Clinical Proton MR Spectroscopy in Central Nervous System Disorders¹

A large body of published work shows that proton (hydrogen 1 [¹H]) magnetic resonance (MR) spectroscopy has evolved from a research tool into a clinical neuroimaging modality. Herein, the authors present a summary of brain disorders in which MR spectroscopy has an impact on patient management, together with a critical consideration of common data acquisition and processing procedures. The article documents the impact of ¹H MR spectroscopy in the clinical evaluation of disorders of the central nervous system. The clinical usefulness of ¹H MR spectroscopy has been established for brain neoplasms, neonatal and pediatric disorders (hypoxia-ischemia, inherited metabolic diseases, and traumatic brain injury), demyelinating disorders, and infectious brain lesions. The growing list of disorders for which ¹H MR spectroscopy may contribute to patient management extends to neurodegenerative diseases, epilepsy, and stroke. To facilitate expanded clinical acceptance and standardization of MR spectroscopy methodology, guidelines are provided for data acquisition and analysis, quality assessment, and interpretation. Finally, the authors offer recommendations to expedite the use of robust MR spectroscopy methodology in the clinical setting, including incorporation of technical advances on clinical units.

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ince the inception of magnetic resonance (MR) imaging in the 1980s, its employment in the diagnostic evaluation of the central nervous system (CNS) has had a major impact on patient management. With the advent of 1.5-T whole-body magnets, imaging of the CNS with unprecedented detail became possible by using the proton (hydrogen 1 \(^{1}\text{H}\)) signal of water. Complementary to structural MR imaging, \(^{1}\text{H}\) MR spectroscopy has become an attractive approach with which to assess the levels of metabolites in normal and diseased CNS, especially as image-controlled, localized MR spectroscopy acquisition techniques were developed. These early localization techniques included point-resolved spectroscopy (PRESS) (1,2) and stimulated echo acquisition mode (STEAM) (3), methods that are now widely used in clinical MR spectroscopy applications.

Preliminary studies revealed large differences in metabolite levels in acute stroke (4), chronic multiple sclerosis (5), and brain tumors compared with healthy brain (6). Although this work stimulated a surge of interest in \(^{1}\text{H}\) MR spectroscopy for diagnosing and assessing CNS disorders during the early days of the “Decade of the Brain” (1990–1999), many suboptimal patient studies (7) and the lack of consistent guidelines have led to a situation where, 20 years later, MR spectroscopy is still considered an “investigational technique” by some medical professionals and health care organizations. However, the ability to make an early, noninvasive diagnosis or to increase confidence in a suspected diagnosis is highly valued by patients and clinicians alike. As a result, an increasing number of imaging centers are incorporating MR spectroscopy into their clinical protocols for brain examinations in selected patients. To facilitate expanded use of MR spectroscopy in the clinical setting, this consensus statement encourages standardization of data acquisition, analysis, and reporting of results.

When assessing the impact of imaging techniques on health care (8), it is recommended that six criteria be evaluated: (a) technical feasibility, (b) diagnostic accuracy, (c) diagnostic impact, (d) therapeutic impact, (e) impact on outcome, and (f) societal impact (9). Although MR spectroscopy certainly fulfills the first two criteria, only a few studies have demonstrated that it has a wide impact on differential diagnosis, patient treatment, and outcome and none have measured the societal impact (ie, cost-benefit analysis) (8). Thus, it remains a challenge and task of high priority for the MR spectroscopy community to focus on studies that will quantify the extent to which MR spectroscopy improves diagnosis and leads to changes in patient treatment and resulting in improved outcomes. This consensus article has been produced by an international group of imaging scientists, neuroradiologists, neurologists, oncologists, and clinical neuroscientists from universities and MR vendors to document the impact of \(^{1}\text{H}\) MR spectroscopy in the clinical evaluation of disorders of the CNS. The MR Spectroscopy Consensus Group was formed from October 2011 to April 2012. The group drafted and finalized the manuscript jointly through e-mail correspondence and teleconferences with the group members and by means of two special interest group meetings held in connection to the 20th Scientific Meeting of the International Society for Magnetic Resonance in Medicine in May 2012 and the 21st Scientific Meeting of the International Society for Magnetic Resonance in Medicine in April 2013.

**Essentials**

- Hydrogen 1 \(^{1}\text{H}\) MR spectroscopy is complementary to MR imaging and adds clinically relevant information about metabolites in common brain abnormalities.
- MR spectroscopy is clinic-ready for diagnostic, prognostic, and treatment assessment of brain tumors, various neonatal and pediatric disorders (hypoxia-ischemia, inherited metabolic diseases, and traumatic brain injury), demyelinating disorders, and infectious brain lesions; it is expected to contribute to patient management in neurodegenerative disorders, epilepsy, and stroke.
- Provided that spectra are acquired reproducibly with a protocol that adheres to quality standards, clinical MR spectroscopy can be performed successfully at either 1.5 or 3.0 T.
- MR spectroscopy data acquisition and processing procedures must be harmonized across vendors for expanded clinical acceptance, as lack of standardization and quality assurance of MR spectroscopy data acquisition and analysis methods is a current impediment to widespread clinical use.

