, tumor-bearing mice at day 21 after tumor inoculation, and tumor-bearing mice immediately after 3 doses of local tumor irradiation on days 21, 22 and 23. Harvesting of T cells and enrichment using anti-Thy 1.2 columns were performed on day 23. Whereas the percentage of CD4\(^+\) and CD8\(^+\) T cells among mononuclear cells in the representative normal spleen was 21% and 10%, respectively (Fig. 3A), the percentage was reduced to about 7% and 2%, respectively in the unirradiated tumor-bearing spleen (Fig. 3B) or irradiated tumor-bearing spleen (Fig. 3C).

The reduction in the percentage of CD4\(^+\) and CD8\(^+\) T cells was due to the increase in Gr-1\(^+\)CD11b\(^+\) cells, from about 2% in normal mice to about 70% in the tumor-bearing mice (Fig. 3A–C). The latter cells have been identified previously as myeloid derived suppressor cells (MDSCs) (37). Figure 3D shows that on day 21 there was a massive infiltration of the MDSCs in the spleen, and the mean absolute number rose from about 2 \( \times 10^6 \) cells per spleen to about 300 \( \times 10^6 \) cells per spleen. Nevertheless, the mean absolute numbers of CD4\(^+\) and CD8\(^+\) T cells in the normal spleens and tumor-bearing spleens were similar to one another with about 9 \( \times 10^6 \) CD8\(^+\) T cells and about 18–23 \( \times 10^6 \) CD4\(^+\) T cells, respectively.

Among the CD4\(^+\) T cells, the percentages of CD4\(^+\)CD25\(^+\) Tregs were similar in the normal, unirradiated tumor-bearing spleen (Fig. 3B) or irradiated tumor-bearing spleen (Fig. 3C).

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**FIG. 1.** Effect of local tumor irradiation alone or in combination with cyclophosphamide on growth of 4T1 breast tumors. Panel A: Survival of BALB/C mice given subcutaneous injections of \( 1 \times 10^4 \) 4T1 wild-type tumor cells and 4T1 luc\(^+\) tumor cells compared to survival of 4T1 WT or 4T1 luc\(^+\) tumor-bearing mice given 60 Gy (3 daily doses of 20 Gy each) local tumor irradiation (LTI) on days 14, 15 and 16. There were five mice in each group. Panel B: Bioluminescence imaging (BLI) of mice at serial time points after injection of 4T1 luc\(^+\) tumor cells. Note signals above diaphragm at days 21 and 28. Panel C: Representative tissue section (H&E, 100x magnification) of lung showing tumor cell cluster (arrows) in untreated mouse. Panel D: BLI of mice given 4T1 luc\(^+\) tumor cells after 60 Gy (3 daily doses of 20 Gy each) local tumor irradiation on days 14, 15 and 16 or on days 21, 22 and 23. Blank areas indicate death of mice. Panel E: BLI of mice given 4T1 luc\(^+\) cells, 60 Gy (3 daily doses of 20 Gy each) local tumor irradiation and intraperitoneal injection of cyclophosphamide (500 mg/kg) on day 23. There were 5–8 mice in each group in D and E.
16–20%. The percentage of CD8\(^+\) T cells that expressed the PD-1\(^+\) Tim-3\(^+\) phenotype, associated with “exhaustion” (38), was less than 1% in all three types of spleen cells. The percentage of NKT cells among T cells was similar in normal and tumor-bearing mice (data not shown). Thus, with the exception of the increase in PD-1 on CD4\(^+\)CD25\(^+\) cells, the phenotypic profiles and absolute numbers of CD4\(^+\) and CD8\(^+\) T cells in tumor-bearing spleens were similar to normal.

**DISCUSSION**

We studied the 4T1 breast cancer model to determine whether high-dose tumor irradiation can effectively treat both primary tumors and metastases in wild-type mice. Subcutaneous injection of 4T1 tumor cells results in local tumor growth, followed by the development of metastases in the lungs (34, 35). The kinetics of this spread were confirmed in the current study by BLI using luciferase-labeled 4T1 tumor cells. Lung metastases were identified at day 21 by both imaging and histopathology. Accordingly, radiation was administered to the tumors starting on day 21. The 4T1 primary tumors were not controlled by a single radiation dose of 30 Gy, but rather required 3 daily radiation doses of 20 Gy each to induce complete remissions in day 14 tumors. However, even the 3 \(\times\) 20 Gy tumor irradiation, which cured the primary tumors failed to control metastases in most mice and almost all mice died by day 84.

Since autologous T-cell infusion after lymphodepletive conditioning has been reported to be effective in treating melanoma (27, 28), we used this strategy to treat 4T1 metastases. We hypothesized that local tumor irradiation would activate T cells in the tumor and/or local lymph nodes, and that the activated T cells would disseminate in the lymphoid tissues. Immediately after tumor irradiation on days 21–23, T cells were harvested and cryopreserved. A

![FIG. 2. T-cell therapy facilitates complete remissions of 4T1 metastases when used in combination with radiotherapy and chemotherapy. BALB/C mice were given 4T1 luc\(^+\) cells and local tumor irradiation and cyclophosphamide as described in Fig 1E and splenic Thy 1.2\(^+\) T cells were harvested just before cyclophosphamide injection on day 23. T cells were cryopreserved, then thawed and injected (2.5 \(\times\) 10\(^6\)) intravenous 48 h after injection of cyclophosphamide. BLI at serial time points is shown, as well as survival of untreated and experimental groups. There were significant differences in survival between groups given local tumor irradiation alone or local tumor irradiation and cyclophosphamide vs. local tumor irradiation and cyclophosphamide and T cells (\(P < 0.05\), but not between groups given local tumor irradiation and cyclophosphamide vs. local tumor irradiation alone (\(P > 0.05\)). There were 5–7 mice in each group.](image1)

![FIG. 3. Analysis of T-cell subsets in spleens from normal and tumor-bearing mice. Panel A: Single cell suspensions were prepared from normal spleens and analyzed for expression of CD25, PD-1 and Tim-3 on CD4\(^+\) and CD8\(^+\) T cells and for MDSCs (CD11b\(^+\)Gr-1\(^+\)). Percentages of each subset in boxes on representative two color analysis panels are shown and arrows identify gated subsets. Staining for CD11b, Gr-1, CD4 and CD8 used a mononuclear cell gate. Panel B shows the same analysis of spleens from tumor-bearing mice at day 21 after tumor induction. Panel C shows subset analysis just after local tumor irradiation 3 \(\times\) 20 Gy (days 21, 22 and 23). Panel D: Absolute numbers of MDSCs in the spleens of normal and tumor-bearing (day 21) mice. Panel E: Absolute numbers of CD4\(^+\) and CD8\(^+\) T cells in normal and tumor-bearing mice (day 21).](image2)
A Normal spleen

B Tumor-bearing (day 21); no LTI

C Tumor-bearing (day 23); LTI at day 21

D Absolute number of MDSCs in spleen (×10^6 cells)

E Absolute number of T cells in spleen (×10^6 cells)


