Lecture 9: Introduction to Genomics

Serena Yeung

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Announcements

- Upcoming deadlines:
 - Project proposal was due last Friday
 - A2 due next Wed Oct 21



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Some biology basics: starting from DNA

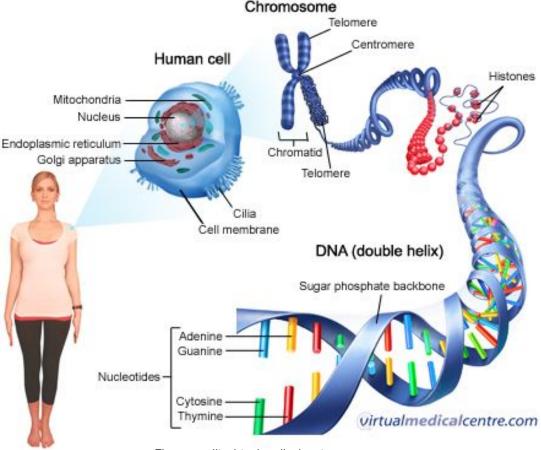


Figure credit: virtualmedicalcentre.com

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Some biology basics: starting from DNA

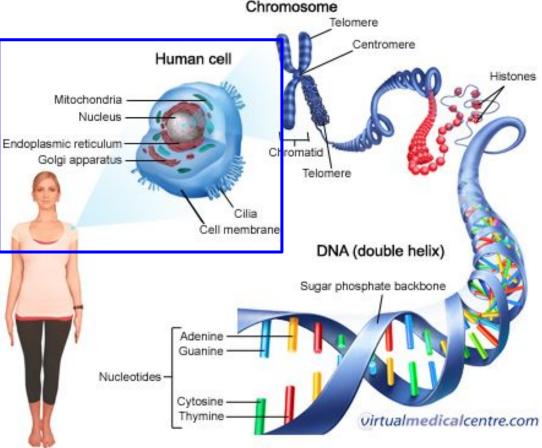


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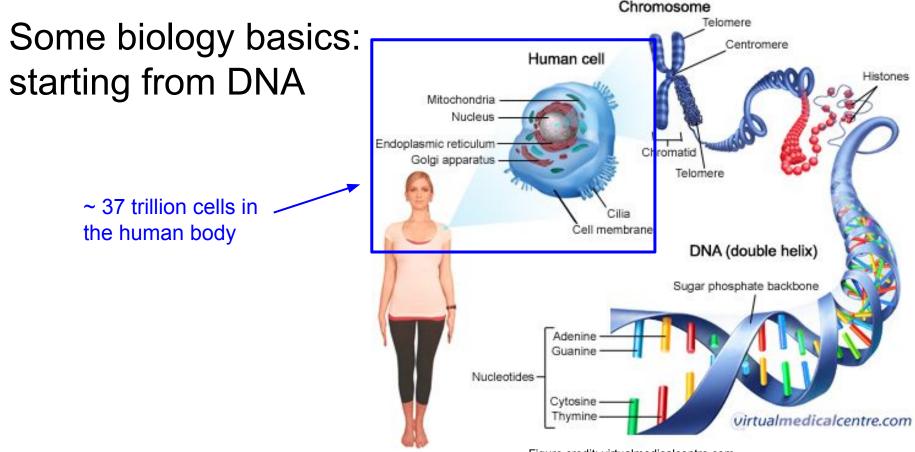


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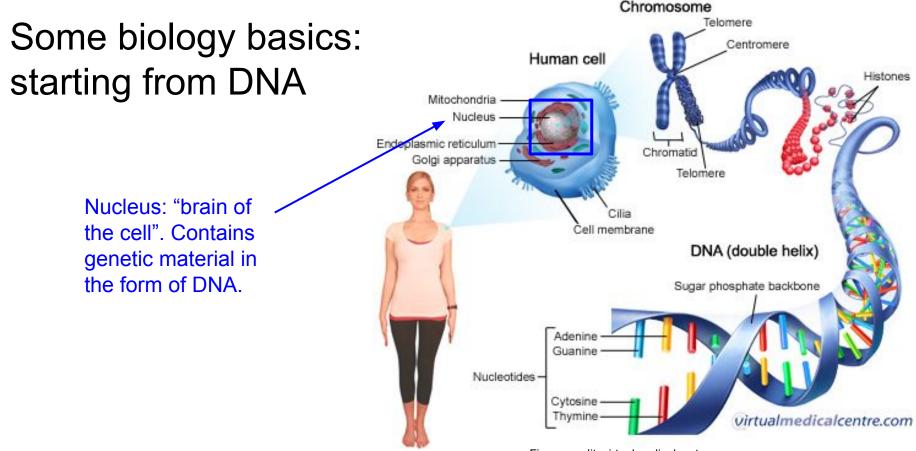


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Some biology basics: starting from DNA

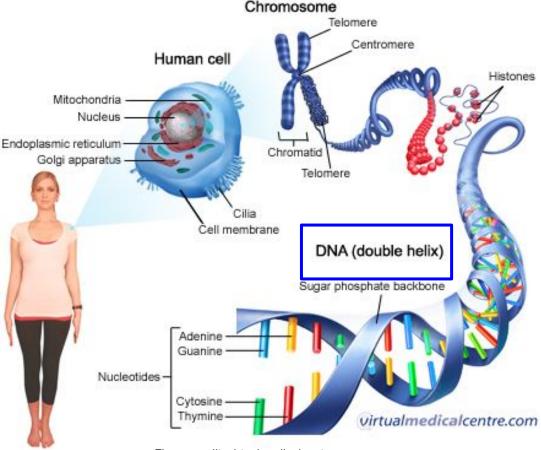
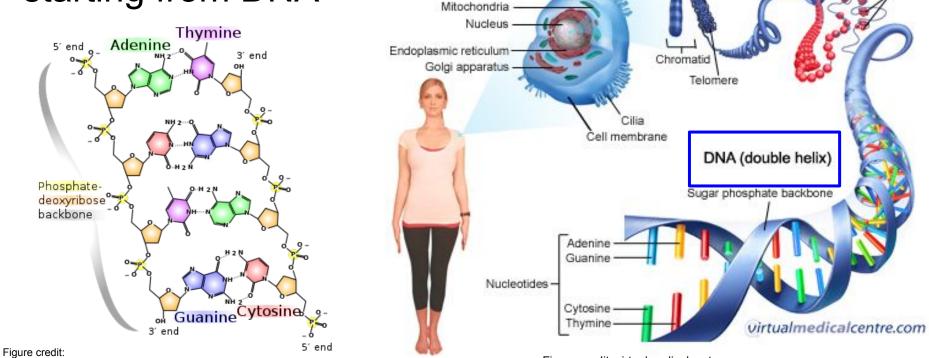


Figure credit: virtualmedicalcentre.com

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Some biology basics: starting from DNA



https://en.wikipedia.org/wiki/Nucleobase#/media/File:DNA_chemical_structure.svg

Figure credit: virtualmedicalcentre.com

Chromosome

Human cell

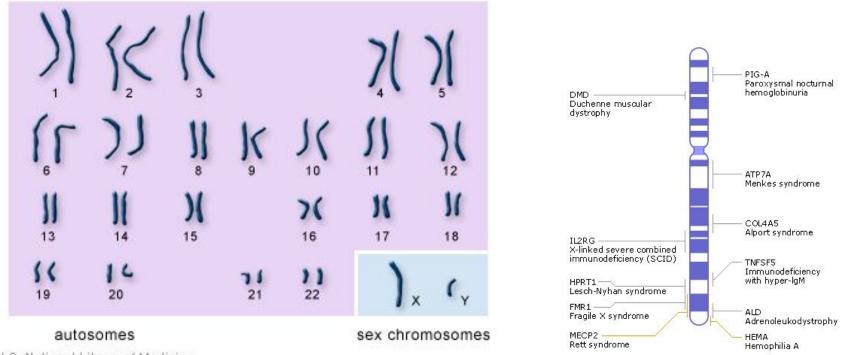
Telomere

Centromere

Histones

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Figure credit: https://ghr.nlm.nih.gov/primer/illustrations/chromosomes.jpg

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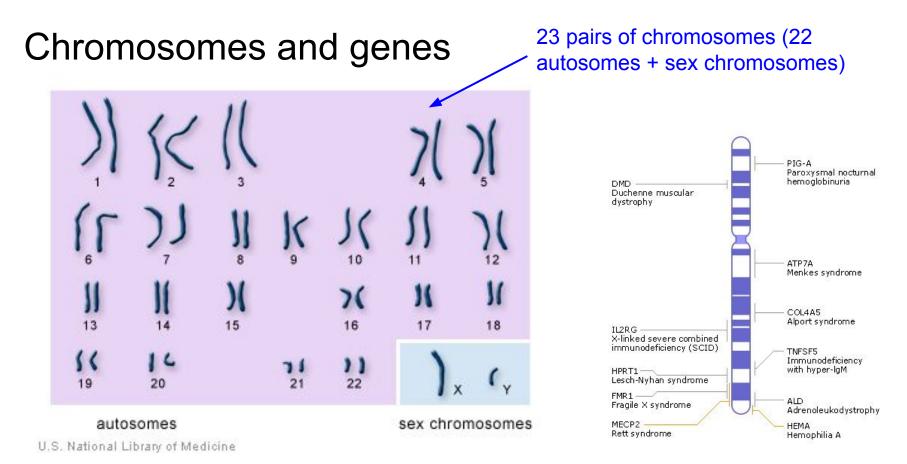


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Genes: segments of DNA within chromosomes

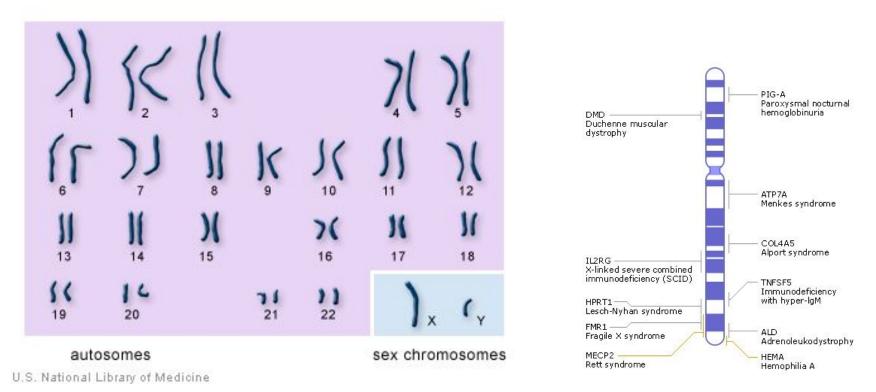
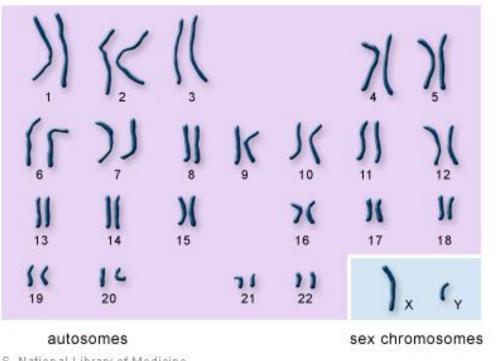


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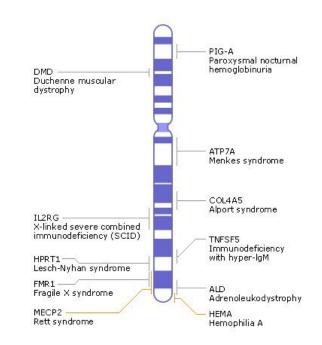
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Genes: segments of DNA within chromosomes

Genes provide code for proteins



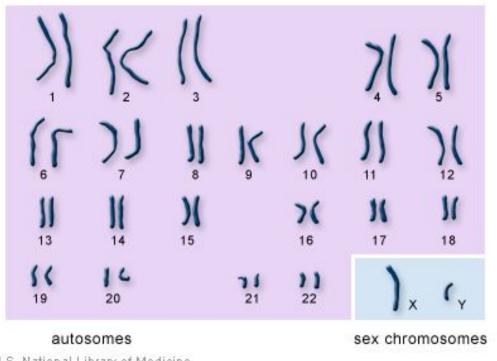
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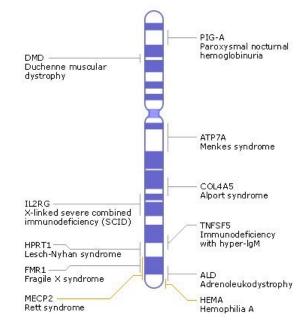
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Genes: segments of DNA within chromosomes

Genes provide code for proteins But 99% of genes are "non-coding!"