**MR Spectrum of the Brain: Metabolites and Their Biomarker Potential**

MR spectroscopy provides a very different basic “readout” than MR imaging, namely a spectrum rather than an

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**Abbreviations:**

CNS = central nervous system
Cr = creatine
Gn = glutamine
Glu = glutamate
Lac = lactate
mIns = myo-inositol
NAA = N-acetylaspartate
PRESS = point-resolved spectroscopy
SNR = signal-to-noise ratio
STEAM = stimulated echo acquisition mode
tCho = total choline
tCr = total creatine
tE = echo time
tNAA = N-acetylaspartylglutamate

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Conflicts of interest are listed at the end of this article.
**SPECIAL REVIEW**: Clinical Proton MR Spectroscopy in Central Nervous System Disorders

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**Image (Fig 1)**: 1H MR spectrum acquired at 3.0 T from a volume of interest in occipital lobe (20 × 20 × 20 mm³, T1-weighted axial image) of healthy subject with the STEAM sequence (repetition time msec/echo time [TE] msec = 5000/8; 128 repetitions). tNAA = total N-acetylaspartate (NAA), tCr = total creatine (Cr), tCho = total choline, Glu = glutamate, Gln = glutamine, mlns = myo-inositol, MM = macromolecules.

Although MR images are conventionally displayed as gray-scale images that radiologists interpret by means of visual inspection of signal intensities and geometric structures, the MR spectrum consists of resonances or peaks that represent signal intensities as a function of frequency (commonly expressed as parts per million, a relative, magnetic field–independent frequency scale). Spectra are obtained either from one selected brain region in the case of single-voxel spectroscopy or from multiple brain regions in the case of MR spectroscopic imaging. The spectral data format has no antecedent in radiology, as MR images do in radiographic films, which may be one of the reasons for the relatively slow acceptance of MR spectroscopy in the clinical imaging community. Nevertheless, currently available analysis methods can help automatically and reliably quantify MR spectra in the clinical setting.

In vivo 1H MR spectroscopy focuses on carbon-bound protons in the 1–5 ppm range of the chemical shift scale (Fig 1) and can depict metabolites that are present at high enough concentrations (within the micromoles per gram range) and mobile on the MR spectroscopy time scale. These include the neuronal metabolite NAA, the glial metabolite mlns, choline-containing compounds such as glycerophosphocholine and phosphocholine, neurotransmitters Glu and γ-aminobutyric acid, antioxidants glutathione and ascorbate, and other important metabolites such as Cr, phosphocreatine, Gln, and lactate (Lac) (10,11). Additional metabolites arise in specific clinical conditions, such as succinate and acetate in abscesses (12), lipids in various abnormalities (13,14), and even exogenous substances that cross the blood-brain barrier, such as propylene glycol after administration of some parenteral preparations (15) and ethanol after at least moderate alcohol consumption (16).

The number of quantifiable metabolites depends on the chosen pulse sequence and parameters, as well as the spectral resolution and signal-to-noise ratio (SNR), which are affected by many factors including the static magnetic field strength, quality of B₀ field homogeneity, and radiofrequency coil used (17,18). The major singlet resonances originating from total MR spectroscopy–visible NAA (tNAA) (ie, NAA + N-acetylaspartylglutamate), tCr (ie, Cr + phosphocreatine), and tCho (ie, primarily phosphocholine + glycerophosphocholine) can be quantified at all clinical magnetic field strengths and at almost all practical TE's up to 280 msec (19,20). At 1.5 T and short TE's (25–35 msec for PRESS, 20 msec or shorter for STEAM), mlns and combined Glu and Gln can also be quantified (21). At field strengths of 3.0 T and higher, additional metabolites are detected at short TE’s (eg, γ-aminobutyric acid and glutathione) and the separation of Glu and Gln is feasible (22,23). Up to 18 metabolites can be quantified at short TE’s and field strengths of 7.0 or 9.4 T (23–25).

A subset of the metabolites detectable by using MR spectroscopy may serve as biomarkers in the context of physiologic and pathologic states. For at least one MR spectroscopy–detected metabolite, NAA, evidence from cell (26), ex vivo brain (27), and histologic studies (28) show unequivocally that, in the mature CNS, NAA is present only in neurons, axons, and dendrites—not in glial cells. Together with 1H MR spectroscopy results of human brain ex vivo specimens (29) and in vivo data (30), these observations make a strong case that NAA is a biomarker for neuronal integrity. In addition, NAA levels may reflect mitochondrial (dys)function (31). tNAA (comprised primarily of NAA, with a small contribution from N-acetylaspartylglutamate) is therefore commonly used as a positive or negative in vivo biomarker either for the presence of viable neurons or the assessment of parenchymal damage. Elevated mlns is generally considered a marker for gliosis (32,33), and high tCho may be a marker for cellular proliferation, increased membrane turnover, or inflammation (13,29,34,35). Elevated Lac is indicative of anaerobic glycolysis and may be considered an unspecific MR spectroscopy biomarker for several abnormalities (36,37).

**MR Spectroscopy of CNS Disorders**

Neurologic diseases affect as many as 1 billion people worldwide and are a major cause of disability and human suffering. Diagnosis is often complex, and the time window for effective therapy may be limited. MR imaging, with its excellent
Soft-tissue contrast, is commonly the modality of choice for the detection of brain lesions. The morphologic details and the sensitivity to changes in content and physical properties of water are exquisite. However, conventional MR imaging is not able to depict changes in cell density, cell type, or biochemical composition—all of which can be investigated with MR spectroscopy. Furthermore, lesions of different underlying pathophysiology often manifest with a similar MR imaging appearance. Accordingly, MR imaging and MR spectroscopy are complementary tools for diagnosing disease and monitoring disease progression and response to therapy.

In the next sections, we will first report on the clinical impact of 1H MR spectroscopy in the evaluation of diseases in which it has already been demonstrated to be valuable and next on the potential clinical utility of MR spectroscopy in disorders where substantial research activity has occurred in the past 2 decades with consistent results across laboratories. The breakdown is based on (a) the demonstration of improved diagnostic accuracy of MR spectroscopy over other commonly used clinical imaging modalities, (b) the presence of disease-linked specific metabolites in the 1H MR spectrum, and (c) the demonstration of reduced need for invasive diagnostic procedures. In general, the “patient-ready” applications involve large disease effects detectable in an individual MR spectrum, whereas disorders for which 1H MR spectroscopy is expected to contribute to future patient management involve subtle spectroscopic changes that are more challenging to detect in individual cases. Table 1 summarizes the CNS disorder entities that are covered herein and lists metabolites of interest for these disorders.

