Dynamics in the Interior of AOT Lamellae Investigated with Two-Dimensional Infrared Spectroscopy

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ABSTRACT: The dynamics inside the organic regions of aerosol-OT (AOT)/water mixtures in the lamellar mesophase, bicontinuous cubic (BC) phase, and in an analogous molecule without the charged sulfonate headgroup are investigated by observing spectral diffusion, orientational relaxation and population relaxation using ultrafast two-dimensional infrared (2D IR) vibrational echo spectroscopy and IR pump–probe experiments on the asymmetric CO stretch of a vibrational probe, tungsten hexacarbonyl (W(CO)6). The water layer thickness between the bilayer planes in the lamellar phase was varied. For comparison, the dynamics of W(CO)6 in the normal liquid bis(2-ethylhexyl) succinate (EHS), which is analogous to AOT but has no charged sulfonate headgroup, were also studied. The 2D IR experiments measure spectral diffusion, which results from the structural evolution of the system. Spectral diffusion is quantified by the frequency–frequency correlation function (FFCF). In addition to a homogeneous component, the FFCFs are biexponential decays with fast and slow time components of \( \sim 12.5 \) and \( \sim 150 \) ps in the lamellar phase. Both components of the FFCF are independent of the number of water molecules per headgroup for the lamellae, but they slow somewhat in the BC phase. The dynamics in the ordered phases are in sharp contrast to the dynamics in EHS, which displays fast and slow components of the FFCF of \( 5 \) and \( 80 \) ps, respectively. As the hydration level of AOT increases, vibrational lifetime decreases, suggesting some change in the local environment of W(CO)6 with water content.

1. INTRODUCTION

Mesomorphic phases appear in many surfactant systems and with increasing surfactant concentration. For example, the surfactant AOT (aerosol-OT, sodium bis(2-ethylhexyl) sulfosuccinate) in mixtures with water with increasing AOT concentration at room temperature goes through the phases: isotropic, lamellar, bicontinuous cubic (BC), and inverse hexagonal.1 The lamellar structure occurring in surfactant systems has been shown by a variety of techniques as being composed of macroscopically extended bilayers of amphiphilic molecules alternating with water layers.2,3 Lyotropic lamellar phases are important as models of logical membranes4–6 and in various scientific and technological applications.7,8 Biological membranes have a basic bilayer structure. In addition to a permeability barrier, the plasma membrane serves as the "solvent" for membrane proteins and other biological molecules. Therefore, the nature of the internal dynamics of lamellar structures (bilayers) is important for understanding processes that occur within them.9–13 The BC phase has an interesting thermodynamically stable structure consisting of what is essentially a branching network of surfactant cylinders in three dimensions with congruent networks of water channels.14,15 It has been suggested that the BC phase is an intermediate between the inverted hexagonal phase (parallel cylinders) and the lamellar phase.

AOT gives rise to lamellar structures when mixed with water in a wide range of AOT/water ratios. At room temperature, the AOT lamellae phase occurs in the range from \( \sim 12 \) to \( \sim 78 \) wt% AOT.1 The lamellar repeat distances have been characterized by X-ray diffraction and scale linearly with the reciprocal of the AOT volume fraction.3,16,17 The structure of AOT is shown in Figure 1A. It contains a sulfonate headgroup and a sodium counterion that preferentially partition into the water phase. There are ester moieties near the headgroup that may also interact with the water as well as the sodium counterion depending on the packing and configuration of the AOT molecules.18–21 The alkyl chains are branched and bulky and their steric interactions with one another play an important role in determining the surface area per AOT molecule in lamellar phases and in reverse micelles.17 Various studies have been carried out to investigate the hydration of lipid membranes,5,23–25 and experiments and simulations have shown that water molecules penetrate the interfacial region of bilayers to some extent, but have a low probability of being found in the hydrophobic core.19,26–28 Dynamical studies of the AOT/water system in the lamellar phase have been somewhat sparse. Some of these studies have focused on water diffusion29–31 while others have explored the effect of water/surface interactions and confinement geometry on the dynamics of the water.18,32 X-ray, NMR, and fluorescence experiments have also been used to study the model membranes mainly to study the effect of cholesterol on the depth of water penetration.33–37

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In this paper we investigate the internal dynamics of AOT lamellar structures, the BC phase, and a related model compound. Measurements of spectral diffusion, orientational relaxation, and population relaxation were made using two-dimensional infrared (2D IR) vibrational echo spectroscopy and pump probe experiments. The experiments were performed on a vibrational dynamics probe molecule, the triply degenerate asymmetric CO stretching mode of tungsten hexacarbonyl (W(CO)$_6$) at room temperature. W(CO)$_6$ is completely insoluble in water and therefore resides in the hydrophobic interior of the AOT lamellar and BC structures. The number of water molecules per surfactant is given by the parameter $\lambda = [H_2O]/[AOT]$. The lamellar phase was studied over a wide range of hydration levels from $\lambda = 98$ to 16. The BC phase only occurs in a very narrow range of hydration, and measurements were made with $\lambda = 6$.