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DNA replication and transcription

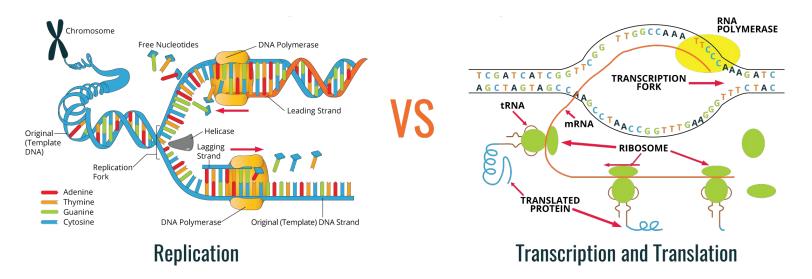
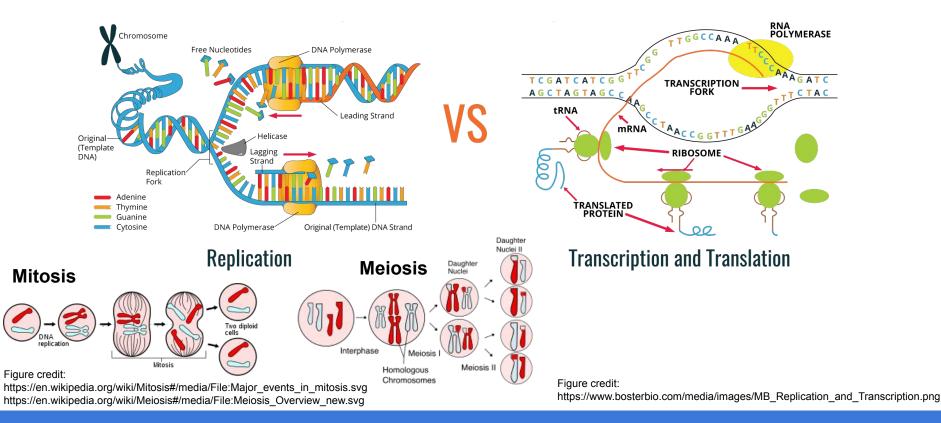


Figure credit: https://www.bosterbio.com/media/images/MB_Replication_and_Transcription.png

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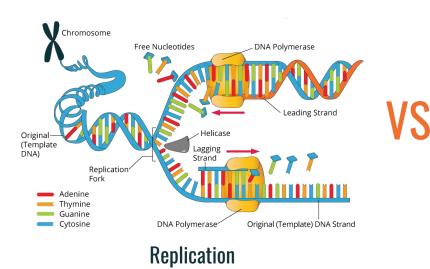
DNA replication and transcription



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DNA replication and transcription



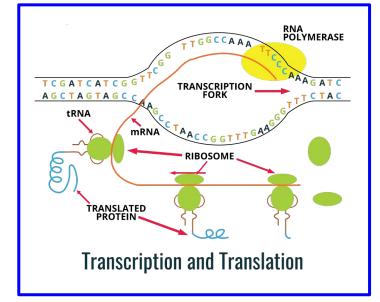


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Transcription and translation

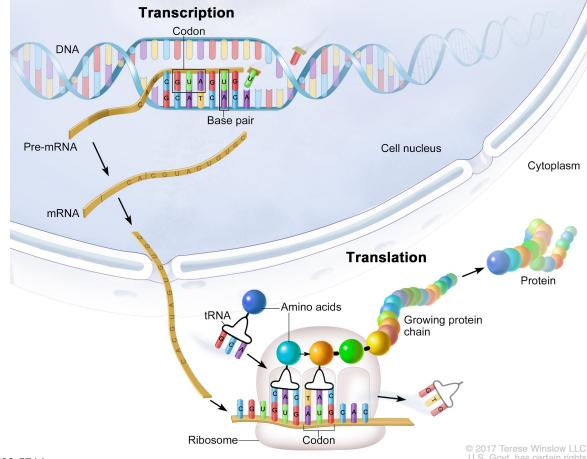


Figure credit: https://www.cancer.gov/images/cdr/live/CDR761782-571.jpg

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Transcription and translation

Transcription: DNA -> RNA

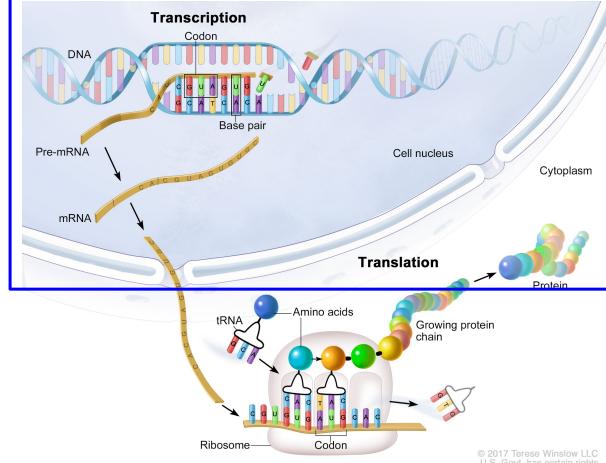


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Transcription and translation

Transcription: DNA -> RNA

Translation: RNA -> Protein

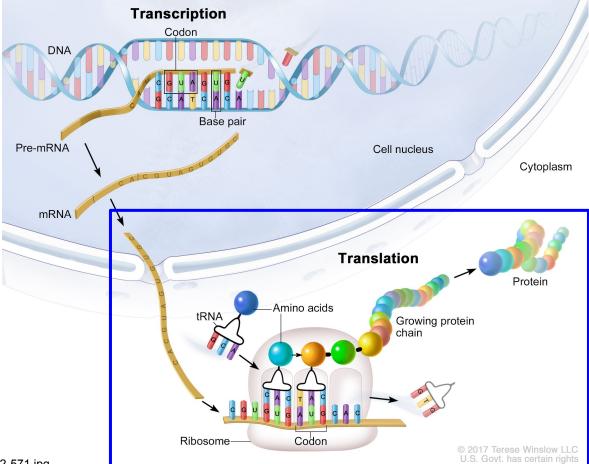
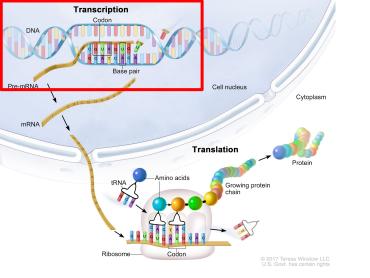


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DNA -> Pre-mRNA

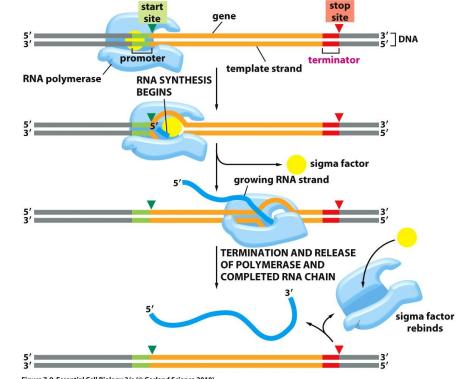


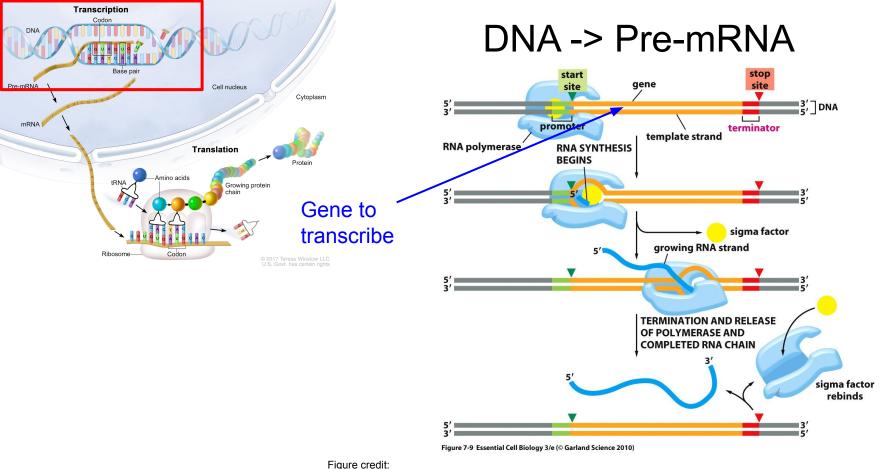
Figure 7-9 Essential Cell Biology 3/e (© Garland Science 2010)

http://u18439936.onlinehome-server.com/craig.milgrim/Bio230/Outline/ECBFigures_Tables/Chapter_7/FigureJPGs/figure_07_09.jpg

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Figure credit:



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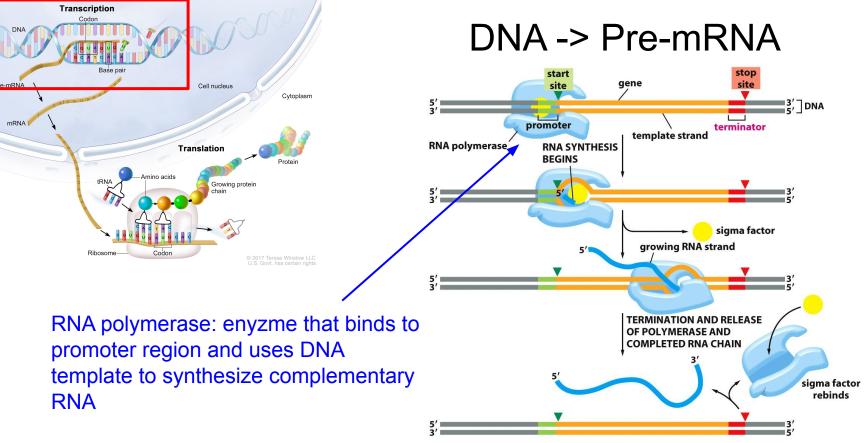