### Neurologic Diseases in Which 1H MR Spectroscopy Is Valuable for Clinical Decision Making

#### Brain Tumors

Clinical decision making in neuro-oncology is achieved by a multidisciplinary team combining information from many sources, including MR imaging. Although it plays a central role in the clinical management of patients with brain tumors, MR imaging alone cannot provide the answer to many important clinical questions. These include differentiating tumor from other focal lesions (e.g., demyelinating plaques, encephalitis), obtaining a definitive diagnosis of atypical ring-enhancing focal lesions (e.g., high-grade gliomas, metastasis, lymphoma, and abscess), identifying the optimal biopsy sites in heterogeneous gliomas, monitoring the response to treatment, and differentiating between treatment-induced changes and recurrent tumor. MR spectroscopy can provide information in all of these key clinical areas, and it is increasingly being used as an adjunct to MR imaging.

The earliest reports in human brain tumors (6), together with work in ex vivo specimens (39,40) and cancer cells (41), demonstrated that MR spectroscopy offers great potential for noninvasive assessment of brain neoplasms. For example, MR spectroscopy in conjunction with perfusion imaging provided a sensitivity of 72% and a specificity of 92% in the differentiation of tumors from nonneoplastic lesions (42). Similarly, a sensitivity of 93% and a specificity of 60% were achieved when using these two methods for identifying high-versus low-grade gliomas, a substantial improvement in sensitivity over that with conventional MR imaging (43).