To investigate the effect of structural ordering in the mesophases on their dynamics, the same experiments were performed on W(CO)$_6$ in a disordered liquid that is chemically equivalent to the interior of the AOT lamellar and BC structures. The model system is the room temperature liquid bis(2-ethylhexyl) succinate (EHS, C$_{20}$H$_{38}$O$_4$), which has the same chemical structure as AOT but without the charged sulfonate headgroup. The structure of EHS is shown in Figure 1B.

Recently, using ultrafast infrared spectroscopy, dynamics of water between the bilayers in AOT lamellar structures and in the water nanopools in AOT spherical reverse micelles were probed.$^{18,38-42}$ The experiments investigated the influence of interactions with the AOT interface on the dynamics of water. The experiments compared water dynamics in the planar lamellar geometry and the spherical reverse micelle geometry as a function of the number of water molecules per AOT headgroup. In contrast to those experiments on water, the current experiments examine the dynamics in the surfactant interiors in the lamellar and BC mesophases.

The 2D IR experiments measure spectral diffusion, which is the time evolution of the vibrational frequencies of the probe molecule vibration within the inhomogeneously broadened absorption line. Spectral diffusion is the result of the structural evolution of the system that causes the local environments of the probe molecules to change, which in turn changes the probe vibrational frequency. Therefore, spectral diffusion reports on the time dependence of the structural evolution of the system. The 2D IR measurements of spectral diffusion yield the frequency–frequency correlation function (FFCF), which quantifies the dynamic time scales and amplitudes of contributions to the structural changes in the system.$^{33-47}$ The FFCFs obtained from the 2D IR experiments yield a homogeneous (ultrafast structural fluctuations) component and two spectral diffusion components. In the lamellar phase, the spectral diffusion components are independent of the number of water molecules per headgroup ($\lambda$) within experimental error, and have fast and slow component values of $\sim 12.5$ and $\sim 150$ ps, respectively. When the amount of water is reduced, the phase changes to BC, and both components of the spectral diffusion slow ($\sim 15$ and $\sim 180$ ps), showing that the dynamics are sensitive to the structural phase but not sensitive to the amount of water so long as the system remains in the lamellar phase. In contrast to these ordered phases, the two components of the spectral diffusion in the EHS disordered liquid model of AOT are $\sim 5$ and $\sim 80$ ps, which shows that the mesophase structure has a substantial impact on the internal dynamics of the lamellar and BC phases. Although the spectral diffusion does not change with $\lambda$ (number of water molecules between bilayers) in the lamellar phase, the vibrational lifetime does change to some extent, indicating a local change in the environment of the vibrational probe.

2. EXPERIMENTAL PROCEDURES

Succinic anhydride, 2-ethyl-1-hexanol, toluene, and p-toluenesulfonic acid (Sigma-Aldrich) were used as received. Aerosol-OT (AOT, sodium bis(2-ethylhexyl) sulfosuccinate) (Sigma-Aldrich) was purified by dissolving the compound in methanol and stirring overnight with activated charcoal. The charcoal was removed by vacuum filtration, and methanol was removed by rotary evaporation.

Bis(2-ethylhexyl) succinate (EHS), the model compound, was synthesized and purified following a related procedure for the synthesis of a variant of AOT$^{48}$ as follows: 10 g of succinic anhydride and 21 g of 2-ethyl-1-hexanol were dissolved in 250 mL of toluene, and 2.2 g of p-toluenesulfonic acid were added as a catalyst. The reaction mixture was heated to 120 °C for 15 h in a round-bottom flask with an attached Dean–Stark water-trap. The reaction mixture was subsequently washed a few times with aqueous NaHCO$_3$ solution. The organic phase was dried over MgSO$_4$ and the solvent was removed by rotary evaporation. The mixture was then purified using flash column chromatography over silica with a 10–20% EtO$_2$/petroleum ether solvent gradient.

Tungsten hexacarbonyl was dissolved in methylene chloride and added to the surfactant or EHS, such that the ratio of probe molecule to surfactant or to EHS was 1:600. In the surfactant systems, water was added after slow evaporation of methylene chloride overnight. AOT/ water mixtures were made with AOT weight percentages 20, 30, 40, 50, 60, and 80, which correspond to water to surfactant ratios $\lambda = 98$, 57, 37, 24, 16, and 6. The water thickness of lamellar samples of varying water to surfactant ratios has been determined by X-ray diffraction.$^{3,16,17}$ Samples for FT-IR absorption experiments and 2D IR and IR pump–probe experiments were housed in sample cells with CaF$_2$ windows separated by 12 μm Teflon spacers.