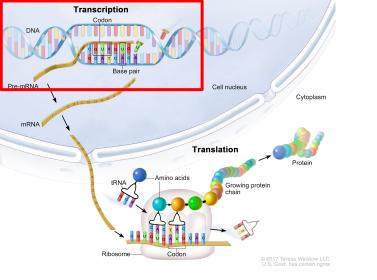
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Figure credit:



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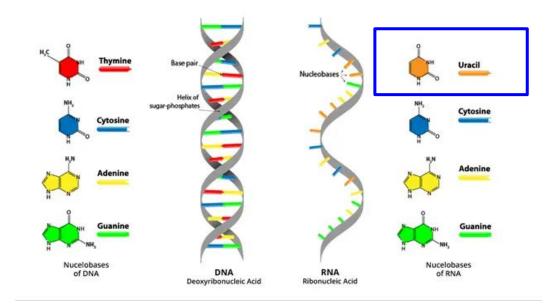
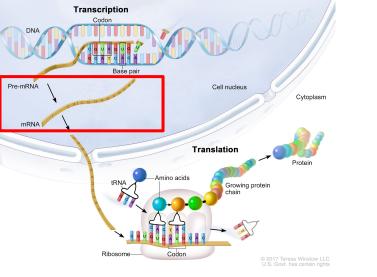


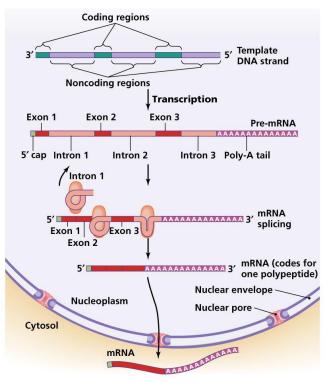
Figure credit: https://cdn.technologynetworks.com/tn/images/thumbs/webp/640_360/what-are-the-key-differences-between-dna-and-rna-296719.webp?v=9503516

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Pre-mRNA -> mRNA



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Figure credit: http://academic.pgcc.edu/~kroberts/Lecture/Chapter%207/transcription.html

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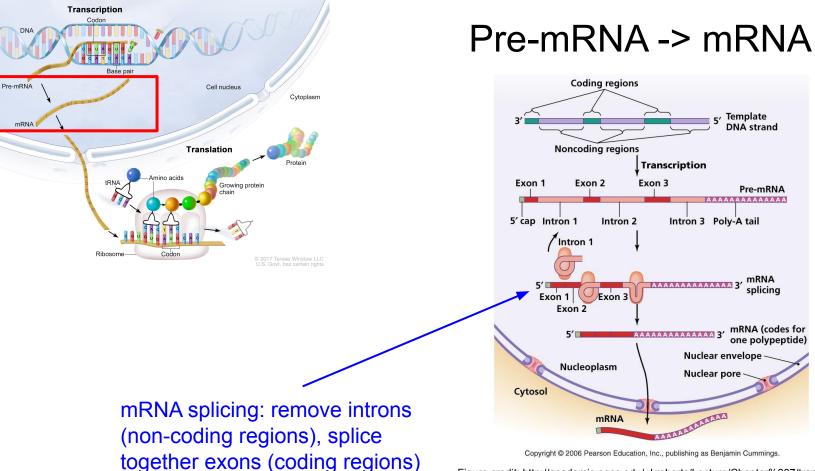
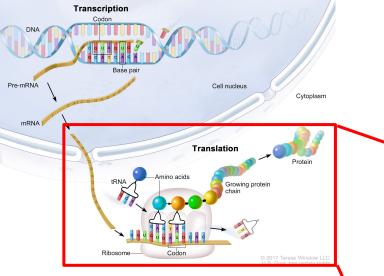


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mRNA -> Proteins

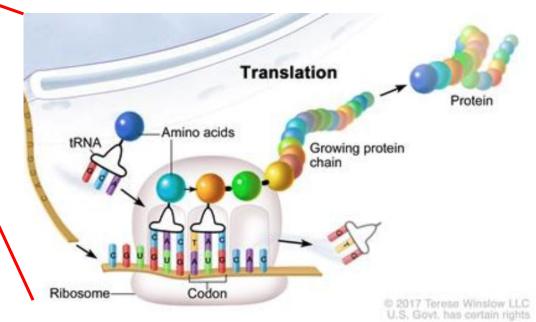


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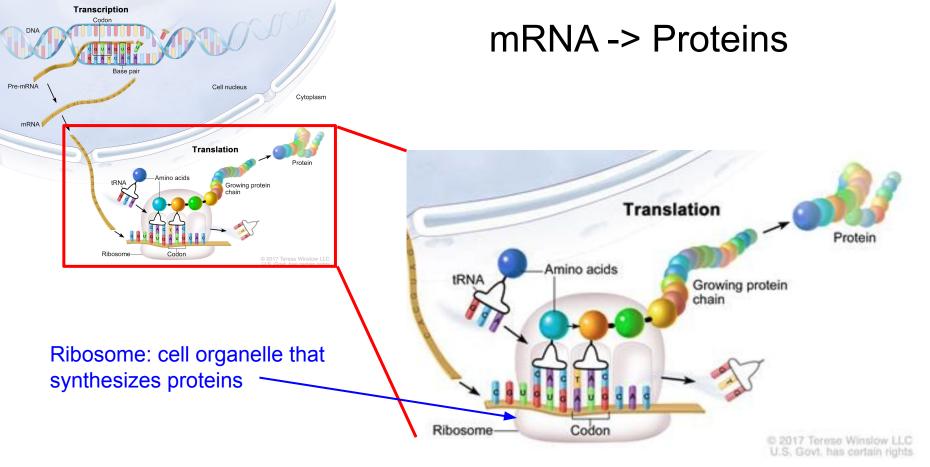


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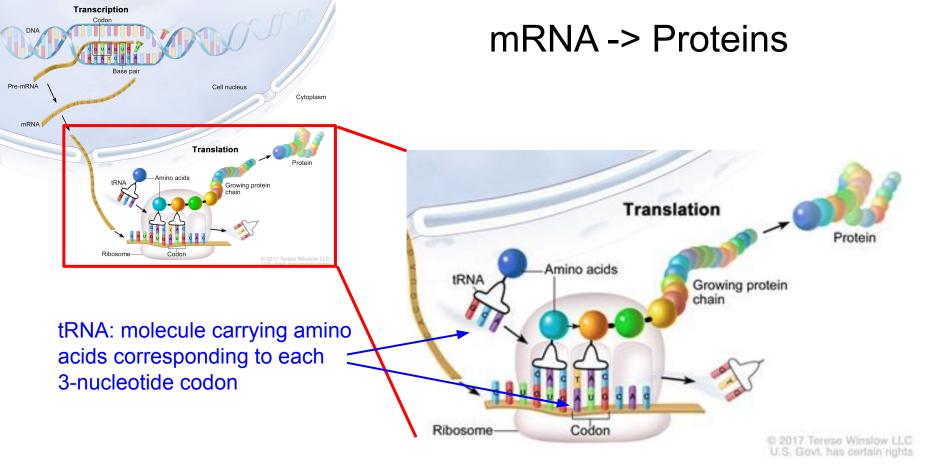
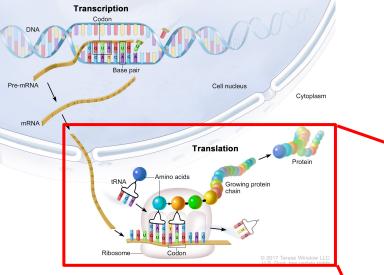


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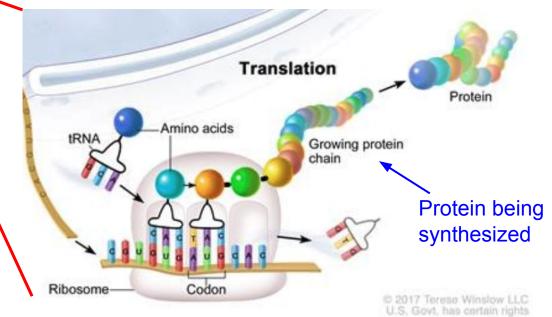
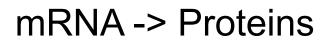
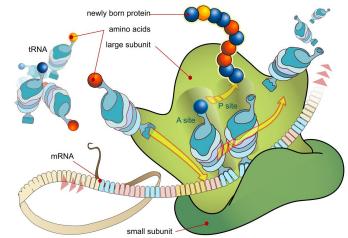


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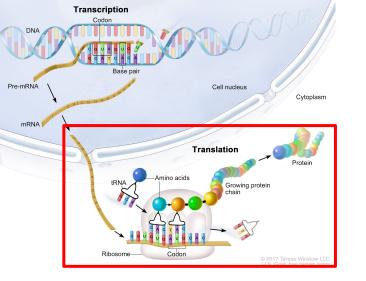


Figure credit:

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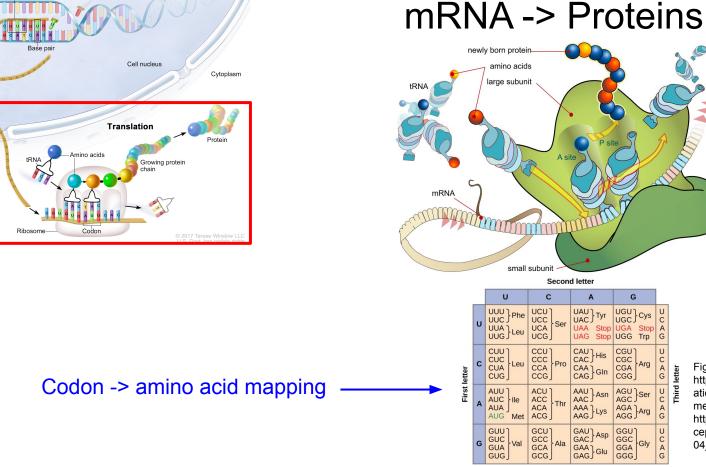


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Transcription Codon

Pre-mRNA

mRNA

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Study of processes that regulate how and when genes are turned on and off ("gene expression")



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Study of processes that regulate how and when genes are turned on and off ("gene expression")

- E.g. **transcription factors**: proteins that bind to the promoter and other noncoding regions, can enhance or repress transcription

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- E.g. DNA methylation: addition of large methyl group to promoter region makes it difficult for proteins to bind
 represses transcription

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Study of processes that regulate how and when genes are turned on and off ("gene expression")

- E.g. **transcription factors**: proteins that bind to the promoter and other noncoding regions, can enhance or repress transcription
- E.g. DNA methylation: addition of large methyl group to promoter region makes it difficult for proteins to bind
 -> represses transcription
- E.g. **Histone modification**: addition or removal of acetyl groups affects charge interaction to relax or tighten chromatin structure (easier for proteins to bind)

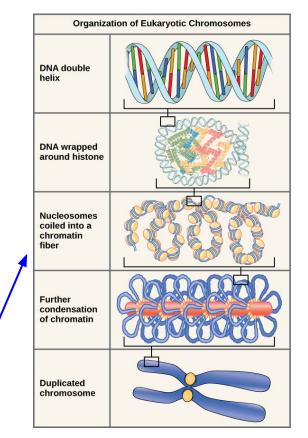


Figure credit:

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Lecture 9 - 35

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Transcriptomics

- Study of the transcriptome (the RNA of a cell)
- One reason of interest: Harder to measure proteins (the functional molecules!), but we can sequence RNA as a (highly imperfect) proxy for proteins to quantify cell state