Large multicenter studies have determined the accuracy of single-voxel MR spectroscopy with pattern recognition algorithms for diagnosing brain tumor histology and grade (44–46). Short TE MR spectroscopy gives an accuracy of approximately 90% for all pairwise comparisons of the main adult tumor types (meningiomas, low-grade glioma, glioblastoma multiforme, metastases) except for glioblastoma multiforme versus metastasis, where the accuracy was 78% (44,46). Combining short and long TE MR spectroscopy gives a diagnostic accuracy for the main childhood brain tumor types (pilocytic astrocytoma, medulloblastoma, and ependymoma) of 98% (45). More recently, MR spectroscopy helped identify molecular subtypes of gliomas with isocitrate dehydrogenase mutations, an example of molecular fingerprinting in vivo, on the basis of levels of 2-hydroxylglutarate (47). Few studies compared the diagnostic accuracy of MR spectroscopy with that of conventional MR imaging, but one study established added value for a decision support system constructed from multicenter data. Namely, 1H MR spectroscopy data improved low- and high-grade tumor prediction relative to MR imaging alone; the area under the receiver operating characteristic curve for low-grade tumors was 0.93 for MR imaging plus MR spectroscopy versus 0.81 for MR imaging alone, and the area under the receiver operating characteristic curve for high-grade tumors was 0.93 for MR imaging plus MR spectroscopy versus 0.85 for MR imaging alone (48). Elevated tCho along with decreased tNAA is typically regarded as a diagnostic feature of brain tumors (13) (Fig 2). In addition, the prominent signal at 1.3 ppm, which arises from lipids present in cytoplasmic droplets associated with necrosis or hypoxia, is generally associated with higher grade and poor survival (49–51) (Fig 2). Conversely, nonneoplastic lesions such as abscesses and tuberculomas often demonstrate elevated amino acids and lipids (52). Other metabolites observed in brain neoplasms include taurine in primitive neuroectodermal tumor (53), alanine in meningiomas (13), and glycine in high-grade pediatric tumors (54). If biopsy is needed for diagnosis, the tCho/tNAA ratio can help differentiate areas of solid tumor with the highest cell density from edema (53,56). The detection of an increased tCho/tNAA ratio in the peritumoral region further reflects tumor invasiveness and can thus be used to differentiate high-grade gliomas from brain metastases that exhibit a near-normal spectrum in the peritumoral region (57,58). MR spectroscopy has also been shown to have a decisive role in the diagnosis of low-grade versus high-grade tumors, as well as in the diagnosis of metastasis versus high-grade tumors, as part of a diagnostic work-up that includes conventional MR imaging.
<table>
<thead>
<tr>
<th>Disorder and MR Spectroscopy Method*</th>
<th>Location of VOI or ROI†</th>
<th>Metabolites of Interest‡</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Suspected tumor &gt;10 mL at gadolinium-enhanced T1-weighted MR imaging or FLAIR MR imaging</strong></td>
<td>VOI on the contrast-enhancing region of tumor if it exists, avoiding necrotic core, and on FLAIR abnormality for nonenhancing tumors</td>
<td>tNAA, tCho, tCr, Lac, mIns, lipids</td>
</tr>
<tr>
<td>SVS, STE or LTE PRESS, STE STEAM</td>
<td></td>
<td></td>
</tr>
<tr>
<td>LTE MRSI</td>
<td>VOI or ROI on the contrast-enhancing region of tumor if it exists, avoiding necrotic core, and on FLAIR abnormality for nonenhancing tumors</td>
<td>tNAA, tCho, tCr, Lac</td>
</tr>
<tr>
<td><strong>Suspected tumor &lt;10 mL at gadolinium-enhanced T1-weighted MR imaging or FLAIR MR imaging</strong></td>
<td>VOI or ROI on the contrast-enhancing region of tumor if it exists, avoiding necrotic core, and on FLAIR abnormality for nonenhancing tumors</td>
<td>tNAA, tCho, tCr, Lac, lipids</td>
</tr>
<tr>
<td>STE or LTE MRSI</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Suspected infective focal lesion</strong></td>
<td>VOI within the lesion</td>
<td>Ac, Suc, Lac, lipids, amino acids (Ala, Leu, Isoleu, Val)</td>
</tr>
<tr>
<td>SVS, STE or LTE PRESS</td>
<td></td>
<td></td>
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<tr>
<td><strong>Suspected metabolic disorder</strong></td>
<td>VOI according to metabolic disorder, e.g., in parietal cortex for Cr deficiencies, in basal ganglia for Leigh disease, white matter in most cases</td>
<td>tNAA, tCr, Lac, Gly, Ala, Pyr, Suc</td>
</tr>
<tr>
<td>SVS, STE or LTE PRESS, STE STEAM</td>
<td>VOI according to metabolic disorder, e.g., in parietal cortex for Cr deficiencies, in basal ganglia for Leigh disease, white matter in most cases</td>
<td>tNAA, tCr, Lac, Gly, Ala, Pyr, Suc</td>
</tr>
<tr>
<td>LTE MRSI</td>
<td>VOI within the lesion</td>
<td>Ac, Suc, Lac, lipids, amino acids (Ala, Leu, Isoleu, Val)</td>
</tr>
<tr>
<td><strong>Neonatal hypoxia-ischemia</strong></td>
<td>VOI within the lesion</td>
<td>tNAA, tCr, Lac, MM</td>
</tr>
<tr>
<td>SVS, STE or LTE PRESS, STE STEAM</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Suspected demyelinating disorder</strong></td>
<td>VOI in T2 hyperintense white matter lesion</td>
<td>tNAA, tCr, tCho, mIns, MM</td>
</tr>
<tr>
<td>SVS, STE or LTE PRESS</td>
<td></td>
<td></td>
</tr>
<tr>
<td>LTE MRSI</td>
<td>Section covering T2 hyperintense white matter lesion</td>
<td>tNAA, tCr, tCho</td>
</tr>
<tr>
<td><strong>Multiple sclerosis</strong></td>
<td>VOI in T2 hyperintense white matter lesion</td>
<td>tNAA, tCr, tCho, mIns, MM</td>
</tr>
<tr>
<td>SVS, STE or LTE PRESS</td>
<td></td>
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<tr>
<td>LTE MRSI</td>
<td>Section covering white matter including corpus callosum</td>
<td>tNAA, tCr, tCho</td>
</tr>
<tr>
<td><strong>Suspected dementia</strong></td>
<td>VOIs in posterior cingulate and mesial temporal lobes</td>
<td>tNAA, tCr, tCho, mIns, Glx</td>
</tr>
<tr>
<td>SVS, STE PRESS, STE STEAM</td>
<td></td>
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<tr>
<td>LTE MRSI</td>
<td>Section angulated along planum temporale and above the lateral ventricles</td>
<td>tNAA, tCr, tCho, (mIns)</td>
</tr>
<tr>
<td><strong>Focal epilepsy</strong></td>
<td>VOI best defined by clinical data</td>
<td>tNAA, tCr, tCho</td>
</tr>
<tr>
<td>LTE MRSI</td>
<td></td>
<td></td>
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<tr>
<td><strong>Mesial temporal lobe epilepsy</strong></td>
<td>VOIs in mesial temporal structures, planum temporale angulation</td>
<td>tNAA, tCr, tCho</td>
</tr>
<tr>
<td>SVS, STE or LTE PRESS</td>
<td></td>
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<tr>
<td><strong>Ischemic lesion</strong></td>
<td>VOI within reduced diffusion volume</td>
<td>tNAA, tCr, tCho</td>
</tr>
<tr>
<td>SVS, STE or LTE PRESS</td>
<td></td>
<td></td>
</tr>
<tr>
<td>LTE MRSI</td>
<td>Section through reduced diffusion volume</td>
<td>tNAA, tCr, tCho</td>
</tr>
</tbody>
</table>

Note.—FLAIR = fluid-attenuated inversion recovery.
* Clinically available MR spectroscopy methods that have been widely used for the abnormalities indicated are listed. Short TE is 25–35 msec for PRESS sequence and 20 msec for STEAM sequence. Long TE is 135–270 msec typically only used for single-voxel spectroscopy. 144 msec is used for Lac inversion, however, precaution is required about chemical shift displacement errors at 3.0 T (38). LTE = long TE, MRSI = MR spectroscopic imaging, STE = short TE, SVS = single-voxel spectroscopy.
† ROI = region of interest, VOI = volume of interest.
‡ Note that mIns, a combination of Glu and Gln (Glx), macromolecules (MM), and lipids are detected reliably with short TE sequences, whereas glycine (Gly) is detected reliably with long TE sequences. Ac = acetate, Ala = alanine, Isoleu = isoleucine, Leu = leucine, Pyr = pyruvate, Suc = succinate, Val = valine.
with gadolinium and diffusion-weighted and perfusion MR imaging (59).