![Figure 1. Structures of (A) AOT (aerosol-OT, sodium bis(2-ethylhexyl) sulfosuccinate) and (B) bis(2-ethylhexyl) succinate (EHS).](image-url)
A Ti:sapphire oscillator/regenerative amplifier with a repetition rate of 1 kHz, which produced 100 fs pulses with pulse energies of ~700 μJ and a wavelength centered at 800 nm, was used to pump a home-built mid-IR optical parametric amplifier to generate ~170 fs pulses centered at 1980 cm⁻¹ with ~5 μJ pulse energy. The mid-IR pulses were then co-aligned with a He–Ne beam and a 90:10 ZnSe beam-splitter was used to split the incoming pulse into a weaker probe pulse and a stronger pump pulse. The weak pulse is routed through a mechanical delay line which is used to set the waiting time T₁ (see below). The strong pulse was sent to a mid-IR Fourier-domain pulse-shaper. Details of the laser system and the pulse shaper have been presented.⁴⁹ The output beam from the pulse shaper, which generated pulses 1 and 2 in the vibrational echo experiments, is focused into the sample using a 150 mm focal length CaF₂ lens to give spot size diameters of ~250 μm and ~180 μm, respectively. The probe is collimated after the sample then sent to a spectrometer equipped with a 32-element HgCdTe IR array detector.

In a 2D IR vibrational echo experiment, the time between these pulses is scanned for a fixed T₂ (the time between pulse 2 and 3), and a 2D spectrum is obtained by Fourier transforming along the τ dimension. Then T₁ is changed, and τ is scanned to obtain another 2D spectrum. A series of such spectra is recorded. The desired information is contained in the peak positions and the T₁ dependence of the amplitudes and shapes of the peaks in the 2D spectrum. In the pump–probe experiments, two pulses, a strong pump and a weak probe, impinge on the sample. The time between the two pulses is τ. The pump produces a transient change in the transmission of the probe. The probe is frequency resolved by the spectrometer, and its amplitude at each frequency is measured as a function of τ. The change in probe transmission is obtained with the pump polarization parallel to the probe and at the magic angle of 54.7°.

The FFCF, which quantifies the spectral diffusion in terms of the amplitudes and time scales of the dynamics, is the joint probability that an absorption amplitude at each frequency is measured as a function of τ. The FFCF is described with a multiexponential model, where the Δ are the frequency fluctuation amplitudes of each component, and the τ are their associated time constants. A component of the FFCF with Δτ < 1 is motionally narrowed, and it is the dominant source of the homogeneous broadening of the absorption line. When a component is motionally narrowed, Δ and τ cannot be determined separately. The motionally narrowed homogeneous contribution to the absorption spectrum has a pure dephasing time, which is the pure dephasing time, τₚ. The observed homogeneous dephasing time, T₂, also has contributions from the vibrational lifetime and orientational relaxation.

\[
\frac{1}{T_2} = \frac{1}{T_{2p}} + \frac{1}{2T_{1p}} + \frac{1}{3T_{1r}}
\]

where T₁p, T₂p, and T₁r are the pure dephasing time, vibrational lifetime, and orientational relaxation times, respectively. The total homogeneous line width is \( \Gamma = 1/\pi T_{2p} \).

CLS analysis of the T₁-dependant 2D IR line shapes permits the FFCF to be determined in the following manner. ⁴⁶,⁴⁷ The 2D spectrum at a given T₁ is sliced parallel to the \( \omega_{o} \) axis (the vertical axis in the 2D spectra) over a range of frequencies about the center of the spectrum. Each slice is fit to a Gaussian line shape function to obtain its peak position. The peak positions are plotted against the corresponding \( \omega_{o} \) frequencies of the slices. The set of peak positions (\( \omega_{o} \)) are referred to as center line data, and the resulting line is fit with a linear regression to obtain its slope. Slopes are obtained for each T₂ and plotted. The slopes can range from 1 to 0. A fit to the CLS vs T₂ curve gives the spectral diffusion time constants and amplitude factors. The difference between the T₂ = 0 intercept of the CLS plot and 1 is related to the homogeneous component of the FFCF. To obtain homogeneous component, the CLS fitting results are used to simultaneously fit the FT-IR absorption spectrum and the CLS data. ⁴⁶,⁴⁷ The homogeneous component is included in the full FFCF.