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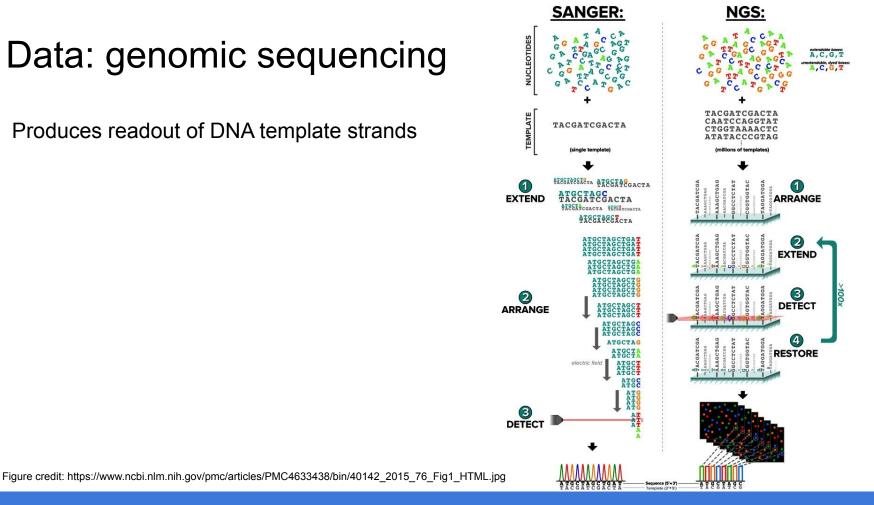
Transcriptomics

- Study of the transcriptome (the RNA of a cell)
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Proteomics

- Study of the proteins in a cell

Produces readout of DNA template strands

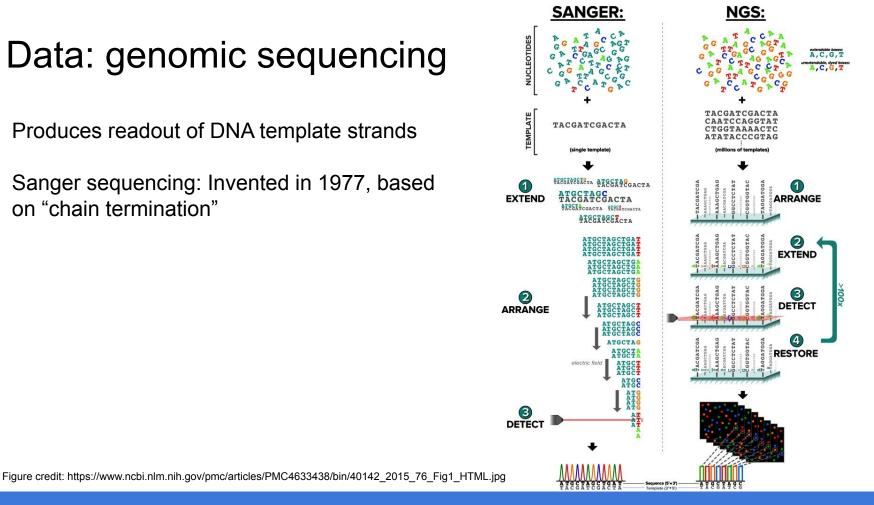


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Produces readout of DNA template strands

Sanger sequencing: Invented in 1977, based on "chain termination"



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Produces readout of DNA template strands

Sanger sequencing: Invented in 1977, based on "chain termination"

Add some special (and fluorescently labeled) nucleotides that cause a chain being synthesized to terminate

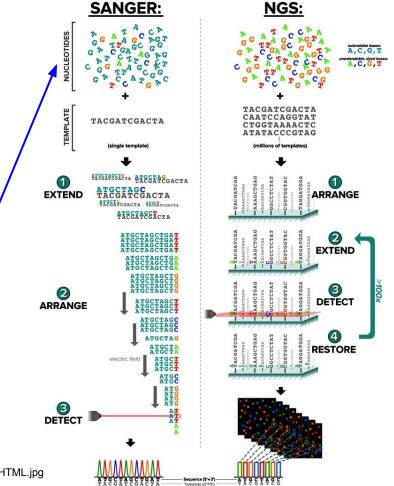


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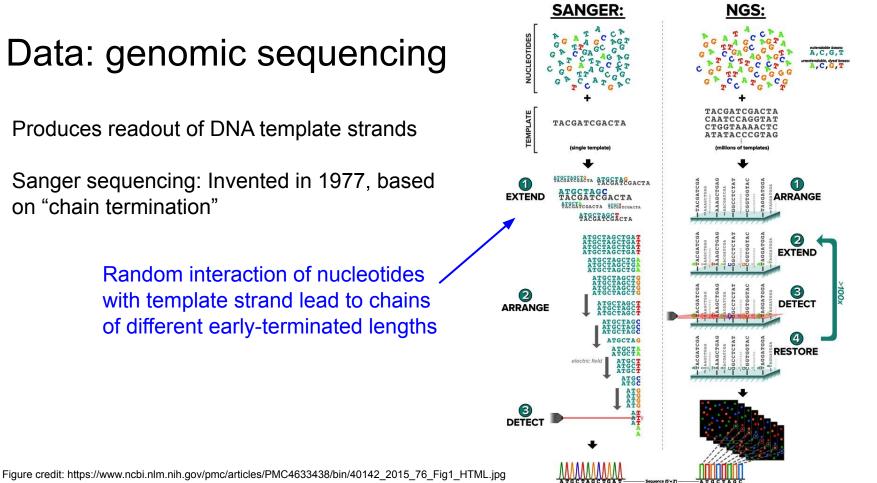
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Produces readout of DNA template strands

Sanger sequencing: Invented in 1977, based on "chain termination"

> Random interaction of nucleotides with template strand lead to chains of different early-terminated lengths



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Produces readout of DNA template strands

Sanger sequencing: Invented in 1977, based on "chain termination"

Sorting by length (e.g. electrophoresis) gives sequence readout

Figure credit: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4633438/bin/40142_2015_76_Fig1_HTML.jpg

SANGER: NGS: A,C,G,T C, G, T CAATCCAGGTAT TACGATCGACTA TEMPL TGGTAAAACTC CTA ATCCATCGACTA CGATCGACTA EXTEND ARRANGE ATSSTACGACTA ATSSTCCACT ATSETASET 2 EXTEND 3 2 DETECT ARRANGE TGCTAG RESTORE ATGCT ATGCT electric field ATGCT ATGCT ATGCT ATGC DETECT

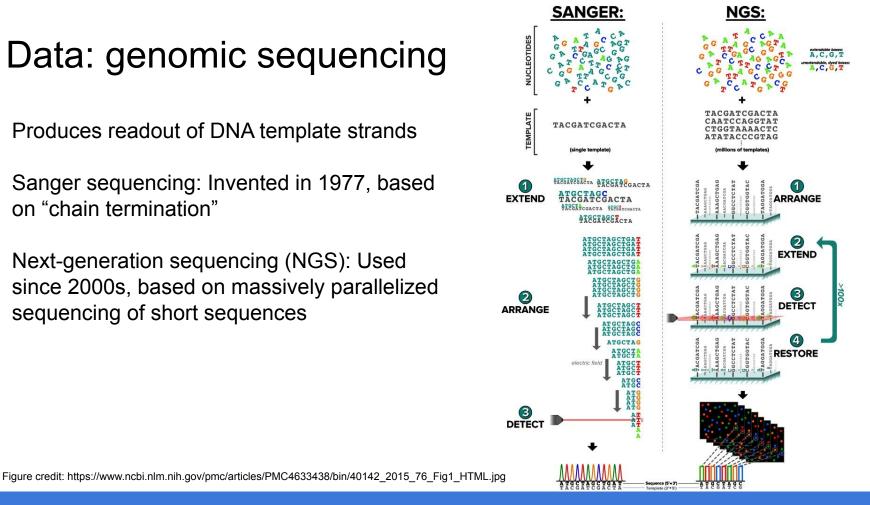
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Produces readout of DNA template strands

Sanger sequencing: Invented in 1977, based on "chain termination"

Next-generation sequencing (NGS): Used since 2000s, based on massively parallelized sequencing of short sequences



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SANGER: NGS: A,C,G,T TACGATCGACTA TA ATECTAGACTA CATCGACTA EXTEND ARRANGE ATSSTASST EXTEND ARRANGE DETECT RESTORE ATGCT electric field ATGCT ATGCT ATGCT

ATGC

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DETECT

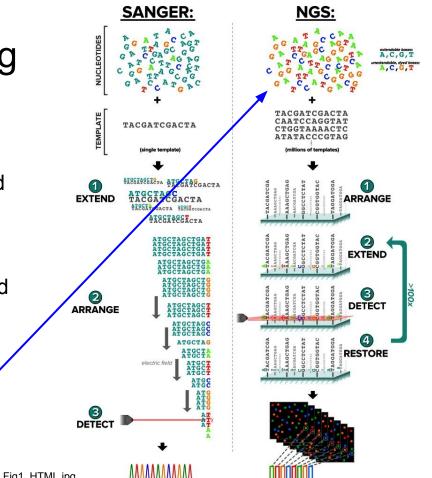
Produces readout of DNA template strands

Sanger sequencing: Invented in 1977, based on "chain termination"

Next-generation sequencing (NGS): Used since 2000s, based on massively parallelized sequencing of short sequences

Now all added nucleotides are chain-terminating

Figure credit: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4633438/bin/40142_2015_76_Fig1_HTML.jpg



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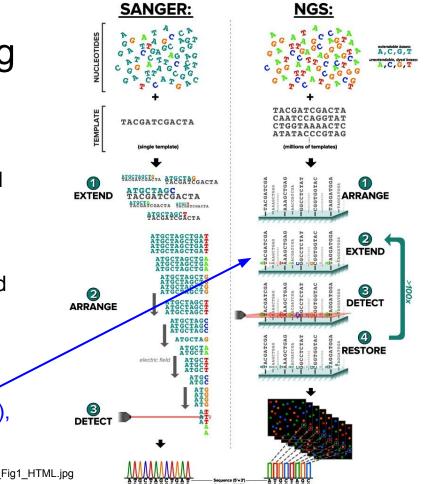
Produces readout of DNA template strands

Sanger sequencing: Invented in 1977, based on "chain termination"

Next-generation sequencing (NGS): Used since 2000s, based on massively parallelized sequencing of short sequences

All templates get next sequence / element attached (and terminated), then read

Figure credit: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4633438/bin/40142_2015_76_Fig1_HTML.jpg



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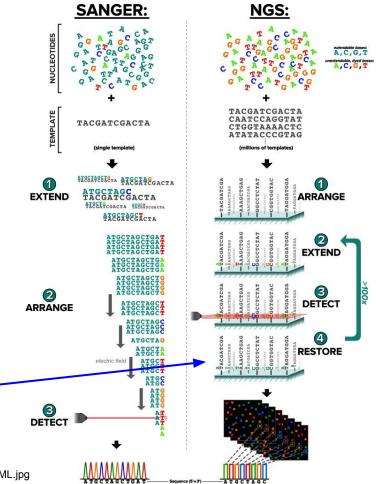
Produces readout of DNA template strands

Sanger sequencing: Invented in 1977, based on "chain termination"

Next-generation sequencing (NGS): Used since 2000s, based on massively parallelized sequencing of short sequences

Apply process to "restore" the chain-terminating nucleotides to be normal, – then repeat to extend synthesizing sequence by one more nucleotide

Figure credit: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4633438/bin/40142_2015_76_Fig1_HTML.jpg



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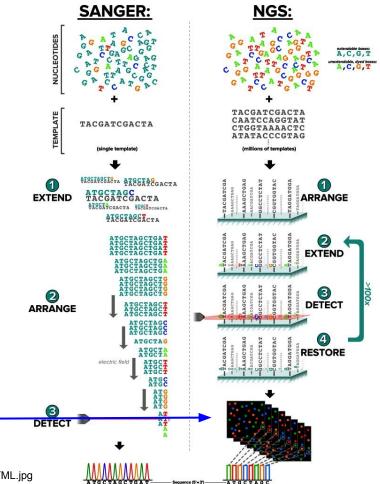
Produces readout of DNA template strands

Sanger sequencing: Invented in 1977, based on "chain termination"

Next-generation sequencing (NGS): Used since 2000s, based on massively parallelized sequencing of short sequences

Set of read-out images at every step gives sequences of all template strands. Then analyze data to reconstruct longer sequences.