MR spectroscopy may be used to determine prognosis and to guide treatment planning in oncology patients when surgery is not indicated, such as in diffuse brainstem gliomas and intramedullary tumors in the spinal cord (60). A tCho/tNAA peak amplitude ratio of at least 2.1 (either at single-voxel spectroscopy with a TE of 144 or 270 msec or at MR spectroscopic imaging with a TE of 280 msec) was found prognostic of unfavorable outcome in pediatric diffuse pontine gliomas (61). Prognostic MR spectroscopy markers are important for treatment stratification and can help identify patients who need more intensive treatment from the outset for some tumor types (47,62,63). These include the detection of 2-hydroxyglutarate in isocitrinate dehydrogenase-1 mutated gliomas (47), citrate in proliferating pediatric astrocytomas (62), and highly MR spectroscopy–visible saturated lipids with elevated scyllo-inositol and low glutamine in high-risk pediatric brain tumors (64). A tCho/tNAA ratio of more than 2.1 at long TE MR spectroscopic imaging has been used to identify regions of more aggressive phenotype within a heterogeneous glioblastoma multiforme to improve gamma knife radiosurgery (64).

For neurosurgical treatment planning, MR spectroscopy plays a role in differentiating areas of tumor from benign processes and, together with other MR imaging methods, in establishing their relationship to key normal brain structures (56), particularly in gliomas. Infiltrative gliomas extend well beyond the T2-defined main tumor bulk. One study reported that the MR spectroscopy–defined abnormal area was an average of 24% larger than that delineated by T2 hyperintensity and confirmed the accuracy of an elevated tCho/tNAA ratio with histologic and immunohistochemistry findings for tumor cells (65). Another study demonstrated increased mlns and Glu levels in the contralateral hemisphere of patients with untreated glioblastoma multiforme, a finding that was indicative of early neoplastic infiltration (66). In addition, gliomas of all grades may have intratumoral heterogeneity (67), sometimes even despite apparent homogeneous imaging characteristics. It is common to find low-grade oligodendrogliomas with malignant imaging features, nonenhancing high-grade gliomas with benign imaging features, and focal areas of malignancy in low-grade gliomas. In low-grade gliomas, detection of areas with infiltrative tumor cells (close or distant to the main mass) is very important as these can be the primary sites of tumor recurrence. Delineation of tumor infiltration is an essential part of (a) preoperative decision making, (b) intraoperative MR imaging–guided resections, and (c) postoperative follow-up and application of additional therapies (post-surgery radiation and/or chemotherapy). MR spectroscopy was shown to spatially correlate with histologic type and grade and to reflect heterogeneity in brain tumors before surgery: A tCho/tNAA ratio greater than 2, a Lac/tNAA ratio greater than 0.25, and the presence of lipid at MR spectroscopic imaging with a long TE (144 msec) are characteristics of a high-grade tumor, allowing demarcation
of brain parenchyma adjacent to MR imaging–delineated tumor (56). In addition, recent intraoperative $^1$H MR spectroscopy at 3.0 T helped differentiate tumor from a nontumoral abnormality, as indicated by a high tCho/tCr ratio and the presence of Lac, in 57% of suspected cases and had a positive effect on surgical success and patient outcome (68).

MR spectroscopy can help avoid the incorrect diagnosis of tumor progression, which can lead to inappropriate surgery, other treatment, and patient distress in cases of posttreatment-induced changes that are ambiguous at conventional MR imaging. For example, the tCho/tNAA ratio was shown to reliably differentiate recurrent glioma from postradiation injury (69) (Fig 3). Similarly, MR spectroscopy (tCho/water), either alone or in combination with conventional MR imaging, can further contribute to the assessment of response to anticancer treatment (70).

MR spectroscopy (tCho/tCr and tCho/tNAA) and dynamic susceptibility contrast MR imaging was reported to have 100% positive and negative predictive values for discriminating posttreatment change, which is more accurate than both conventional MR imaging (positive predictive value, 50%) and fluorine 18 deoxyglucose positron emission tomography (PET) (positive predictive value, 67%; negative predictive value, 60%) (72). However, dynamic susceptibility contrast MR imaging showed a substantial false-positive rate, which was not the case with MR spectroscopy—a finding that points to an incremental value of MR spectroscopy in separating tumor recurrence and posttreatment injury (72).

In summary, MR spectroscopy adds diagnostic and prognostic benefits to MR imaging and aids in treatment planning and monitoring of brain cancers.

**Pediatric Disorders: Hypoxia-Ischemia, Inherited Metabolic Diseases, and Traumatic Brain Injury**

$^1$H MR spectroscopy was used for pediatric brain imaging as early as 1990–1991 (73–75), and it is part of routine imaging protocols in many specialized academic health centers and children’s hospitals. For the newborn infant, quantitative assessment of cerebral Lac due to hypoxia-ischemia is one of the earliest imaging signs indicative of clinical brain injury (37,76) (Fig 4), and persistence of high Lac is associated with poor outcome (77). MR spectroscopy can be used as a means to assess treatment efficacy of hypothermia, a proven neuroprotective treatment for perinatal asphyxia (78).

Although rare, inherited metabolic disorders are a significant disease entity in neuropaediatrics. Clinical symptoms in certain inherited metabolic diseases are due to the accumulation of metabolites that are either neurotoxic or interfere with normal function. If the accumulating substance is visible at MR spectroscopy, its presence or elevation in the spectrum can be used for diagnosis. MR spectroscopy has proved clinically useful in neonates suspected of having metabolic disorders (79–81) owing to the unique ability to noninvasively detect the metabolic defect in vivo (82–85). For example, the presence of pyruvate (plus Lac and/or alanine) and succinate are early indicators of pyruvate and succinate dehydrogenase complex deficiencies, respectively (79,86–88). Detection of elevated glycine, in particular at long TEs, is clinically diagnostic in nonketotic hyperglycinemia (82), although intracerebral hemorrhage presents a confound in the interpretation of high glycine levels (89). A grossly elevated tNAA level is a diagnostic hallmark of Canavan disease (90).