The polarization selective pump–probe technique tracks the decay of the probe transmission with polarizations parallel (Iₚ) and at the magic angle (MA = 54.7°) relative to the pump pulse polarization. The data contain information about both population and orientational dynamics.

\[
I_p = P(t)(1 + 0.8C_2(t))
\]

(3)

\[
I_{MA} = P(t)
\]

(4)

\( P(t) \) is the vibrational population relaxation and \( C_2(t) \) is the second Legendre polynomial correlation function, which is the orientational correlation function for the vibrational mode. Measurements of \( I_{MA} \) enable \( C_2(t) \) to be extracted from measurements of \( I_p \). Here \( C_2(t) \) is a single-exponential decay characterized by the orientational relaxation time, \( \tau_r \).

### 3. RESULTS AND DISCUSSION

#### 3.1. FT-IR Absorption Spectra

\( W(CO)_{6} \) is used as a vibrational spectral probe of dynamics and structure. \( W(CO)_{6} \) is substantially smaller than AOT. It has a molecular volume of \( \sim 95 \text{ Å}^3 \), compared to the molecular volume of AOT, which is \( \sim 455 \text{ Å}^3 \). So the probe is about a fifth the size of AOT. In addition, its concentration in the bilayers is small, with 1 \( W(CO)_{6}/600 \text{ AOT} \). Therefore, the presence of \( W(CO)_{6} \) should not have a significant influence on the global structure of the surfactant bilayers.

Figure 2 displays FT-IR absorption spectra of the asymmetric CO stretching mode of \( W(CO)_{6} \) in AOT/water mixture with λ.

![Figure 2](image-url)  
*Background subtracted infrared absorbance spectrum of the CO stretching mode of tungsten hexacarbonyl (\( W(CO)_{6} \)) in AOT/water binary mixture with various water to surfactant ratios.*

\( \lambda = [\text{H}_2\text{O}]/[\text{AOT}] \) from 98 to 16 are in the lamellar phase and \( \lambda = 6 \) is in the bicontinuous cubic phase.
the larger red tails. The $\lambda = 6$ (BC phase) spectrum is clearly narrower than the other spectra and its peak is shifted by $\approx 0.5$ cm$^{-1}$ to the blue. The full widths at half-maximum (fwhm) from low to high water content are 11.02, 11.92, 12.27, 12.62, and 12.62 cm$^{-1}$.

$\text{W(CO)}_6$ is completely insoluble in water. Therefore, it resides in the hydrophobic interior of the lamellar and BC structures. Spectra can shed light on the location of the vibrational probe within the lamellae and BC phase. Figure 3A shows the FT-IR absorption spectra of the CO asymmetric stretches of $\text{W(CO)}_6$ in the lamellae, dodecane, and EHS (blue curve) (Alta 1). The $\lambda = 37$ spectrum (red curve) and a two-Gaussian fit (black dashed curve). The two Gaussians that result from the fit are the blue and green curves. The blue Gaussian is virtually identical to the EHS spectrum shown in A. The green curve is slightly shifted and somewhat broader than the dodecane spectrum shown in A.

Figure 3. (A) Spectra of the CO stretching mode of $\text{W(CO)}_6$ in $\lambda = 37$ lamellae (red curve), in dodecane (green curve), and in EHS, the model liquid that is AOT without the sulfonate headgroup (see Figure 1). (B) The $\lambda = 37$ spectrum (red curve) and a two-Gaussian fit (black dashed curve). The two Gaussians that result from the fit are the blue and green curves. The blue Gaussian is virtually identical to the EHS spectrum shown in A. The green curve is slightly shifted and somewhat broader than the dodecane spectrum shown in A.

As can be seen in Figure 3A, $\text{W(CO)}_6$ in dodecane has an exceedingly narrow line width of 4.2 cm$^{-1}$ compared to its width in EHS (14.8 cm$^{-1}$). The line width in EHS is more typical. For example, the CO antisymmetric stretches of $\text{W(CO)}_6$ in CCl$_4$ and in CHCl$_3$ are 10 and 19 cm$^{-1}$, respectively. Long linear hydrocarbons like dodecane tend to have all-trans configurations. The all-trans configuration will produce little variation in the local structure experienced by $\text{W(CO)}_6$ giving rise to the narrow line width. It is not unreasonable to assume that the branched alkane inside the lamellae is less ordered than in dodecane. Therefore, $\text{W(CO)}_6$ in the alkane region of AOT will have a wider the line width then it has in dodecane. The branching of the alkane portion of AOT and differences in packing are likely responsible for the somewhat increased width of Gaussian 1 (green curve) in the fit to the AOT spectrum shown in Figure 3B compared to the $\text{W(CO)}_6$ spectrum in dodecane. In EHS, $\text{W(CO)}_6$ can experience a wider range of environments giving rise to a broader line. In EHS, $\text{W(CO)}_6$ can be in all alkane environments or it could be in contact with or near the ester moieties (see Figure 1). Given the spectrum in dodecane, presumably the blue side of the EHS line corresponds to more alkane-like environments and the on the red side of the line there are more interactions with the esters. It is important to note that $\text{W(CO)}_6$ does not have to come in direct contact with an ester oxygen to have its frequency shifted. As has been shown by comparison between experiments and simulations, frequency shifts that give rise to inhomogeneous broadening of vibrational absorption lines can arise from variations in the local electric field.