Figure credit: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4633438/bin/40142_2015_76_Fig1_HTML.jpg



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Produces relative expression of genes in normal vs disease tissue samples

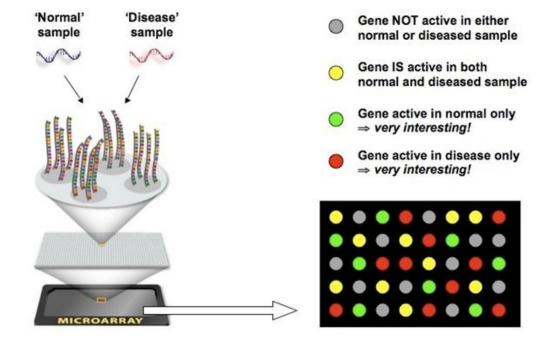


Figure credit: http://www.vce.bioninja.com.au/_Media/microarray_med.jpeg

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Produces relative expression of genes in normal vs disease tissue samples

Isolate mRNA ("expressed genes") from tissue samples and synthesize complementary DNA (cDNA).

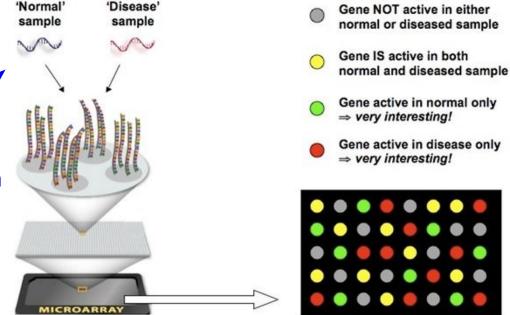


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Produces relative expression of genes in normal vs disease tissue samples

Isolate mRNA ("expressed genes") from tissue samples and synthesize complementary DNA (cDNA).

Use fluorescent tags to label cDNA from normal tissue green, and from disease tissue red

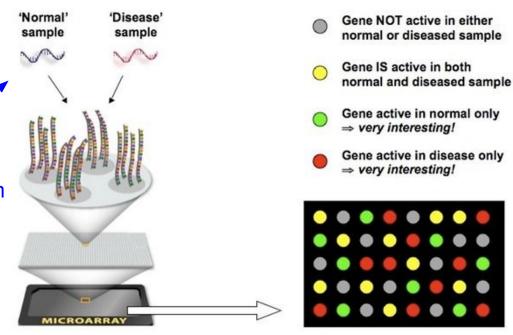


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Produces relative expression of genes in normal vs disease tissue samples

Each spot of DNA microarray contains single-stranded DNA

corresponding to a gene

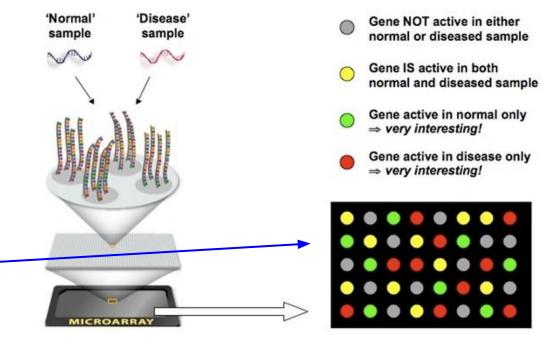


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Produces relative expression of genes in normal vs disease tissue samples

cDNA will bind to the corresponding DNA strands on __ microarray. Color indicates ratio of cDNA (relative gene expression) in the normal vs disease tissue

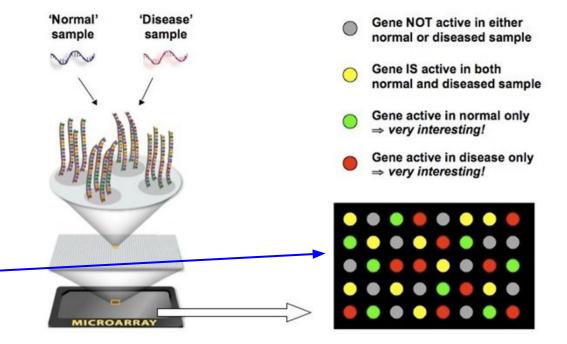


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Produces readout of mRNA content in a tissue sample

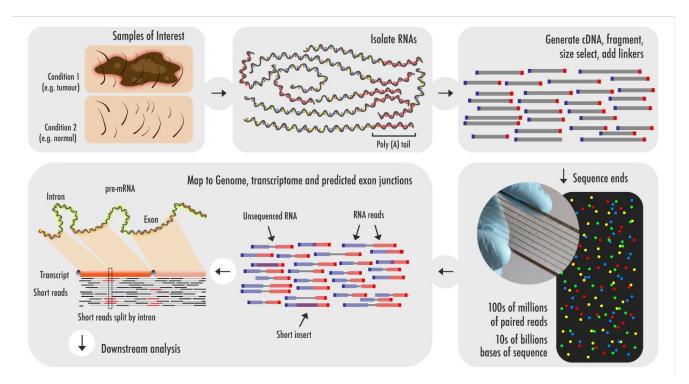


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Produces readout of mRNA content in a tissue sample

Isolate RNA and generate cDNA

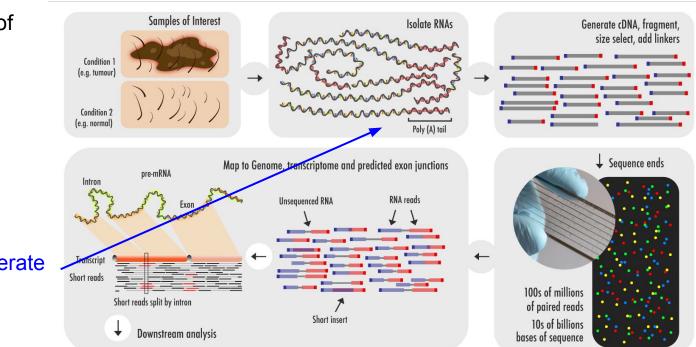


Figure credit: https://cdn.technologynetworks.com/tn/images/body/dnasequencinga1529596208892.png

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Produces readout of mRNA content in a tissue sample

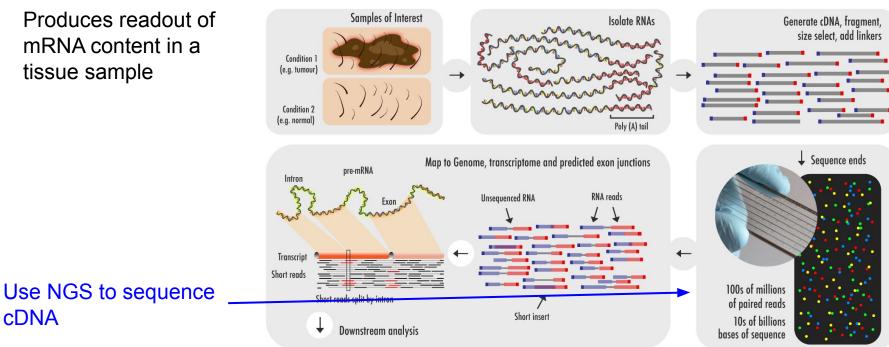


Figure credit: https://cdn.technologynetworks.com/tn/images/body/dnaseguencinga1529596208892.png

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cDNA

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Produces readout of mRNA content in a tissue sample

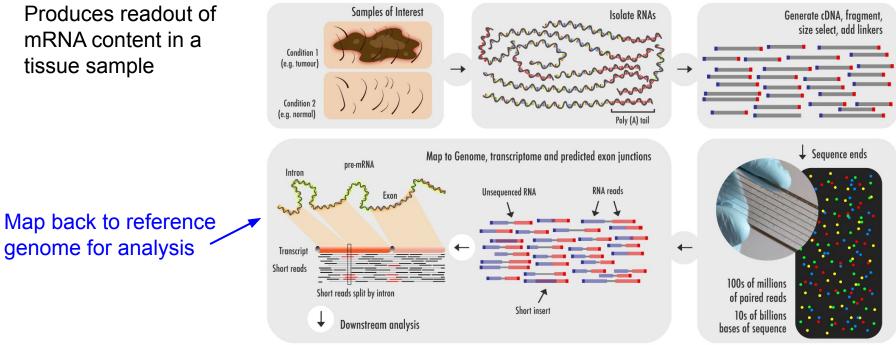


Figure credit: https://cdn.technologynetworks.com/tn/images/body/dnasequencinga1529596208892.png

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Produces readout of mRNA content in a tissue sample

Map back to reference genome for analysis

Now standard approach for transcriptomics study

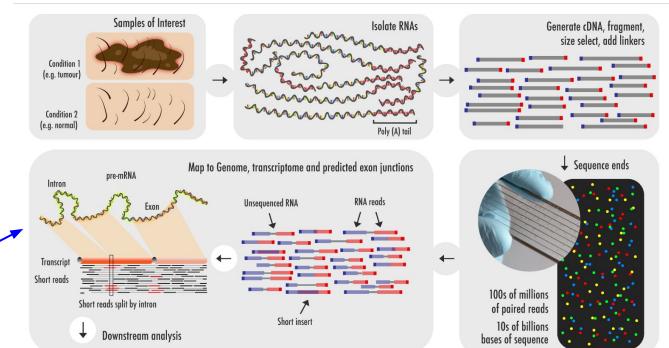


Figure credit: https://cdn.technologynetworks.com/tn/images/body/dnasequencinga1529596208892.png

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More recently in 2010s, single-cell RNA-seq!