In other inherited diseases, the reduction of metabolites owing to
For effective clinical management, objective means to evaluate long-term outcome are required, especially for comatose patients. In a cohort of children with traumatic brain injury, a regression model, incorporating age, initial Glasgow coma scale, and presence of retinal hemorrhage and supplemented with tNAA/tCr ratio and MR spectroscopy–visible Lac within the 1st month after incidence, was shown to differentiate between good and poor outcomes (102). In pediatric near-drowning accidents, an MR spectroscopy index based on tNAA, Lac, and combined Glu and Gln was shown to correctly differentiate between good and poor outcomes—with no false-positive results (103).

These data support the clinical utility of MR spectroscopy in combination with MR imaging to provide insights into cerebral metabolism and viability. However, the interpretation of MR spectroscopy data requires careful consideration of the limitations associated with the presence of significant background noise and the varying sensitivity of different metabolites across the frequency spectrum. Further research is needed to standardize the acquisition and analysis protocols to enable more accurate and robust clinical application of MR spectroscopy in the pediatric population.
with clinical measures for predicting outcome.

**Demyelinating Diseases**

MR spectroscopy plays an important role (104) or in addition to other semiquantitative MR techniques (105) in the differential diagnosis of hereditary leukoencephalopathies. MR spectroscopy provides valuable information about tissue pathophysiology for at least three different metabolic profiles: (a) hypomyelination, (b) white matter rarefaction, and (c) demyelination, which were differentiated with tCho/tCr and tNAA/tCr ratios in a study of 70 children (104).

Hematopoietic stem cell transplantation is currently the only treatment option for inherited demyelinating disorders such as X-linked adrenoleukodystrophy, metachromatic leukodystrophy, and globoid cell leukodystrophy (106). MR spectroscopy is used to monitor the onset of demyelination in neurologically asymptomatic patients with X-linked adrenoleukodystrophy with high genotypic variability (14,32,107). Interval elevation of mlNs/tNAA and tCho/tNAA ratios in normal-appearing white matter at follow-up is an indication for treatment with hematopoietic stem cell transplantation (Fig 6). Hematopoietic stem cell transplantation performed before substantial tissue degeneration as assessed with tNAA results in clinical stabilization (108). In patients who are newly diagnosed with juvenile or adult metachromatic leukodystrophy, a combination of MR imaging and MR spectroscopy can be used to judge the state of brain tissue inflammation (109,110). Although mlNs is typically increased even in the early stages of metachromatic leukodystrophy, as long as tNAA is still within the normal range, hematopoietic stem cell transplantation is indicated (111,112).

The clinical use of MR spectroscopy in multiple sclerosis, an acquired demyelinating disease, remains limited despite the various insights into disease pathology that it has offered as well as its ability to assess the burden of axonal damage (113). MR spectroscopy of chronic multiple sclerosis plaques in white matter shows a consistently reduced tNAA/tCr ratio (5,35) and, sometimes, an elevated tCho/tCr ratio (35). Spectra from plaques undergoing active inflammation show an elevated tCho/tCr ratio, normal or reduced tNAA/tCr ratio (35), and elevated macromolecular signals, possibly arising from myelin breakdown products (114). The tNAA/tCr ratio in the normal-appearing white matter of patients with varying clinical presentations helps differentiate patients from healthy control subjects (115,116) and inversely correlates with disability scores—especially at an early stage (117). In addition, the tCho/tCr ratio is elevated in normal-appearing white matter months before lesions become detectable at conventional MR imaging (118). These observations underscore the ability of MR spectroscopy to characterize white matter abnormality in evolving multiple sclerosis (119). In addition, increasing evidence for gray matter involvement in multiple sclerosis (120) provides motivation to study these lesions with MR spectroscopy as well (113). Finally, MR spectroscopic imaging might play an important role in the differential diagnosis of multiple sclerosis, with acute disseminated encephalomyelitis showing recovery of tNAA signal losses as a favorable prognostic sign (121).

**Focal Lesions Caused by Infectious Agents**

Brain infections can be life threatening and, hence, require an early diagnosis for optimal clinical management. Definitive laboratory diagnostic tests can be time consuming, thus delaying therapy. MR spectroscopy is valuable in the differential diagnosis of intracranial ring-enhancing lesions. When a ring-enhancing mass lesion manifests with nonspecific clinical and conventional MR imaging features, the ability of MR spectroscopy to characterize white matter abnormality in multiple sclerosis, with acute disseminated encephalomyelitis showing recovery of tNAA signal losses as a favorable prognostic sign (121).
typically localized to the region(s) affected by the degenerative process (124,125). The tNAA levels reflect pathologic severity (33,126) (Fig 8) and correlate with clinical measures in cross-sectional studies (127,128). Consistently, tNAA/tCr tends to be lower in subjects with mild cognitive impairment who convert to dementia compared with those who remain stable (129). Therefore, the tNAA/tCr ratio or tNAA concentration may be a valuable prognostic indicator of disease progression, either alone or in combination with volumetric measurements (130).

Other 1H MR spectroscopy changes associated with neurodegeneration include a decreased Glu level (128,131,132), an elevated tCho level (125), and an elevated mIns level (132,133). The elevation in mIns may be associated with glial or microglial activation, a characteristic feature of these diseases (134). An elevated mIns level appears early in dementia, preceding the decrease in tNAA concentration (Fig 8), atrophy, and associated neuronal loss and cognitive impairment, as demonstrated in presymptomatic carriers for familial Alzheimer disease (135) and in patients with frontotemporal lobar degeneration mutations (136).

1H MR spectroscopy may also be used to monitor treatment response in neurodegenerative diseases. For example, a transient increase in tNAA concentration was associated with short-term functional response during donepezil treatment in Alzheimer disease, suggesting that tNAA also reflects functional integrity and recovery (137). Other studies have shown a decreased mIns/tCr ratio following donepezil treatment (138) and an increased Glu level after galantamine treatment for Alzheimer disease (139).