The results of the fits in Figure 3B suggest that the $\text{W(CO)}_6$ experiences both alkane and ester environments but with the alkane environments dominating. Based on the molecular volume of EHS (392 Å$^3$) and the volume of the ester portion only (124 Å$^3$), EHS, (see Figure 1B) is very approximately composed of a one-third ester region and a two-thirds alkane region. The spectrum of $\text{W(CO)}_6$ in the pure alkane shows that the absorption is on the blue side of the peak of the spectrum in the lamellae. To fit the spectrum in the lamellae requires the EHS spectrum plus additional absorbance on the blue side of the line, indicating that the $\text{W(CO)}_6$ is exposed more to the ester portion of the AOT than to the ester portion. Given the areas of the Gaussians in the fits in Figure 3B (ratio blue/green = 2.1) and the very approximate partitioning of the EHS into 33%/66% ester/alkane, a very crude estimate suggests that the $\text{W(CO)}_6$ is exposed to 77%/23% alkane/ester in the interior of the lamellae. This is one extreme. However, in EHS, $\text{W(CO)}_6$ will frequently be exposed to ester and alkane portions of the surrounding solvent simultaneously. If we take all of the EHS portion of the fit in Figure 3B (blue curve) to represent some interactions of $\text{W(CO)}_6$ with ester moieties and the green curve to represent interactions only with the alkane portions of AOT, then the ratio is 32%/68% alkane/ester. Here ester means some influence on $\text{W(CO)}_6$ because it is either in contact or near an ester. This is a wide range, but the results indicate that the ensemble of $\text{W(CO)}_6$ molecules is not located strictly in the alkane portion of the bilayers. It may be possible to obtain more detailed information on the $\text{W(CO)}_6$ environments through simulations.

$\text{W(CO)}_6$ is not polar. However, it is polarizable. Its polarizability will produce some attraction to the polar ester region of AOT. Returning to the spectra in Figure 2, the increasing tails to the red with increasing water content may
indicate that the additional water, which penetrates to some extent into the ester regions of the AOT headgroup,\textsuperscript{18} causes W(CO)\textsubscript{6} to partition to a greater extent into the ester region. Conversely, the fact the spectrum in the BC phase (λ = 6) has less spectral density to the red may indicate that in this phase, tungsten hexacarbonyl partitions less into the ester region of the AOT head groups.

3.2. Ultrafast IR Spectroscopy. To illustrate the nature of the data, 2D IR vibrational echo spectra of the asymmetric CO stretch of W(CO)\textsubscript{6} inside a lamellar structure (λ = 24) are shown in Figure 4 at two waiting times (T\textsubscript{w}). The positive going bands (red) arise from the 0-1 vibrational transition. The negative going bands (blue), which are only partially shown below the 0-1 bands, arise from vibrational echo emission at the 1-2 vibrational frequency. These bands are shifted by the vibrational anharmonicity to lower frequency along the ω\textsubscript{m} frequency axis. Changes are clearly visible in the 0-1 spectra. At early time (T\textsubscript{w} = 4 ps, top panel), the spectrum is substantially elongated along the diagonal (black dashed line). The width perpendicular to the diagonal shows there is substantial homogeneous broadening of the absorption line. The elongation along the diagonal at short time arises because the line is inhomogeneously broadened and structural evolution of the system has not caused significant sampling at this short time of the range of inhomogeneous frequencies. At long time (T\textsubscript{w} = 95 ps, bottom panel), the 2D IR spectrum is close to symmetrical, which shows that spectral diffusion is almost complete.