Produces readout of mRNA content in a tissue sample

Map back to reference genome for analysis

Now standard approach for transcriptomics study

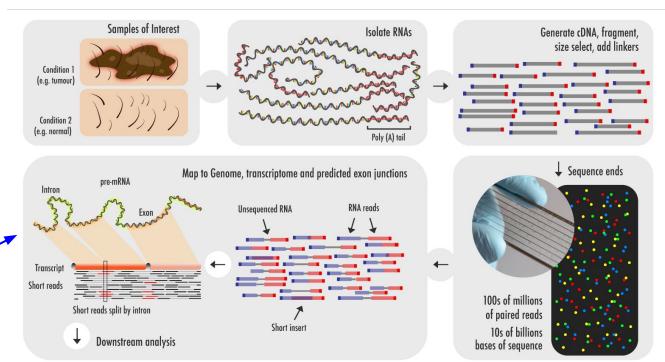
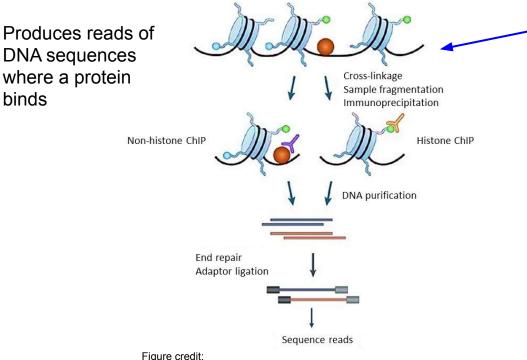


Figure credit: https://cdn.technologynetworks.com/tn/images/body/dnasequencinga1529596208892.png

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Use formaledehyde treatment to cross-link (fix) proteins to their bound DNA

https://www.france-genomique.org/wp-content/uploads/2019/08/CHIP-selon-P ark-1-e1566900408602.jpg

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binds

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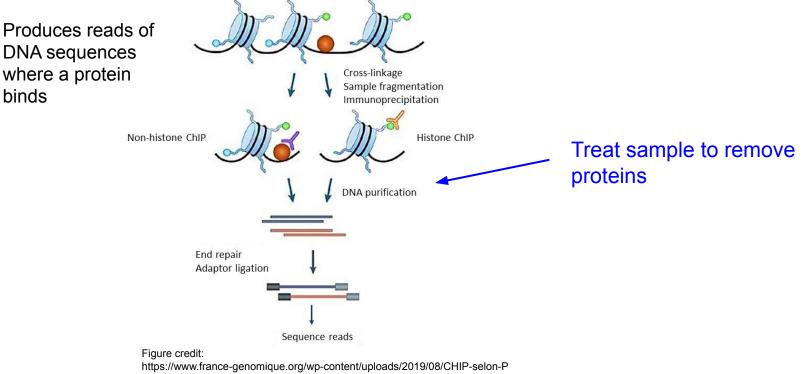
Produces reads of DNA sequences where a protein Cross-linkage Sample fragmentation binds Immunoprecipitation Non-histone ChIP Histone ChIP **DNA** purification End repair Adaptor ligation Sequence reads

Disintegrate non-bound DNA -> what is left is DNA segments bound to protein

Figure credit: https://www.france-genomique.org/wp-content/uploads/2019/08/CHIP-selon-P ark-1-e1566900408602.jpg

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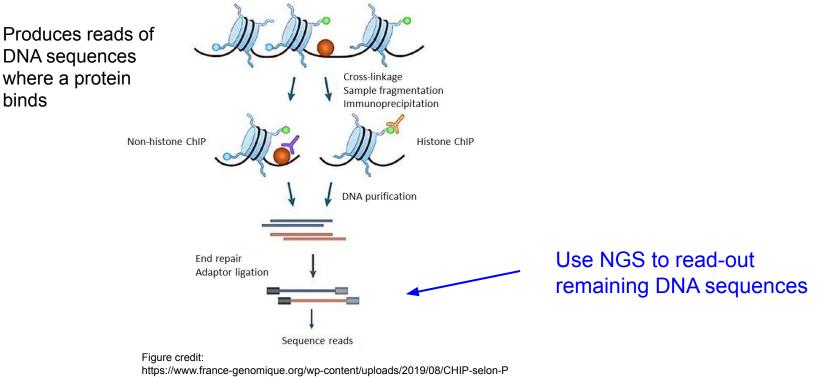
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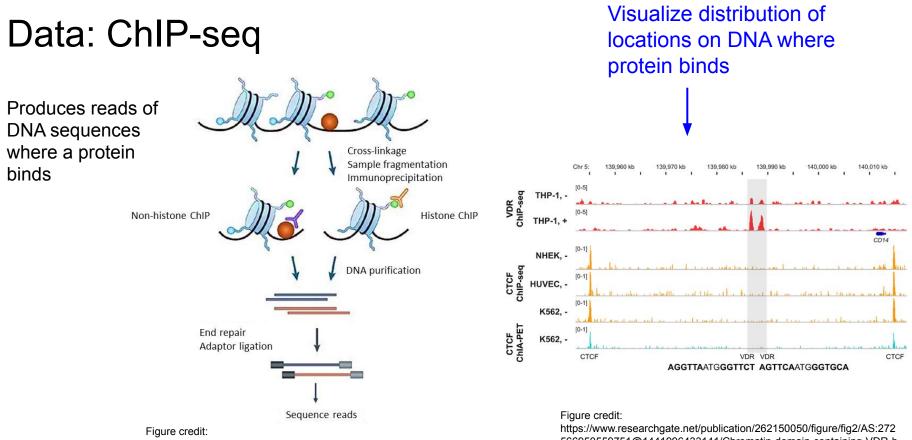
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https://www.france-genomique.org/wp-content/uploads/2019/08/CHIP-selon-P ark-1-e1566900408602.jpg

https://www.researchgate.net/publication/262150050/figure/fig2/AS:272 566950559751@1441996433141/Chromatin-domain-containing-VDR-b inding-sites-The-IGV-browser-was-used-to-display-the.png

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ENCODE: identifying and analyzing all functional elements in the human genome

- Launched by US National Human Genome Research Institute in 2003
- Contributions from worldwide consortium of research groups

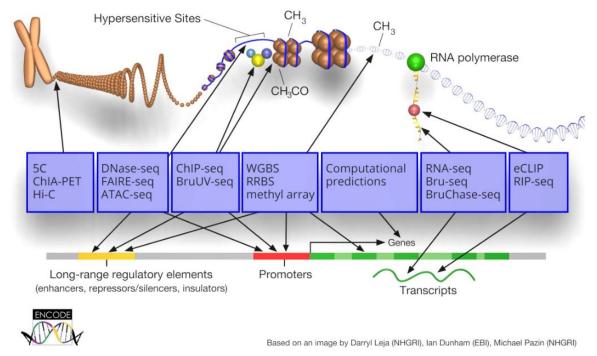
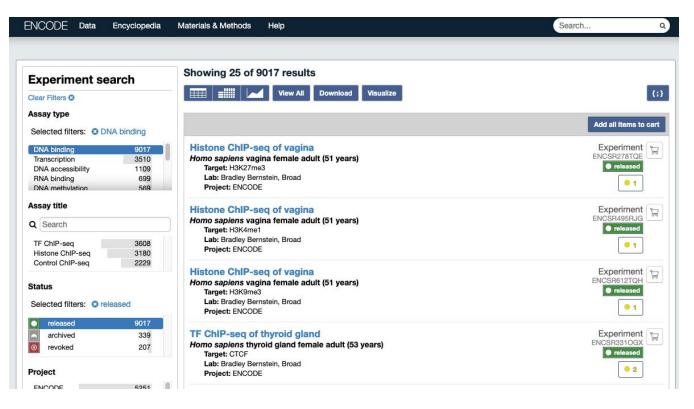


Figure credit: https://www.encodeproject.org/

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ENCODE data



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ENCODE data

ENCODE Data 6	Encyclopedia	Materials & Methods Help	Search Q
Experiment sea	arch	Showing 25 of 3510 results	
Clear Filters 3		Image: Second state Image: Second state View All Download Visualize	(;)
Assay type			
Selected filters: 3 Tran	scription		Add all items to cart
DNA binding Transcription DNA accessibility RNA binding DNA methylation	9017 3510 1109 699 569	Iong read RNA-seq of left lung Homo sapiens left lung male adult (40 years) Lab: Ali Mortazavi, UCI Project: ENCODE	Experiment ENCSR426KOP released
Assay title Q Search		long read RNA-seq of left lung Homo sapiens left lung female child (16 years) Lab: Ali Mortazavi, UCI	Experiment ENCSR746ITG released
polyA plus RNA-seq total RNA-seq	770 704	Project: ENCODE	b 1
shRNA RNA-seq small RNA-seq	640 214	long read RNA-seq of ovary	Experiment
microRNA-sea	201	Homo sapiens ovary female adult (41 years) Lab: Ali Mortazavi, UCI	ENCSR587WPR
Status		Project: ENCODE	B 1
Selected filters: 3 relea	ased		
o released	3510	long read RNA-seq of mucosa of descending colon Homo sapiens mucosa of descending colon female adult (61 years)	
archived revoked	287 24	Lab: Ali Mortazavi, UCI Project: ENCODE	• released
Broloot	27	FIJEG: ENCODE	🖺 1

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ENCODE data

Common Cell Types: Tier 1 and Tier 2

Cell, tissue or DNA sample: Cell line or tissue used as the source of experimental material.

cell ^{↓1}	Tier ^{↓2}	Description ¹³	Lineage ^{↓4}	Tissue ^{↓5}	Karyotype	Sex	Documents	Vendor ID
GM12878		B-lymphocyte, lymphoblastoid, International HapMap Project - CEPH/Utah - European Caucasion, Epstein-Barr Virus	mesoderm	blood	normal	F		Coriell GM12878
H1-hESC	1	lempryonic stem cells		embryonic stem cell	normal	М	ENCODE	WiCell Research Institute WA01
K562	1	leukemia, "The continuous cell line K-562 was established by Lozzio and Lozzio from the pleural effusion of a 53-year-old female with chronic myelogenous leukemia in terminal blast crises." - ATCC	mesoderm	blood	cancer	F		ATCC CCL-243

Total = 3

Cell, tissue or DNA sample: Cell line or tissue used as the source of experimental material.

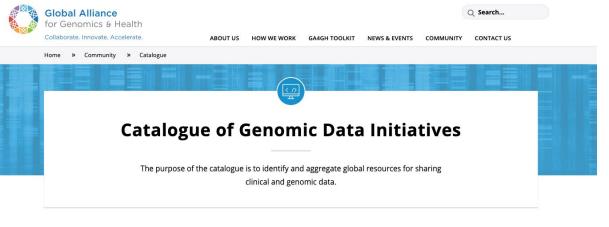
cell ^{↓1}	Tier ^{↓2}	Description ^{↓3}	Lineage ^{↓4}	Tissue ^{↓5}	Karyotype	Sex	Documents	Vendor ID	Τ
A549	2	epithelial cell line derived from a lung carcinoma tissue. (PMID: 175022), "This line was initiated in 1972 by D.J. Giard, et al. through explant culture of lung carcinomatous tissue from a 58-year-old caucasian male." - ATCC, newly promoted to tier 2: not in 2011 analysis		epithelium	cancer	М	Myers Crawford Stam	ATCC CCL-185	E
									T

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Other datasets

https://www.ga4gh.org/community/catalogue



Enter Keyword		40 of 220 Initiative	es	« 1 2 3 4 »
Filters <u>R</u>	<u>eset</u>	1000 Genomes		
Category	÷	CATEGORY: eHealth		INITIATIVE TYPE: Database
(19/102) Mendelian Genetic Disorders (13/41) eHealth (12/92)	÷	individuals from 26 popula data, high-density genotyp the International Genome human genome assembly	ations. Additional work was done to inve ing chip calls and cell lines from the Pro Sample Resource (IGSR), which is realig and also expanding the data resources	genetic variation, publishing a set of variations based on sequencing o estigate structural variations in the human genome. Variant calls, sequ oject are all available. Data from the 1000 Genomes Project is now hou gning sequence data from the 1000 Genomes Project to the updated G s produced by 1000 Genomes to include new samples with similarly op er information, access to data and user support are a
Dibiobank/Repository (17/17)		CONTACT:	test1	
Consortium/Collaborat Network (75/75)	tive	Antigenic Variati	on Database (VarDB)	
Database (40/40) GA4GH Driver Project		CATEGORY: INITIATIVE TYPE:		

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Other datasets

https://www.ga4gh.org/community/catalogue

nature

THE INTERNALL. I WEEKLY JOURNAL OF SCIENCE

UK

Genetic and health data

from half a million people in the United Kingdom

PAGES 194, 203 & 210

BIOBAN



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itiative

aborative

Database (40/40)

GA4GH Driver Project

Catalogue of Genomic Data Initiatives

The purpose of the catalogue is to identify and aggregate global resources for sharing clinical and genomic data.