Epilepsy

Epilepsy is a common disorder, with a prevalence of 0.5%–1.0% worldwide. The specific etiology underlying the seizures can be variable, with 60%–70% of all patients responding to medications (140,141). Surgical intervention can be effective in the remaining 30%–40% of
patients (142,143). In the more common type of focal epilepsy, surgical outcomes are improved if the region of seizure onset can be clearly defined (142,143). Conventional MR imaging can accurately localize the seizure onset region, for example, by identifying unilateral hippocampal atrophy or malformations of cortical development. However, MR imaging may often be negative or ambiguous (eg, bilateral involvement) and, in some cases, lesions seen at MR imaging may not match the focus of seizure onset identified by means of invasive electroencephalographic measurements.

Given the close physiologic relationship between brain function and metabolism (144), MR spectroscopy has been extensively used to better understand and localize human epilepsy (145,146). Abnormalities in tNAA concentration and the tNAA/tCr ratio have been useful for detecting injured brain in the seizure onset focus (145–149). MR spectroscopic imaging measures have also been extended to neurotransmitters, for example, to assess γ-aminobutyric acid in patients with epilepsy at ultra-high field strengths (150).

The most common abnormality in temporal lobe epilepsy is mesial temporal sclerosis, which may often be effectively treated with unilateral temporal lobectomy. Multimodal evaluation, which involves scalp or intracranial electroencephalography, conventional MR imaging, and/or metabolic imaging with PET, is commonly used to lateralize the epileptogenic zone in mesial temporal sclerosis. A meta-analysis of 1H MR spectroscopy literature comprising 22 studies.
Acute Stroke and Brain Ischemia
Overall, MR imaging plays a limited role in decision making for clinical management of patients with acute stroke,
usually because of a lack of immediate availability of the imaging unit and of patient-related MR imaging safety information. The decision to thrombolise or to apply any other form of therapeutic intervention in the hyperacute phase is based on clinical grounds and exclusively involves computed tomography to rule out either brain hemorrhage or very large ischemic lesions, which usually have unfavorable outcomes (155). Diffusion-weighted and perfusion MR imaging are superior imaging techniques for detecting acute ischemia and highlight the penumbra, but they are rarely used outside of specialized acute stroke clinics that have rapid access to MR imaging. Similarly, 1H MR spectroscopy offers great potential after the hyperacute phase of stroke (beyond 4.5 hours) to assess several key characteristics of ischemic brain for prognostic purposes, such as severity of ischemia and neuronal dysfunction and damage.

Preclinical work has shown that tNAA decreases in ischemic brain parenchyma in a linear fashion for the first 6 hours, followed by a slower decrease for the subsequent 24 hours (156,157). Therefore, quantitative metabolite data for tNAA and Lac are of value for evaluating the nature of ischemia and predicting risk for new ischemic events (161).

### Technical Considerations

#### Data Acquisition

Any application of MR spectroscopy to a clinical question starts with the decision about a pulse sequence and parameters. In general, this choice is dictated by the disease (Table 2). When the affected brain region is well defined, single-voxel spectroscopy is the preferred method and provides robust metabolite quantification in the selected volume of interest, whereas MR spectroscopic imaging is the method of choice in diseases where the focal point of pathology is unclear, if there are multiple lesions, or if the lesions are heterogeneous. For example, MR spectroscopic imaging is advantageous in the accurate evaluation of tissue status in localization-related epilepsy (Table 1) and in the investigation of the heterogeneity of large tumors (67). In
many abnormalities, single-voxel spectroscopy and MR spectroscopic imaging can be used in combination; for example, MR spectroscopic imaging to first identify the lesion location and single-voxel spectroscopy to quantify metabolites that can be reliably obtained from high-quality, short TE spectra in the identified lesion (Table 1).

All of the major clinical MR imaging vendors provide MR spectroscopy protocols, primarily with use of the basic PRESS (1,2) and STEAM (3) sequences (Table 1). In addition, other state-of-the-art single-voxel spectroscopy and MR spectroscopic imaging sequences, which offer various advantages over the basic STEAM and PRESS sequences, have been implemented on some clinical platforms (Appendix E1 [online]).

Which field strength is optimal for a particular clinical application of MR spectroscopy is another important question for the practicing neuroradiologist and clinical trialist. Although 3.0 T is becoming the preferred platform over 1.5-T for MR spectroscopy owing to potential gains in SNR and spectral resolution, it is important to note that field strength is not the sole determinant of the information content of spectra. In fact, a spectrum obtained at 1.5 T with a protocol adhering to spectral quality standards (Appendix E1 [online], Fig 9) provides more reliable metabolite information than a poor-quality spectrum obtained at 3.0 T. Overall, clinical MR spectroscopy can be successfully performed at either 1.5 or 3.0 T for the majority of applications. Although the potential gains at magnetic fields higher than 3.0 T for clinical MR spectroscopy are still being assessed, significant improvements in spectral and spatial resolution at 7.0 T have been reported. For example, previously inaccessible alterations in low-concentration metabolites may be uncovered at 7.0 T (162). For MR spectroscopic imaging, the nominal spatial resolution can be reduced to 0.14 mL at 7.0 T (163).