For each sample, 2D IR spectra are taken at many T\textsubscript{w} values, and the change in shape with time is quantified with the CLS method.\textsuperscript{37} The CLS data are presented in Figure 5. The data have important qualitative features. All of the AOT lamellar samples (λ = 98, 75, 37, 24, and 16) have almost identical decay curves. However, the AOT sample in the BC phase (λ = 6, blue curve) displays a significantly slower decay. In contrast, the data for W(CO)\textsubscript{6} in the EHS model liquid (red curve, AOT like structure but without the sulfonate, see Figure 2B) decay much faster than any of the other data. These results show that the dynamics interior to the lamellar structures have little dependence on the number of water molecules between the AOT bilayers. However, when the phase changes from lamellar to BC there is a change in dynamics. In addition, the decay curves show that the dynamics interior to the ordered phases are quite different from the dynamics in EHS liquid, which is a model of AOT without the sulfonate headgroup that causes AOT to form ordered phases in the presence of water.

The qualitative ideas are borne out by the quantitative analysis of the CLS curves that yield the FFCFs for each sample.\textsuperscript{46,47} All of the CLS curves shown in Figure 5 are fit very well by a biexponential function. The difference between the T\textsubscript{w} = 0 value of the CLS and 1 is related to the homogeneous component. Combining the CLS fit with the linear absorption spectrum makes it possible to calculate the full FFCF.\textsuperscript{46,47} The FFCF parameters are given in Table 1. First, consider the lamellar samples, λ = 98 to 16. The slowest component of the decay of the FFCF is τ\textsubscript{2}. All of the τ\textsubscript{2} values for the five lamellar samples are basically the same within experimental error. The average of the values is 147 ps. The faster components, τ\textsubscript{1}, are also all the same within experimental error with an average for the five lamellar samples of 12.6 ps. The corresponding amplitude factors, Δ\textsubscript{1} and Δ\textsubscript{2}, are also all the same within experimental error. Note that the deltas in Table 1 are the standard deviations. The total inhomogeneous line width is the convolution of these two Gaussians with standard deviation (Δ\textsubscript{1} + Δ\textsubscript{2})\textsuperscript{1/2}. The fwhm of the inhomogeneous line width is the total standard deviation multiplied by 2.35.

Table 1 also gives the homogeneous linewidths, Γ, and the corresponding homogeneous dephasing times, T\textsubscript{2}. As with the spectral diffusion dynamics, the homogeneous dephasing is the same for the five lamellar samples (λ = 98 to 16) within experimental error. As discussed above, the homogeneous line width has contributions from pure dephasing, the vibrational lifetime, and the orientational relaxation (see eq 2). The vibrational lifetime (see Table 2) contributes only ~0.07 cm\textsuperscript{-1}.
to the homogeneous line width. Orientational relaxation contributes ~0.9 cm\(^{-1}\). Therefore, the homogeneous line width is dominated by pure dephasing, which results from ultrafast motions that produce a motionally narrowed component of the total absorption line width.

When the phase changes, the dynamics change. In the BC phase (\(\lambda = 6\)), the spectral diffusion is slower. The slow component \(\tau_2 = 178 \pm 4\) ps, in contrast to the average of 147 ps for the lamellar phase samples. The fast component \(\tau_1 = 14.7 \pm 0.4\) ps, in contrast to the average of 12.6 ps for the lamellar phase samples. Therefore, both components of the spectral diffusion are slower in the BC phase.

The water at the surface of AOT lamellae and at the AOT surface of large reverse micelles undergoes orientational relaxation in 18 ps.\(^{18,38,39}\) The water in reverse micelles undergoes slower orientational relaxation as the number of water molecules per AOT decreases.\(^{39}\) For 10 waters per headgroup it is 26 ps, and for 5 waters, it is 30 ps.\(^{39}\) Given these results, it is likely that the water orientational relaxation in the BC phase with \(\lambda = 6\) is somewhat slower than the 18 ps orientational relaxation of water at the surfaces of the lamellae. These water orientational relaxation times are much faster than the slow components of the spectral diffusion in either the lamellar or BC phases, although they are comparable to the fast components. It is unlikely that the water dynamics play a significant role in the spectral diffusion measured inside of the surfactant structures, although it cannot be ruled out for the fast component. Therefore, it is reasonable to assume that the slowing of structural dynamics in the BC phase results from the difference in geometry of the BC phase compared to the lamellar phase rather than from the reduction in the number of water molecule per AOT per se. The organization of the AOT in the BC phase is different than it is in the lamellar phase. The difference in the physical arrangement of the surfactant molecules in the two phases results in a difference in the rates of structural dynamics.

While going from the lamellar phase to the BC phase causes a modest slowing of the structural dynamics, the dynamics in the EHS model liquid (see Figure 1) are substantially different. The CO mode of W(CO)\(_6\) in EHS has \(\tau_2 = 80 \pm 9\) ps and \(\tau_1 = 4.8 \pm 0.3\) ps. These times are almost factors of two and three faster, respectively, than the corresponding times in the ordered phases. In addition, the homogeneous line width in EHS is almost a factor of 2 wider than it is in the ordered phases. EHS is a normal liquid with no long-range ordering in its structure. Given the disorder, W(CO)\(_6\) should be in proximity to both the alkane and ester portions of the EHS molecule. It was argued in connection with the FT-IR spectra in Figures 2 and 3 that the absorption spectra of the asymmetric CO stretch of W(CO)\(_6\) in the AOT/water ordered phases is similar to that in EHS but with more contribution from the alkane-like spectrum in the AOT/water samples.