Initiatives



1000 Genomes

CATEGORY:

eHealth

INITIATIVE TYPE:

Database

The 1000 Genomes Project set out to catalogue common human genetic variation, publishing a set of variations based on sequencing of 2504 individuals from 26 populations. Additional work was done to investigate structural variations in the human genome. Variant calls, sequence data, high-density genotyping chip calls and cell lines from the Project are all available. Data from the 1000 Genomes Project is now housed in the International Genome Sample Resource (IGSR), which is realigning sequence data from the 1000 Genomes Project to the updated GRCh38 human genome assembly and also expanding the data resources produced by 1000 Genomes to include new samples with similarly open consent, new populations and a wider range of data types. Further information, access to data and user support are a

CONTACT:

Antigenic Variation Database (VarDB)

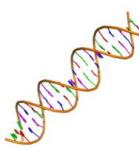
CATEGORY: INITIATIVE TYPE:

test1

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Remember from Lecture 1



1953 - Watson and Crick discover double helix structures of DNA



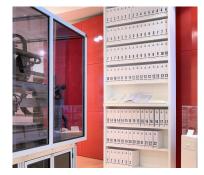
2003: ENCODE project launched to identify and characterize genes in human genome



1977 - Fred Sanger sequences first full genome of a virus



1000 Genomes Project: 2008 - 2015



1990 - 2003: Human Genome Project sequences full human genome

The 100,000 Genomes Project

Genomics England & Partners



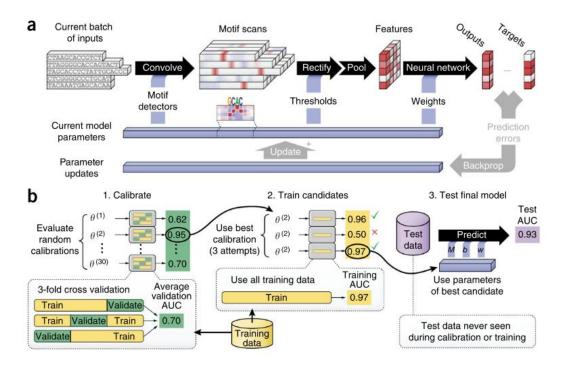
UK100,000 Genomes Project: 2012 - 2018

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DeepBind

Input: DNA sequence Output: Score of whether a particular protein will bind to the sequence or not

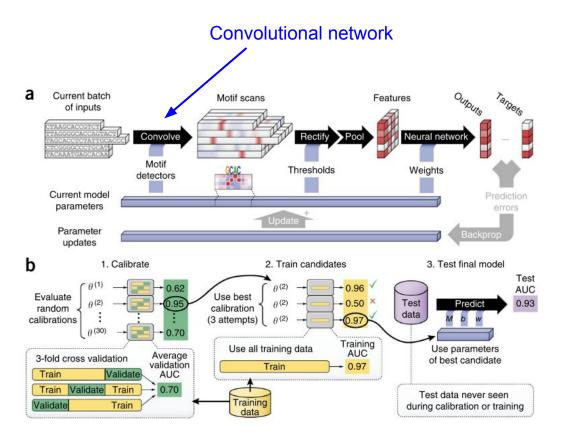


Alipanahi et al. Predicting the sequence specificities of DNA- and RNA-binding proteins by deep learning. Nature Biotechnology, 2015.

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Input: DNA sequence Output: Score of whether a particular protein will bind to the sequence or not



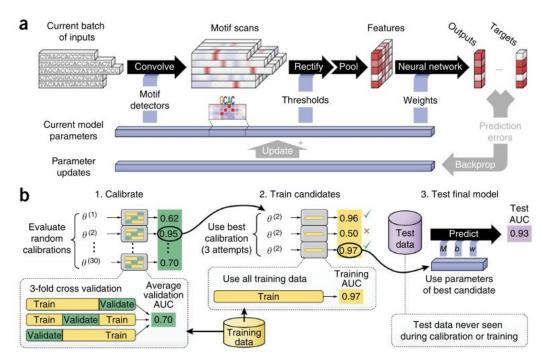
Alipanahi et al. Predicting the sequence specificities of DNA- and RNA-binding proteins by deep learning. Nature Biotechnology, 2015.

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Input: DNA sequence Output: Score of whether a particular protein will bind to the sequence or not

- Processing to handle different sources of experimental (training) data and input / output data formats
- Trained on 12 TB of sequence data; learned 927 DeepBind models representing 538 transcription factor (TF) proteins and 194 RNA-binding proteins (RBPs)



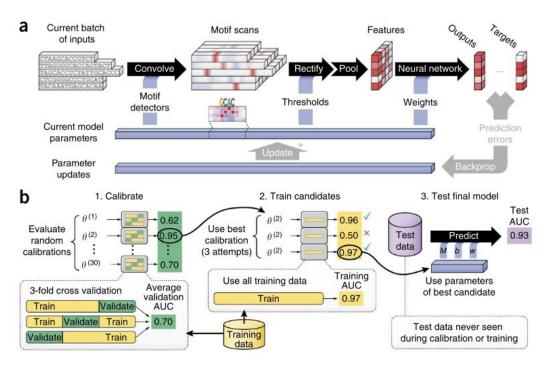
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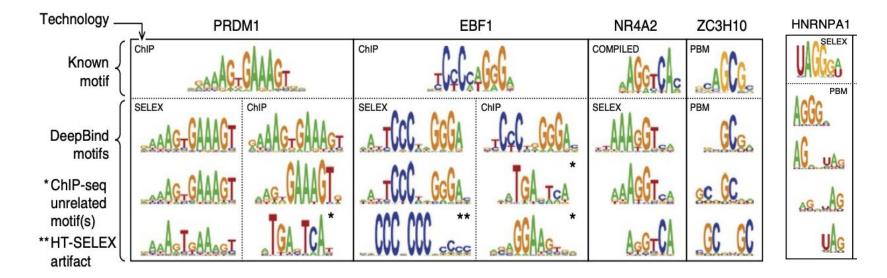
Outperformed prior methods on the DREAM5 TF-DNA Motif Recognition Challenge



Alipanahi et al. Predicting the sequence specificities of DNA- and RNA-binding proteins by deep learning. Nature Biotechnology, 2015.

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Learned DeepBind motifs

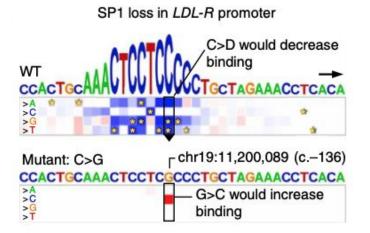


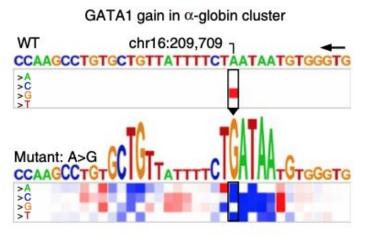
Alipanahi et al. Predicting the sequence specificities of DNA- and RNA-binding proteins by deep learning. Nature Biotechnology, 2015.

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Predicted effect of sequence mutations

Mutation would increase score Mutation would have no effect Mutation would decrease score



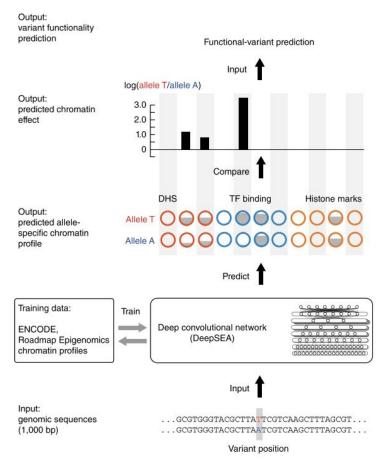


Alipanahi et al. Predicting the sequence specificities of DNA- and RNA-binding proteins by deep learning. Nature Biotechnology, 2015.

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Predict chromatin effects of (non-coding) sequence alterations with single-nucleotide sensitivity (SNPs: single nucleotide polymorphism)



Lecture 9 - 78

Zhou and Troyanskaya. Predicting effects of noncoding variants with deep learning-based sequence model. Nature Methods, 2015.

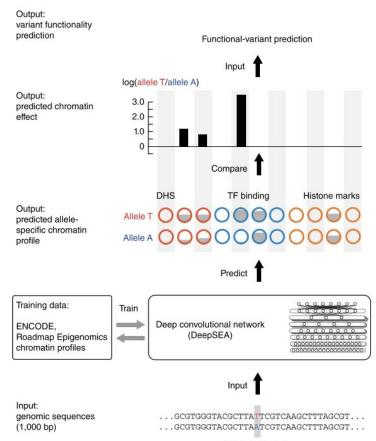
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Predict chromatin effects of (non-coding) sequence alterations with single-nucleotide sensitivity (SNPs: single nucleotide polymorphism)

Input: DNA sequence pair with SNP Output: Predicted chromatin effects (919 total)

- 690 transcription factor profiles
- 125 DNase I hypersensitive sites (DHS) profiles (looser chromatin structure, easier protein binding)
- 104 histone-mark profiles (histone modifications)

Multi-task training!



Variant position

Zhou and Troyanskaya. Predicting effects of noncoding variants with deep learning-based sequence model. Nature Methods, 2015.

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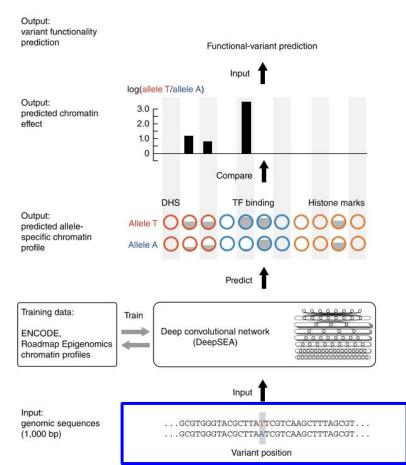
BIODS 220: AI in Healthcare

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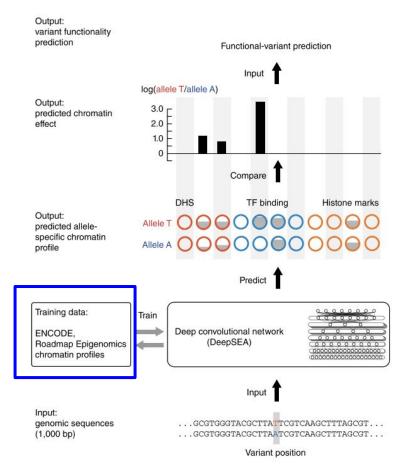
BIODS 220: AI in Healthcare

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Input: DNA sequence pair with SNP Output: Predicted chromatin effects (919 total)

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- 104 histone-mark profiles (histone modifications)

Multi-task training!