Finally, the importance of spectral quality generated with the chosen pulse sequence, parameters, and field strength cannot be underestimated. For reliable clinical decision making based on MR spectroscopy data, obtaining high-quality, artifact-free spectra is crucial. The sources and forms of artifacts in MR spectra have been reviewed in detail (164) and are summarized in Appendix E1 (online). The detection of such artifacts and exclusion of spectra based on predefined quality criteria relies on the human expert in most applications of single-voxel spectroscopy, whereas automated quality assessment of MR spectroscopic imaging data is preferred. A practical guide to determine whether a spectrum is adequate for clinical use is provided in Figure 9. Further considerations regarding the choices for clinical MR spectroscopy data acquisition, including pulse sequence, parameters, field strength, and radiofrequency coils, as well as recommendations for spectral quality assessments, are detailed in Appendix E1 (online).

Data Analysis and Reporting
All clinical imaging units provide MR spectroscopy analysis software, which can be used for visual inspection of spectra and basic quantification of metabolite ratios. In addition, off-line postprocessing tools (165) and sophisticated quantification packages such as LCModel (166) are widely used. These packages provide quantitative error estimates for metabolite quantification (eg, Cramér–Rao lower bounds), with which the reliability of metabolite concentrations can be assessed (see Appendix E1 [online] for recommended criteria). The availability of error estimates is an important requirement for clinical decision making when using quantitative MR spectroscopy measures; therefore, vendors of clinical imaging units are encouraged to implement more robust, U.S. Food and Drug Administration–approved MR spectroscopy analysis packages that provide such quantitative error estimates.

For clinical use, single-voxel spectroscopy data can be reported numerically as metabolite concentrations or as ratios, ideally supplemented with visualization of volume of interest placement (167). On the other hand, information from two- or three-dimensional MR spectroscopic imaging must be made available to the clinician in a quick and easy image format to incorporate into the clinical routine. In addition, implementation of MR spectroscopy into picture archiving and communication systems is recommended to facilitate easy access to MR spectroscopy data in the standard work environment.

Reproducibility and Clinical Translation
Ultimately, test-retest reproducibility of measured metabolite levels determines the utility of MR spectroscopy for disease assessment. To be of clinical value, experimental and biologic variability in the quantified metabolite levels must be smaller than their changes caused by disease. Test-retest coefficients of variance reported at 1.5 and 3.0 T (168–173) show improved accuracy for several metabolites at higher fields and shorter TEs. Test-retest coefficients of variance of 6% or less are achievable for five metabolites (tNAA, tCr, tCho, mlNs, Glu) with single-voxel

<table>
<thead>
<tr>
<th>Figure 9</th>
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<tr>
<td>✓ SNR &gt; 3 for major resonances such as high tCho and low tNAA in tumors; SNR &gt; 2 for detection only of important indicator metabolites such as lactate</td>
</tr>
<tr>
<td>✓ Spectral resolution: FWHM of metabolites &lt; 0.1 ppm</td>
</tr>
<tr>
<td>✓ Line shape: symmetric</td>
</tr>
<tr>
<td>✓ Water suppression &gt; 98%</td>
</tr>
<tr>
<td>✓ No lipid contamination from the scalp</td>
</tr>
<tr>
<td>✓ Artifacts (chemical shift artifact, ghosting, patient motion, eddy currents, volume averaging) are absent or minor</td>
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that identical and optimized acquisition protocols and calibration schemes are used (Fig 10).

When it is desired to use clinical MR spectroscopy data to make decisions affecting management, it is essential that an adequate cohort of subjects has been studied such that the classifier used is robust in terms of both sensitivity and specificity. Recommended cohort size will depend somewhat on the nature of the data, but anything less than several hundred subjects, both healthy subjects and those with the condition of interest, would not yield a classifier that would be certified for use by a regulatory agency. A detailed discussion of these issues has appeared recently (176,177).

In addition, validation of MR spectroscopy biomarkers for clinical use requires their incorporation in robust prospective multicenter clinical trials, where patient selection and treatment meets prespecified criteria and the statistical methodology is set before the trial commences. This requires careful MR spectroscopy protocol design that can be adhered to at all the participating centers. In addition, effective, real-time quality control measures must be put in place to ensure that data that need to be discarded are kept to a minimum to avoid bias and ensure generalizability of the results.

Appendix E1 (online) highlights further recommendations to facilitate translation of MR spectroscopy to routine use in the clinical environment, including steps that must be taken for integration with clinical imaging and for quality management in single- and multisite studies (Fig 10) and a discussion on reimbursement issues, a frequently cited impediment to the widespread use of clinical MR spectroscopy.

Conclusions and Recommendations

In conclusion, MR spectroscopy is used worldwide as an adjunct to MR imaging in several common neurologic diseases, including brain neoplasms, inherited metabolic disorders, demyelinating disorders, and infective focal lesions. The spectrum of disorders for which spectroscopy at 3.0 T (174), and coefficients of variance less than 10% were reported for tNAA, tCr, tCho, and mIns with MR spectroscopic imaging at 3.0 T (172,173). Importantly, standard clinical hardware generates reproducible MR spectroscopy data from the human brain in a multicenter setting provided.
MR spectroscopy will be clinically used is likely to expand; potential examples include neurodegenerative diseases and epilepsy. The standardization of MR spectroscopy data acquisition and analysis techniques for clinical use is encouraged, along with the publication of normative data obtained with these techniques. Multicenter trials are encouraged to establish the utility of MR spectroscopy in large enough sample sizes to definitively establish the value of MR spectroscopy in specific clinical applications. Where possible, these should include assessment of the impact on clinical outcome and economic benefit. Clinical imaging centers specializing in combined use of MR imaging and spectroscopy should be established in all major clinical neurologic centers that offer standardized MR spectroscopy procedures for improved patient management. Manufacturers of MR units and third-party companies (eg, vendors of analysis software) are encouraged to continue to develop their products to incorporate recent technical advances, to obtain U.S. Food and Drug Administration approval for clinical use, and to provide products with manufacturer-independent standardized outputs.

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