The spectral diffusion dynamics as well as the pure dephasing dynamics are very different in the ordered lamellar and BC phases than they are in the EHS model liquid (see Table 1) even though the chemical nature of the environments in the ordered and disordered systems are very similar. These results demonstrate that the organization of the interiors of lamellae and BC structures have a substantial influence on the structural dynamics. Going from one ordered phase to another, lamellar to BC, has some effect on dynamics. While in the lamellar phase, the structures are planar, in the BC phase, the dominant structure is more akin to cylinders. This change will affect the packing of the AOT hydrocarbon tails in the interior of the ordered structures, which evidently has an impact on the structural dynamics. However, the change from one ordered structure to another has a small influence on dynamics compared to the effect of order vs disorder.

In both the ordered phases and the disordered liquid, there are two time scales for the spectral diffusion that are more than an order of magnitude different. The FFCF is described in terms of a multieponential model. The time constants, \(\tau_1\) and \(\tau_2\) given in Table 1 should be thought of as time scales for processes rather than time constants for specific processes as there may be a collection of motions, which occur with about the same rates, that give rise to the time constants obtained from the fits. The time scales may reflect motions of the side chains (fast) and motions of the total AOT or EHS molecules (slow), with the disordered EHS liquid having overall faster motions of both types. Regardless of the exact nature of the motions, it is clear that the order imposed by lamellar and BC phases substantially slows the structural dynamics.

Although the amount of water between the lamellae layers does not influence the structural dynamics as reported by the spectral diffusion within experimental error, there is evidence that there may be minor changes in the local environment of W(CO)\(_6\) with water content. Figure 6 shows the results of

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**Table 1. FFCF Parameters for the CO Stretching Mode of Tungsten Hexacarbonyl in AOT Lamellar Phase (\(\lambda = 98\) to 16), in the Bicontinuous Cubic Phase (\(\lambda = 6\)), and in Bis(2-ethylhexyl) Succinate (EHS, AOT Model Liquid)**

<table>
<thead>
<tr>
<th>sample</th>
<th>(T_r (\text{ps}))</th>
<th>(\Gamma (\text{cm}^{-1}))</th>
<th>(\Delta_1 (\text{cm}^{-1}))</th>
<th>(\tau_1 (\text{ps}))</th>
<th>(\Delta_1 (\text{cm}^{-1}))</th>
<th>(\tau_2 (\text{ps}))</th>
<th>(\Delta_2 (\text{cm}^{-1}))</th>
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</thead>
<tbody>
<tr>
<td>(\lambda = 98)</td>
<td>2.5 ± 0.2</td>
<td>4.3 ± 0.4</td>
<td>3.0 ± 0.2</td>
<td>12.0 ± 0.5</td>
<td>2.2 ± 0.4</td>
<td>149 ± 9</td>
<td></td>
</tr>
<tr>
<td>(\lambda = 57)</td>
<td>2.4 ± 0.2</td>
<td>4.4 ± 0.6</td>
<td>3.2 ± 0.4</td>
<td>13.4 ± 0.5</td>
<td>2.2 ± 0.6</td>
<td>154 ± 12</td>
<td></td>
</tr>
<tr>
<td>(\lambda = 37)</td>
<td>2.2 ± 0.4</td>
<td>4.9 ± 0.2</td>
<td>2.9 ± 0.6</td>
<td>12.5 ± 0.3</td>
<td>2.2 ± 0.6</td>
<td>144 ± 5</td>
<td></td>
</tr>
<tr>
<td>(\lambda = 24)</td>
<td>2.1 ± 0.3</td>
<td>5.0 ± 0.4</td>
<td>2.7 ± 0.5</td>
<td>13.4 ± 0.5</td>
<td>2.2 ± 0.4</td>
<td>139 ± 6</td>
<td></td>
</tr>
<tr>
<td>(\lambda = 16)</td>
<td>2.2 ± 0.2</td>
<td>4.9 ± 0.4</td>
<td>2.9 ± 0.3</td>
<td>11.7 ± 0.3</td>
<td>2.1 ± 0.3</td>
<td>149 ± 5</td>
<td></td>
</tr>
<tr>
<td>(\lambda = 6)</td>
<td>2.5 ± 0.2</td>
<td>4.1 ± 0.5</td>
<td>2.8 ± 0.2</td>
<td>14.7 ± 0.4</td>
<td>2.3 ± 0.5</td>
<td>178 ± 4</td>
<td></td>
</tr>
<tr>
<td>EHS</td>
<td>1.3 ± 0.3</td>
<td>8.2 ± 0.4</td>
<td>4.4 ± 0.2</td>
<td>4.8 ± 0.3</td>
<td>2.4 ± 0.4</td>
<td>80 ± 9</td>
<td></td>
</tr>
</tbody>
</table>
In all of the systems, the anisotropy decay is faster than either of the spectral diffusion time constants. It is important to recognize that the anisotropy relaxation correlation function, the second Legendre polynomial correlation function, obtained from the pump–probe experiments is a different correlation function than the FFCCF. Therefore, the resulting time constants cannot be directly compared. As an example, for Gaussian fluctuation anisotropy relaxation that is single exponential, dielectric relaxation, which measures the first Legendre polynomial correlation function, gives a time constant that is three times as long as that which would be measured by a pump–probe experiment on the same system.59,60