Zhou and Troyanskaya. Predicting effects of noncoding variants with deep learning-based sequence model. Nature Methods, 2015.

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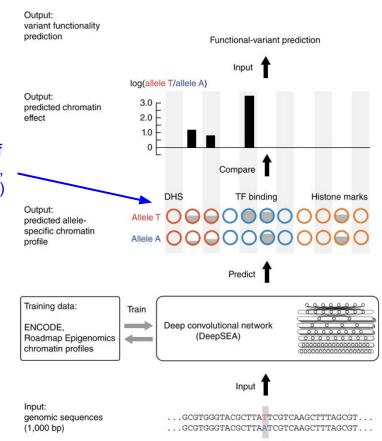
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Predict chromatin effects of (non-coding) sequence alterations with single-nucleotide sensitivity (SNPs: single nucleotide polymorphism) Multi-task prediction of 919 chromatin profiles,

Input: DNA sequence pair with SNP for each allele (variant) Output: Predicted chromatin effects (919 total)

- 690 transcription factor profiles
- 125 DNase I hypersensitive sites (DHS) profiles (looser chromatin structure, easier protein binding)
- 104 histone-mark profiles (histone modifications)

Multi-task training!



Variant position

Zhou and Troyanskaya. Predicting effects of noncoding variants with deep learning-based sequence model. Nature Methods, 2015.

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Predict chromatin effects of (non-coding) sequence

alterations with single-nucleotide sensitivity (SNPs:

Output: Predicted chromatin effects (919 total)

104 histone-mark profiles (histone

125 DNase I hypersensitive sites (DHS)

profiles (looser chromatin structure, easier

690 transcription factor profiles

single nucleotide polymorphism)

protein binding)

modifications)

Multi-task training!

Input: DNA sequence pair with SNP

Output: Interested in relative variant functionality prediction Functional-variant prediction Input log(allele T/allele A) Output: 3.0 predicted chromatin 2.0 effect 1.0 Compare DHS TF binding Histone marks Output: Θ predicted allelespecific chromatin Allele A profile Predict Training data: Train Deep convolutional network ENCODE. (DeepSEA) Roadmap Epigenomics chromatin profiles Input Input: genomic sequences GCGTGGGTACGCTTATTCGTCAAGCTTTAGCGT... (1,000 bp) ... GCGTGGGTACGCTTAATCGTCAAGCTTTAGCGT... Variant position

Zhou and Troyanskaya. Predicting effects of noncoding variants with deep learning-based sequence model. Nature Methods, 2015.

effect

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Model Architecture:

- 1. Convolution layer (320 kernels. Window size: 8. Step size: 1.)
- 2. Pooling layer (Window size: 4. Step size: 4.)
- 3. Convolution layer (480 kernels. Window size: 8. Step size: 1.)
- 4. Pooling layer (Window size: 4. Step size: 4.)
- 5. Convolution layer (960 kernels. Window size: 8. Step size: 1.)
- 6. Fully connected layer (925 neurons)
- 7. Sigmoid output layer

Zhou and Troyanskaya. Predicting effects of noncoding variants with deep learning-based sequence model. Nature Methods, 2015.

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Variant calling: identifying variants from reference genome (SNPs, small indels, etc.)

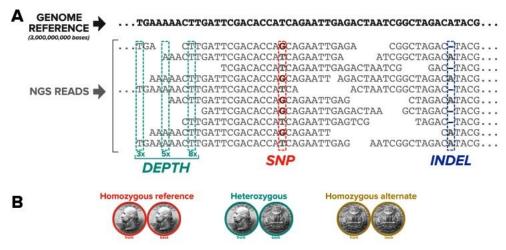
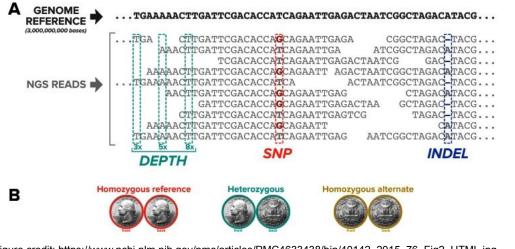


Figure credit: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4633438/bin/40142_2015_76_Fig2_HTML.jpg

Poplin et al. A universal SNP and small-indel variant caller using deep neural networks. Nature Biotechnology, 2018.

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Variant calling: identifying variants from reference genome (SNPs, small indels, etc.)



Challenge with short, errorful

sequence reads from NGS!

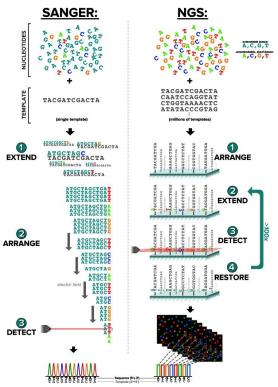


Figure credit: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4633438/bin/40142_2015_76_Fig2_HTML.jpg

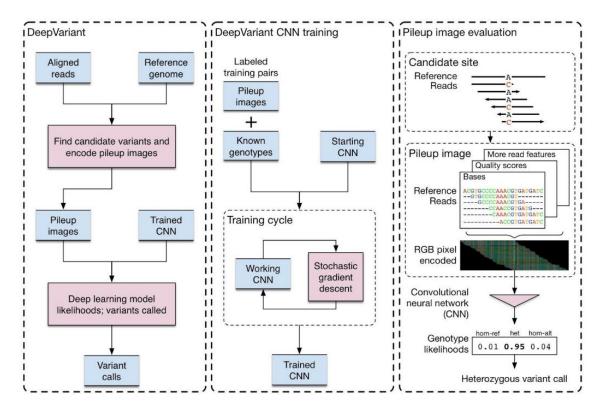
Poplin et al. A universal SNP and small-indel variant caller using deep neural networks. Nature Biotechnology, 2018.

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Input: "Pileup images" of reference sequence + NGS reads, + other features

Output: Categorical prediction of variant type (hom-ref, het, hom-alt), or no variant

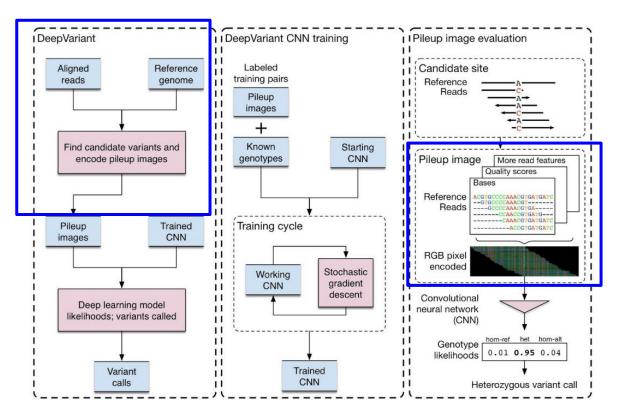


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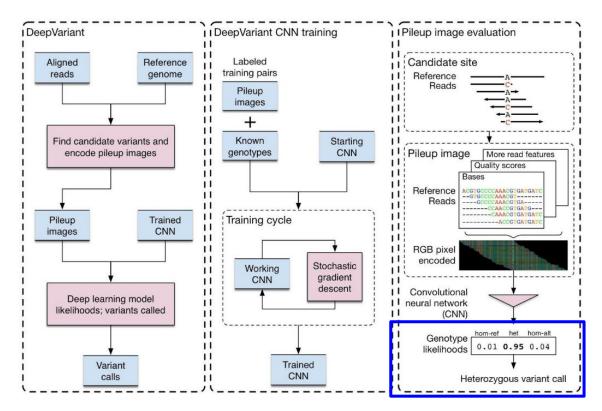


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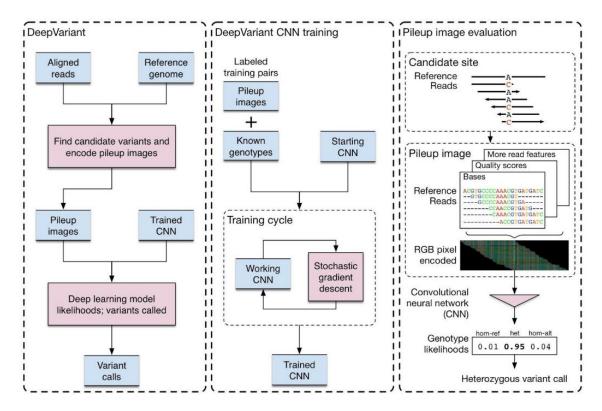
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Output: Categorical prediction of variant type (hom-ref, het, hom-alt), or no variant

Used an Inception v3 CNN



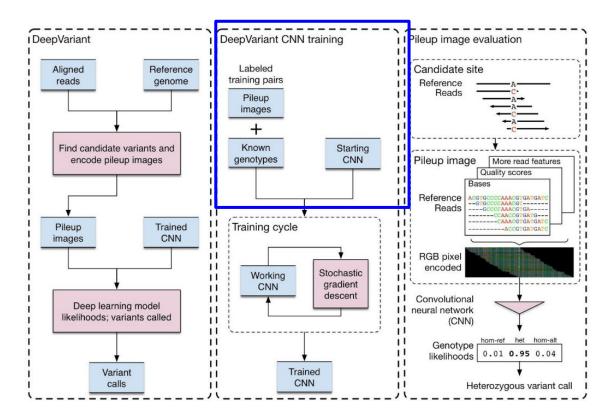
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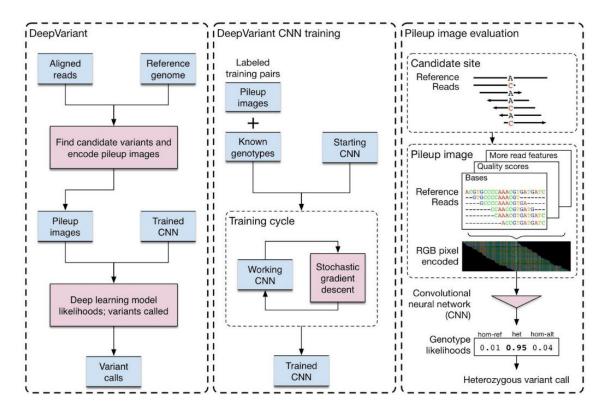
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Input: "Pileup images" of reference sequence + NGS reads, + other features

Output: Categorical prediction of variant type (hom-ref, het, hom-alt), or no variant

Used an Inception v3 CNN

Won highest performance for SNPs in the 2016 FDA variant calling Truth Challenge



Poplin et al. A universal SNP and small-indel variant caller using deep neural networks. Nature Biotechnology, 2018.

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More examples of deep learning in genomics

Epigenomics:

 Predicting methylation states, gene expression from histone modifications, etc.

Transcriptomics:

- Predicting phenotypes from transcriptome, identifying genes associated with transcriptomic data, etc.

Proteomics:

- Predicting secondary structure of proteins, protein-protein interactions, etc.

Summary

Today we covered:

- Biology basics for genomics
- Epigenomics, transcriptomics, proteomics
- Genomics data
- Examples of deep learning for genomics