### 4. CONCLUDING REMARKS

Surfactant bilayers occur very commonly in systems such as vesicles and cell membranes. Here we have examined the dynamics inside surfactant bilayers, which are important for processes that involve bilayers. Molecules can pass through the walls of vesicles and cell membranes. Such transmission of molecules through a bilayer involves the structural fluctuations of the bilayer interior. Cell membranes contain a wide array of transmembrane proteins. For transmembrane proteins, the bilayer is the “solvent” in which biological processes occur. Chemistry depends to a great extent on the nature of the solvent, which provides dynamical intermolecular interactions that are intimately involved in chemical processes. For cytosolic proteins and enzymes, the water medium matters. MD simulations use sophisticated water models that do a reasonable job of reproducing the mechanical degrees of freedom of water. Even when implicit water models are used, these are tweaked to mimic atomistic water models. The bath degrees of freedom can be included in MD simulations of cytosolic proteins and enzymes because we have an extensive understanding of water dynamics, and water simulations have been tested against experiments, principally ultrafast 2D IR vibrational echo experiments.61–64 The equivalent detailed simulations based on 2D IR experimental information that is available for water51,62 are not available for the interiors of surfactant bilayers. The results presented here are an initial step in increasing our understanding of the structural dynamics of surfactant bilayers.

The results presented above provide the first detailed examination of the fast structural dynamics of the interior of surfactant bilayers in both the lamellar phase and the bicontinuous cubic phase. For the lamellae, multibilayers with 16–98 water molecules per AOT surfactant molecule were studied and it was determined that the thickness of the water layer that separates bilayers does not influence the internal structural dynamics of the bilayers. Sixteen water molecules are sufficient to solvate the AOT headgroup and its sodium counterion.18 As the number of water molecules per AOT is increased, the water layer away from the bilayer/water interface becomes increasingly bulk-like. By 98 water molecules per AOT, there is a substantial bulk-like slab of water between the bilayers. The results show that an increased separation of the bilayers by bulk-like water does not change the internal structural dynamics.

Changing the phase does change the dynamics to a small but measurable extent. A reduction in the number of water molecules per AOT to six causes a phase change to BC. The internal structural dynamics in the BC phase are ~20% slower than in the lamellar phase. However, the dynamics in both the lamellar and BC phases are much slower than those of the
model liquid, bis(2-ethylhexyl) succinate (EHS), which is identical to AOT except that it lacks the charged sulfonate headgroup (see Figure 1). Without the sulfonate head groups interacting with water, EHS does not form ordered structural phases. It is essentially identical chemically to the interior of the AOT bilayers but without the ordered structure. In EHS the fast and slow time scale dynamics are approximately factors of 3 and 2 faster, respectively, than the corresponding dynamics in the AOT ordered phases. Thus the order imposed by the interaction of the charged head groups with water has a very substantial impact on the internal structural dynamics of the lamellar and BC phases.

The results provided here are a test bed for MD simulations of the internal structural dynamics of the AOT bilayers. MD simulations of systems such as water and proteins have been compared to 2D IR vibrational echo experimental determinations of frequency–frequency correlation functions. The time scales of the measured dynamics, tens to hundreds of picoseconds are readily amenable to MD simulations. Reasonable reproduction of the 2D IR results by simulations validates the simulations, which then permits the use of the simulations to associate specific molecular motions with the measured time scales.

The experiments on AOT are a first step in understanding the fast internal structural dynamics of multilayers. The experiments used here on AOT systems are currently being extended to the study of phospholipid multilayer models of cell membranes.

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Notes
The authors declare no competing financial interest.

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